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Article

Dysregulated antibody, natural killer cell and immune mediator profiles in autoimmune thyroid diseases

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Abstract: The pathogenesis of autoimmune thyroid diseases (AITD) is poorly understood and the association between different immune features and the germline variants involved in AITD are yet unclear. We previously observed systemic depletion of IgG core fucosylation and antennary α 1,2 fucosylation in peripheral blood mononuclear cells in AITD, correlated with anti-thyroid peroxidase antibody (TPOAb) levels. Fucose depletion is known to potentiate strong antibody-mediated NK cell activation and enhanced target antigen-expressing cell killing. In autoimmunity, this may translate to autoantibody-mediated immune cell recruitment and attack of

self-antigen expressing normal tissues. Hence, we investigated the crosstalk between immune cell traits, secreted proteins, genetic variants and the glycosylation patterns of serum IgG, in a multi-omic and cross-sectional study of 622 individuals from the TwinsUK cohort, 172 of whom were diagnosed with AITD. We observed associations between two genetic variants (rs505922 and rs687621), AITD status, the secretion of Desmoglein-2 protein, and the profile of two IgG N-glycan traits in AITD, but further studies need to be performed to better understand their crosstalk in AITD. On the other side, enhanced afucosylated IgG was positively associated with activatory CD335-CD314+CD158b+ NK cell subsets. Increased levels of the apoptosis and inflammation markers Caspase-2 and Interleukin-1 α positively associated with AITD. Two genetic variants associated with AITD, rs1521 and rs3094228, were also associated with altered expression of the thyrocyte-expressed ligands known to recognize the NK cell immunoreceptors CD314 and CD158b. Our analyses reveal a combination of heightened Fc-active IgG antibodies, effector cells, cytokines and apoptotic signals in AITD, and AITD genetic variants associated with altered expression of thyrocyte-expressed ligands to NK cell immunoreceptors. Together, TPOAb responses, dysregulated immune features, germline variants associated with immunoactivity profiles, are consistent with a positive autoreactive antibody-dependent NK cell-mediated immune response likely drawn to the thyroid gland in AITD.

Keywords: Multi-omic; autoimmune thyroid diseases (AITD); genetic variants; apoptosis; antibody-dependent cell-mediated cytotoxicity (ADCC); anti-thyroid peroxidase antibody (TPOAb)

1. Introduction

Autoimmune thyroid diseases (AITD) are a class of chronic, organ-specific disorders of the thyroid gland with a high genetic heritability (55-75%) [1–4] affecting approximately 5% of the population and with a gender disparity (i.e., women: 5-15%; men: 1-5%) [5–7]. Pathologically, AITD are characterized by autoantibodies against three main thyroid proteins (thyroid peroxidase (TPO), thyroglobulin (Tg), and the thyroid-stimulating hormone (TSH) receptor (TSH-R)), infiltration of the thyroid gland by immune cells (e.g. lymphocytes, NK cells, monocytes, and macrophages), the formation of germinal centers in the thyroid gland [8] and dysregulated TSH levels [9,10]. However, some studies have failed to observe a significant difference in peripheral blood immune cell composition between AITD patients and healthy individuals [11], while others report significant differences in particular cell types or in immune marker expression [12]. Immune cells, thyroid autoantibodies, and secreted proteins including cytokines may play critical roles in AITD development [13] and in immune responses, including in antibody-dependent cell-mediated cytotoxicity (ADCC) pathways [14,15]. However, the underlying autoimmune signatures associated with AITD remain unclear.

ADCC is triggered via antigen/antibody/Fc receptor complex formation, bringing the effector cell (macrophages, NK cells) and the target cell (expressing the antigen) in close contact. The formation and function of antigen/antibody complexes are modulated by various factors including post-translational modifications of glycans decorating antibodies [16,17]. One example is lack of fucose on the N-linked core glycan of IgG. Afucosylated antibodies have a higher affinity (~100-fold) for the immunoglobulin Fc receptor Fc γ RIIIa (CD16a), expressed on NK cells, macrophages and $\gamma\delta$ T cells, and are shown to confer enhanced ADCC potential *in vitro* and anti-tumor activity *in vivo* [18–21]. This could result in antibodies with more potent Fc-mediated effector functions able to more effectively recruit and activate immune effector cells such as NK cells to kill target antigen-expressing cells [22,23]. IgG core fucose, observed in approximately 95% of IgG in healthy individuals, is considered a “safe switch” that can attenuate potentially harmful antibody-dependent damage against self-antigen-expressing normal tissues [18–21,24]. However, it is possible that these processes may be altered in autoimmune diseases.

We previously studied the glycosylation profiles of total immunoglobulin G (IgG) and of peripheral blood mononuclear cells (PBMC) in patients with AITD [4], as well as the glycosylation of IgG-depleted serum proteins in Hashimoto's thyroiditis (HT) patients [25]. In peripheral blood, we identified both depleted core fucosylation of IgG antibodies and decreased antennary α 1,2 fucosylation of PBMC to be associated with autoantibodies to thyroid peroxidase (TPOAb) and AITD status [4]. We also identified a network of genes, including *FUT8* and *IKZF1* that regulate fucosylation, to be implicated in the development of AITD [4,26]. Based on these findings, we speculated that IgG core fucose deficiencies together with elevated levels of autoantibodies may participate in autoimmune responses in AITD by enhancing effector cell activation and heightened immune and inflammatory signals.

Therefore, here we investigated immune features that may signify dysregulated, and likely heightened immune effector cells, antibodies, and immune mediators in AITD. In this *in silico* study in the blood of 622 subjects from the TwinsUK cohort, of whom 172 have AITD features, we aimed to investigate: 1) the association of different components of antigen/antibody/Fc receptor complexes with AITD; 2) the associations between these different immune components in a cohort of samples from volunteers regardless of disease status, and 3) potential genetic drivers on these components (study design summarized in Fig. 1). Specifically, we examined the association of total serum IgG glycosylation, immune traits, such as immune cell subpopulation frequencies (CSFs; i.e. relative frequencies of circulating immune cell subsets), immune cell surface protein expression levels (SPELs; i.e. the measurement of the cell-surface expression of critical proteins) and secreted proteins, in the peripheral blood of patients with AITD compared with those of healthy volunteers (sample sizes of each study performed are summarized in Table S1).

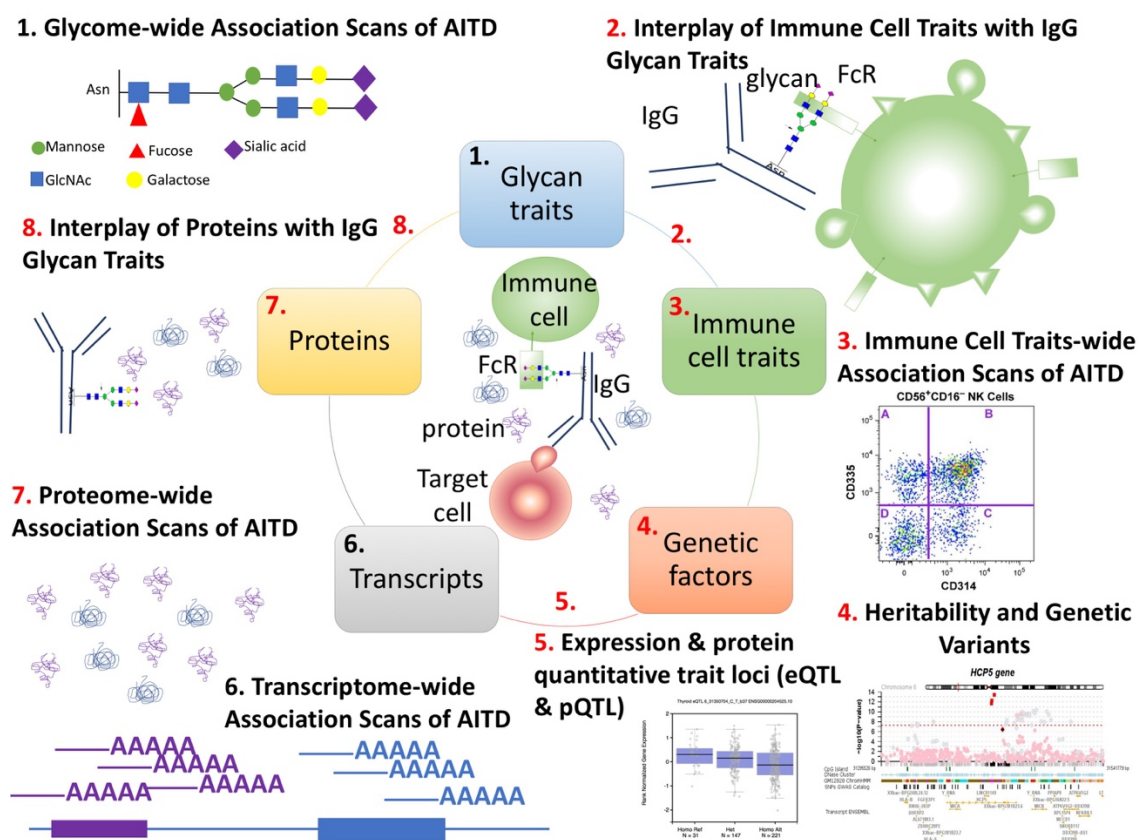


Figure 1. Multi-omics computational analyses were used to study the components of antigen/antibody/effector cell complex structure in AITD. 1) We previously performed glycome-wide association studies of AITD and TPOAb levels using 3,146 individuals from three European cohorts, including the TwinsUK cohort. We identified 17 AITD-IgG N-glycan traits in the discovery TwinsUK cohort, and seven of these 17 have been then replicated in two other cohorts [4]. 2) In the present

study, we studied the association of total IgG N-glycan traits with 23,485 immune cell traits in 383 individuals from the TwinsUK cohort (regardless of disease status). We showed that 6 out of the 17 AITD-IgG glycan traits were correlated with 51 immune cell traits featuring the CD335, CD134, and CD158b receptors. 3) None of these 51 immune cell traits appeared to be associated with AITD in 374 individuals (34 with AITD). 4) The heritability of AITD, TPOAb level and several -omic features (IgG N-glycan traits and immune cell traits) were performed in previous studies of the TwinsUK cohort [4,27–29]. Here we estimated the heritability of secreted proteins, but we could not determine shared additive genetic variance between different phenotypes studied (AITD status, TPOAb level, level of IgG N-glycan traits, of immune cell traits and of circulating proteins in the bloodstream). 5) We identified genetic variants that alter the expression of genes, proteins and cell-bound immune receptors (highlighted in this study) using the previous GWASs performed in the TwinsUK cohort or from GWAS catalog, eQTLs from GTEx project and pQTLs from INTERVAL project [27,28,30–35]. 6) We previously performed transcriptome-wide association studies of AITD, TPOAb level, and N-glycan structures in the whole blood of approximately 300 individuals and we found no significant associations [4]. 7) We observed 3 out of 1,113 circulating proteins tested in plasma of almost 300 individuals shown to be associated with AITD status (TSH, Caspase-2, and Interleukin-1 α). 8) Several secreted proteins were correlated with the level of plasma IgG glycan traits in 164 individuals, but none of them were also associated with AITD. The sample sizes of these different studies are described in **Table S1**. GlcNAc = N-acetylglucosamine. The numbers in black depict analyses performed previously [4,27–29] while the numbers in red depict analyses presented for the first time in the present study.

2. Materials and Methods

2.1. Study Sample

The study was conducted using immune cell traits, glycosylation, proteomics, genotyping, and phenotypes in samples from research volunteers from the UK Adult Twin Registry (TwinsUK cohort). The TwinsUK cohort is comprised of approximately 14,000 monozygotic and dizygotic same-sex adult twins from the UK, unselected for any particular disease or trait (**Table S1**). The cohort is of Northern European/UK ancestry and has been shown to be representative of singleton populations and the UK population in general [36,37]. Ethical approval was granted by the National Research Ethics Service London-Westminster, the St Thomas' Hospital Research Ethics Committee (EC04/015 and 07/H0802/84). Informed consent was obtained from all study participants.

2.2. Data Statement

Multi-omic data were derived from samples in the TwinsUK cohort. Individual-level TwinsUK data, including phenotypes and genotypes, are not permitted to be shared or deposited due to the original consent given at the time of data collection. Access data can be applied for through the TwinsUK data access committee (<http://twinsuk.ac.uk/resources-for-researchers/access-our-data/>).

2.3. Definition of AITD and detection of TSH and TPOAb

The study was performed using a clinical AITD definition and TPOAb as a threshold trait; it was not possible for AITD (Hashimoto's disease and Graves' disease) clinical diagnosis to be confirmed by a clinician. However, approximately 90% of individuals with Hashimoto's disease, about 75% with Graves' disease, <20% with other thyroid diseases, and <10% of normal individuals are known to have TPOAb-positivity [38–40]. Therefore, individuals were considered to have AITD if they either showed significantly higher than normal TPOAb serum titers (set at 3-fold higher than the threshold set by the manufacturer [18 IU/mL for the Abbott assay and 100 IU/mL for the Roche assay]) or had TSH serum levels >10 mIU/L. We considered individuals as controls if they had normal levels of TSH and a negative TPOAb titer, with no previous clinical diagnosis of thyroid disease and who were not treated with thyroid medications or steroids. Individuals with a history of thyroid cancer or thyroid surgery were excluded. Among the 622 individuals studied, 172 (27.65%) were

identified with AITD, 236 (37.94%) considered normal controls, and 214 (34.41%) have TPOAb or TSH serum levels outside the normal range, but do not reach the 3-fold cutoff for inclusion in the AITD cohort. Evaluations of sera to measure TPOAb and TSH levels are described in **Appendix A**.

2.4. Detection of IgG glycosylation profiling for discovery

Plasma specimens for analysis of IgG glycosylation was collected between 1997 and 2013 in 2,279 individuals from the TwinsUK cohort. IgG glycosylation profiling was performed on total plasma IgGs glycome (combined Fc and Fab glycans and all IgG subclasses) in Genos Glycoscience Research Laboratory, Croatia using UPLC analysis of 2AB-labelled glycans. Protocol, data pre-processing and normalization in the TwinsUK cohort were previously described [4] (**Appendix A**).

2.5. Detection of immune cell traits

Plasma samples for assessment of 78,000 immune traits were collected between 2010 and 2012 in 669 female participants from the TwinsUK cohort using high-resolution deep immunophenotyping flow cytometry analysis as previously described [28]. 78,000 different cell surface marker combinations captured by 7 distinct 14-color immunophenotyping panels were detected and described immune cell subset frequencies (CSF) and immune cell-surface protein expression levels (SPELs). After quality control to remove immune cell traits that appeared as poor reproducibility or out of range, 23,485 immune cell traits from 497 individuals of the TwinsUK cohort were analyzed. For this analysis, only 374 twins had immune cell traits data and TPOAb level detected by Roche immunoassay and 245 individuals in a case-control study by combining Roche and Abbott assays (204 controls and 41 AITD). Immune traits were quantile normalized residuals of a linear mixed effect model where age was included as fixed effects, and the batches were considered as random effects.

2.6. Detection of protein profiling in plasma

With an aptamer-based multiplex protein assay (SOMAscan v2, SomaLogic Inc, Boulder, CO) [41,42], 1,129 proteins were measured (2013) on plasma samples collected between 2004 and 2011 from 211 female twins of the TwinsUK cohort (**Appendix A**).

2.7. Statistical analyses

All statistical analyses were run using R version 3.2.3. Linear mixed effect models were conducted using the R lme of package lme4 [43], and linear models were done in using R function lm of package stat. Custom R scripts developed for this study are available at this URL: https://github.com/TiphaineCMartin/multiomic_AITD.git.

For determination of effective number of independent tests for different -omic data, association studies between -omics features and thyroid phenotypes and heritability analysis for proteins (**Appendix A**).

2.8. Genome-wide Association Analysis on IgG N-glycan traits

To define genetic variants (i.e., single nucleotide polymorphisms (SNP), short insertions and deletions (indels)) associated with glycosylation profiles regardless of specific phenotypes in the TwinsUK cohort, we ran analyses with the GenABEL software package [44] designed for genome-wide association study (GWAS) analysis of family-based data by incorporating pairwise kinship matrix calculated using genotyping data in the polygenic model to correct relatedness and hidden population stratification. Data were recently published with other datasets [26,45]. We selected genetic variants for each IgG N-glycan traits with a P-value under the GWAS threshold (P-value < 5×10^{-8}) and added the list of previously-defined genetic variants [29,45] (**Appendix A**).

2.9. Determination of shared genetic variants and genes between IgG N-glycan traits, immune cell traits, protein abundance, and thyroid functions and diseases

To examine whether IgG N-glycan, immune cell traits, proteins, thyroid functions and diseases shared genetic variants or genes, we compared the genetic variants from GWASs on TwinsUK data (NHGRI GWAS catalog and other projects). As genetic variants detected by GWASs could be lead genetic variants but not necessarily causal genetic variants [46], we extended the list of genetic variants to other variants in linkage disequilibrium (LD) with an r^2 threshold of 0.8 from 1000G Phase 1 European population. Using HaploReg V4.1 [47] and GTEx data [32,33], we extracted tissue-specific expression quantitative traits (eQTLs) associated with these genetic variants.

2.10. Visualization

Heatmaps were created in using R package ComplexHeatmap. Correlation plots were created with R package corrplot. Boxplots and scatter plot were created in using R package ggplot2.

3. Results

3.1. Depletion of IgG core fucose is positively associated with increased CD158b+CD314+CD335- NK cell subset counts

IgG N-glycosylation is considered indispensable for the effector functions of IgG and inflammation control [48–52] and plays an essential role in the recognition and binding to Fc receptors of immune cells [51]. Using high-resolution deep immunophenotyping flow cytometry analysis in 669 twins from the TwinsUK cohort and IgG N-Glycan traits in 2,297 twins from the same cohort [4,27,28], we identified 383 samples with measurements of 23,485 immune cell and 17 AITD-IgG N-glycan traits (IGP2, IGP7, IGP8, IGP15, IGP21, IGP33, IGP36, IGP42, IGP45, IGP46, IGP48, IGP56, IGP58, IGP59, IGP60, IGP62 and IGP63) and searched for any associations between them (Table S1).

In our cohort, we identified 1,357 independent immune cell traits among 23,485 potential tested immune cell traits, where the partial correlation between immune cell traits is highlighted in Fig. S1a, 20 independent IgG N-glycan traits among 75 potential tested IgG N-glycan traits, and 6 independent AITD-IgG N-glycan traits among 17 potential tested AITD-IgG N-glycan traits [53]. Association studies of total IgG N-glycan traits with immune cell traits showed that 6 of the 17 identified significant IgG N-glycan traits (IGP2, IGP42, IGP46, IGP58, IGP59, IGP60) previously associated with TPOAb level and AITD status in the TwinsUK cohort, were also associated with 51 immune cell traits, which are all NK cells (CD16+CD56) featuring different combinations of 6 immunoreceptors (CD2, CD158a, CD158b, CD314, CD335, R7) (Table S2, Fig. 2). Three IgG N-glycan traits without core fucose (IGP2, IGP42, IGP46) were negatively associated with the level of the activating subpopulation of CD16+CD56+CD158b-CD335+ NK cells and positively associated with the level of the CD16+CD56+CD335- effector NK cell subpopulation and with the activating subpopulation CD16+CD56+CD158b+CD314+CD335- NK cells [54–59]. In contrast, three other significant IgG N-glycan traits with core fucose (IGP58, IGP59, and IGP60) had the opposite effect associations with the same subpopulations of NK cells (Fig. 2a). In agreement with our previous report, there are therefore negative correlations between the set of IgG N-glycan traits without core fucose (IGP2, IGP42, IGP46) and the set of IgG N-glycan traits describing IgG core fucose (IGP58, IGP59, and IGP60) [4] (highlighted in Fig. 2b). Moreover, we observed strong correlations between these 51 immune cell traits (Fig. 2c). The presence of correlation patterns between the 17 AITD-IgG N-glycan traits (Fig. 2b) as well as between the 51 immune cell traits (Fig. 2c) is consistent with our observation of correlations between the 6 AITD-IgG N-glycan traits and the 51 immune cell traits (Fig. 2a). When we extended our analysis to the 58 remaining IgG N-glycan traits also identified in our samples, but not associated with AITD, we observed no significant association between them and the 23,485 immune cell traits. Moreover, for 23,485 peripheral blood immune cell traits (Table S1), no significant association with AITD or TPOAb level could be identified (Fig. S1b).

We conclude that a subpopulation of NK cells (CD16+CD56) and specifically the activating subpopulation CD16+CD56+CD158b+CD314+CD335- NK cells is associated with fucose-depleted IgG in individuals with AITD.

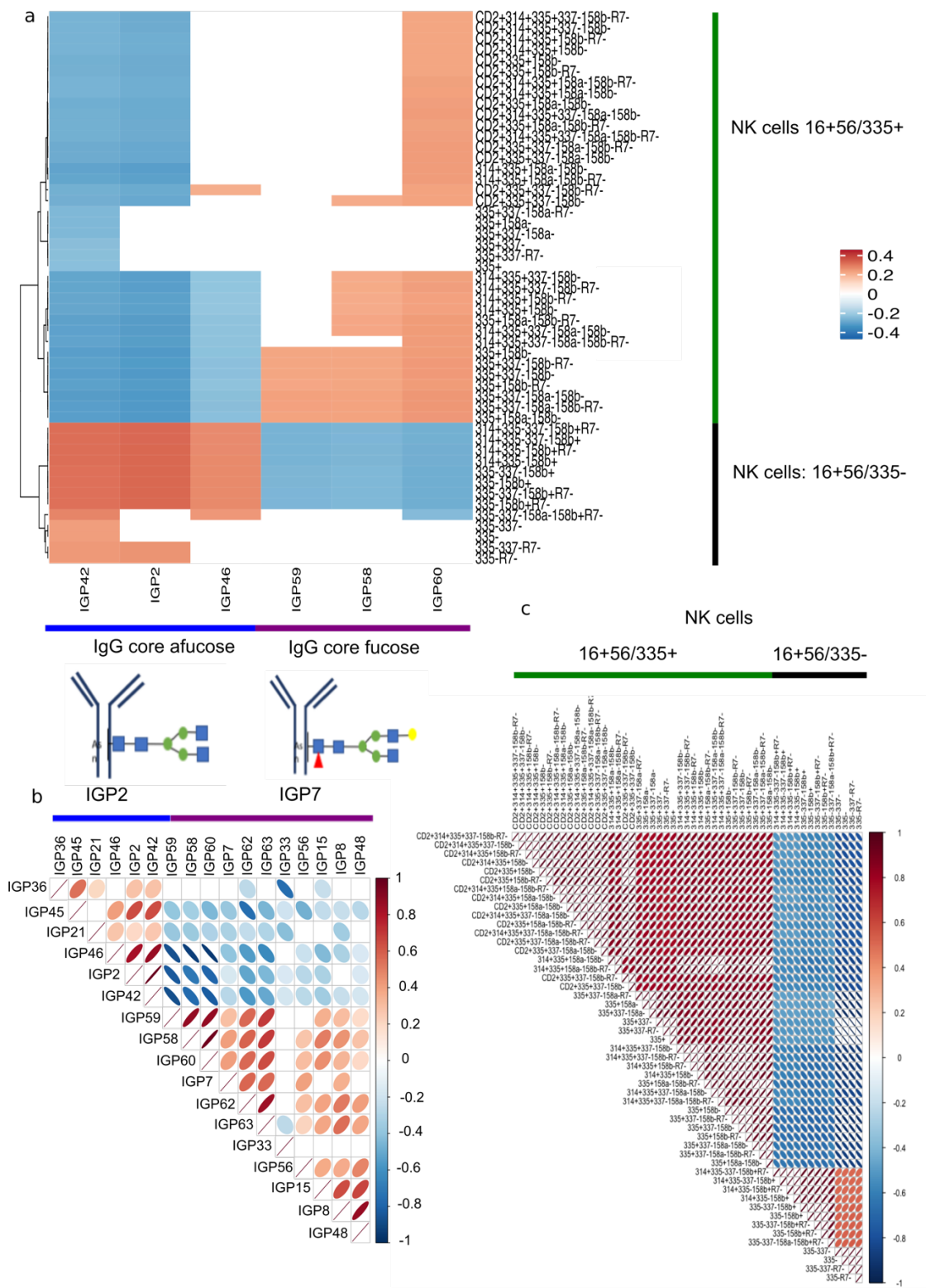


Figure 2. AITD-IgG N-glycan traits associated with a subpopulation of NK cells. (a) Heatmap of immune cell traits associated with AITD-IgG N-glycan traits. The 51 NK cell types were significantly associated with 6 out of 17 AITD-IgG N-glycan traits previously identified [4]. Below the heatmap, there are one representative of IgG core afucose (IGP2) and one representative of IgG core fucose (IGP7), that were both associated with AITD and TPOAb levels [4]. (b) Co-expressions between only 17 IgG N-glycan traits previously associated significantly with AITD status and TPOAb level [4]. (c)

Correlations between the profile of 51 immune cell traits that were associated significantly with at least one of 17 AITD-IgG N-glycan traits. The order of immune cell traits is the same as that in Fig 2a.

3.2. The AITD-associated genetic variants, rs1521 and rs3094228, alter thyroid cell expression of ligands for CD314 and CD158b immunoreceptors

The NK cell receptors, CD335 (NKG2D), CD314 (NKG2D) and the killer cell immunoglobulin-like receptors (KIRs) including CD158b, are normally associated with activated NK cell states, T cell co-stimulation, and mediating tumor cell lysis [55,57]. To determine whether genetic factors could contribute to AITD, related immune features, or their pathways, we inspected genetic variants associated with AITD, TPOAb levels, and immune cell traits from previous GWAS [27,28,30]. We then compared these with recent large-scale studies on tissue-specific expression quantitative traits (eQTLs) [35], mainly from the GTEx project [32–34] in blood and thyroid tissue.

No genetic variants previously associated with AITD or other thyroid phenotypes appeared to be associated with the expression of CD335 or its known ligands in blood and thyroid cells. However, we observed that two genetic variants, rs1521 and rs3094228, associated with Graves' disease and TPOAb-positivity, respectively, fall in the gene regulatory regions of *MIC-A* and *MIC-B* genes, two ligands of CD314 (NKG2D), and alter their gene expressions in thyroid cells [32–34,60–62] (Fig. 3, Table S4, Fig. S2). These two AITD-genetic variants, rs1521 and rs3094228, also alter the expression of the *HLA-C* gene, ligand of CD158b, in thyroid cells. The Graves' disease (GD) risk allele of rs1521 variant is primary associated with a reduced expression of *HLA-C* gene, ligand of CD158b, in thyroid cells. Furthermore, the TPOAb-positivity risk allele of rs3094228 variant is primary associated with an increased expression of *MIC-B* gene, ligand of CD314 (NKG2D), in thyroid cells. As about 75% of patients with Graves' disease have TPOAb-positivity and rs3094228 that has been associated with TPOAb-positivity and Graves' disease [61,63], it is possible that the association of rs1521 with Graves' disease could be also driven by TPOAb-positivity and, so, associated with its phenotypes. Downregulation of *HLA-C* gene expression and upregulation of *MIC-A* and *MIC-B* gene expression in thyrocytes could activate NK cell functions and the cytokine production against thyrocytes when NK cells and thyrocytes are in contact.

Furthermore, three genetic variants, rs2596460, rs2596457 and rs2523691, previously associated with higher levels of the subpopulation of NK cells featuring CD16⁺CD56/CD2-CD314⁺CD335-CD337-CD158a⁺CD158b⁺ [28], are in the same haplotype as the rs3094228 genetic variant, but with a low linkage disequilibrium (LD, $r^2 < 0.8$) (Table S5). Potentially, one of the genetic variants in this locus are the causal genetic variant of higher abundance of CD158b⁺CD314⁺CD335- NK cells. All of the three genetic variants could also alter the expression of the *HLA-C* gene, ligand of CD158b, and *MIC-A*, ligand of CD314 (NKG2D), in immune cells [32–34,64] (Fig. 3, Table S4).

Overall, two genetic variants, rs1521 and rs3094228, associated with, respectively, Graves' disease and TPOAb-positivity, appear to alter thyrocyte expression of ligands of two immunoreceptors of NK cells, CD314 and CD158b; both of which have the capacity to enhance cytotoxicity of NK cells after binding with target cells. Additionally, three genetic variants in the same haplotype than rs3094228 could increase the abundance of the immune active CD158b⁺CD314⁺CD335- NK cell subpopulation.

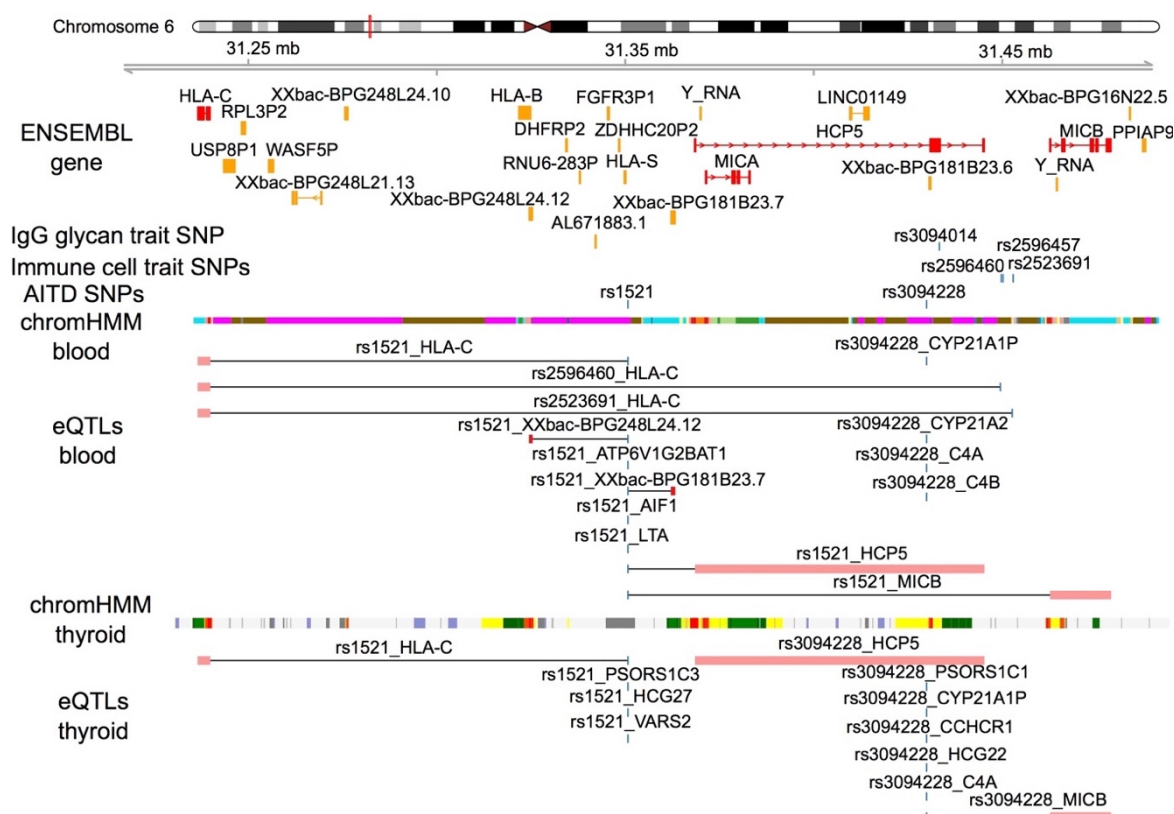


Figure 3. Association of immune cell traits with AITD status. Annotation tracks around *MIC-A*, *MIC-B* and *HLA-C* genes visualize significant GWAS hits for immune cell traits, the ligands of certain immunoreceptors (such as NK), and thyroid phenotypes previously identified in the TwinsUK cohort as well as chromatin states identified using chromHMM from whole blood from ENCODE [65] and thyroid cells from CEMT [66] and eQTLs from GTEx project [32,33]. The plot was produced using functions from R packages Gviz and coMET [67].

3.3. AITD is associated with increased serum Caspase-2 and IL-1 α

We next evaluated whether the abundance of 1,113 free soluble proteins, which are partially correlated between each other (**Fig. 4a**), in peripheral blood may be associated with AITD status (27 AITD patients versus 130 healthy controls) and TPOAb levels (155 individuals of whom 25 have AITD) in the TwinsUK cohort (**Table S1**) using aptamer-based multiplex protein assay (SOMAscan) [68]. Firstly, we observed significant moderate correlations of the TSH levels measured by two clinical-certificated assays (Abbott and Roche) with the TSH levels measured by the SOMAscan assay (**Fig. 4b**). This indicated that the SOMAscan assay could reproduce with good accuracy the estimation of TSH levels and probably also for other proteins. Levels of three proteins were positively associated with AITD status (Bonferroni multiple testing correction, P -value $< 1.9 \times 10^{-4}$): TSH (P -value $= 8.67 \times 10^{-5}$; Beta $= 0.67$; SE $= 0.16$), Caspase-2 (CASP-2; P -value $= 2.72 \times 10^{-7}$; Beta $= 1.10$; SE $= 0.20$) and Interleukin-1 α (IL-1 α ; P -value $= 7.46 \times 10^{-5}$; Beta $= 0.41$; SE $= 0.09$). We also observed higher mean levels of TSH in patients with AITD (mean_{SOMAscan} $= 1443.9$, sd_{SOMAscan} $= 1238.5$; mean_{clinical} $= 7.1$ IU/mL, sd_{clinical} $= 10.47$) or TPOAb-positivity (mean_{SOMAscan} $= 1389.9$, sd_{SOMAscan} $= 1018.3$; mean_{clinical} $= 5.7$ IU/mL, sd_{clinical} $= 7.55$) compared with controls (euthyroidism with TPOAb-negative) (mean_{SOMAscan} $= 851.1$, sd_{SOMAscan} $= 368.6$; mean_{clinical} $= 1.64$ IU/mL, sd_{clinical} $= 0.79$). Although Caspase-2 and IL-1 α levels were associated with AITD status, Caspase-2 and IL-1 α levels were not associated with TPOAb or TSH levels as continuous variables (P -value $> 1.9 \times 10^{-4}$). However, when participants were divided into 4 categories according to TSH and TPOAb levels (**Fig. 4c**), reflecting different clinical categories (hyperthyroidism, euthyroidism/TPOAb-negative, hypothyroidism and euthyroidism/TPOAb-positive), Caspase-2 showed significantly higher mean and variance in two groups: hypothyroidism and euthyroidism/TPOAb-positive (**Fig. 4d**). The hypothyroidism and euthyroidism/TPOAb-positivity in

339 this cohort potentially indicate underlying Hashimoto's thyroiditis (HT). This is because HT is the
340 most common cause of hypothyroidism, spontaneous hypothyroidism (i.e. no previous history of
341 thyroid ablation) is almost always caused by HT, and euthyroid individuals with TPOAb-positivity
342 almost always have HT when studied by cytology and histopathology [69–71]. On the other hand,
343 the variance of IL-1 α was significantly larger in groups with euthyroidism/TPOAb-positive and
344 hyperthyroidism (**Fig. 4e**), but there was no significant difference for their mean values. Hence,
345 individuals from 4 categories have the same levels of IL-1 α , but there are more inter-individual
346 variabilities in euthyroidism/TPOAb-positive and hyperthyroidism than euthyroidism/TPOAb-
347 negative and hypothyroidism.

348 In summary, we confirmed the association of the plasma TSH levels with AITD status, and
349 we found two novel associations of plasma Caspase-2 and IL-1 α with AITD status, but their secretion
350 (mean and variance) seems to also depend on other factors associated with thyroid diseases such as
351 the levels of TSH and TPOAb.

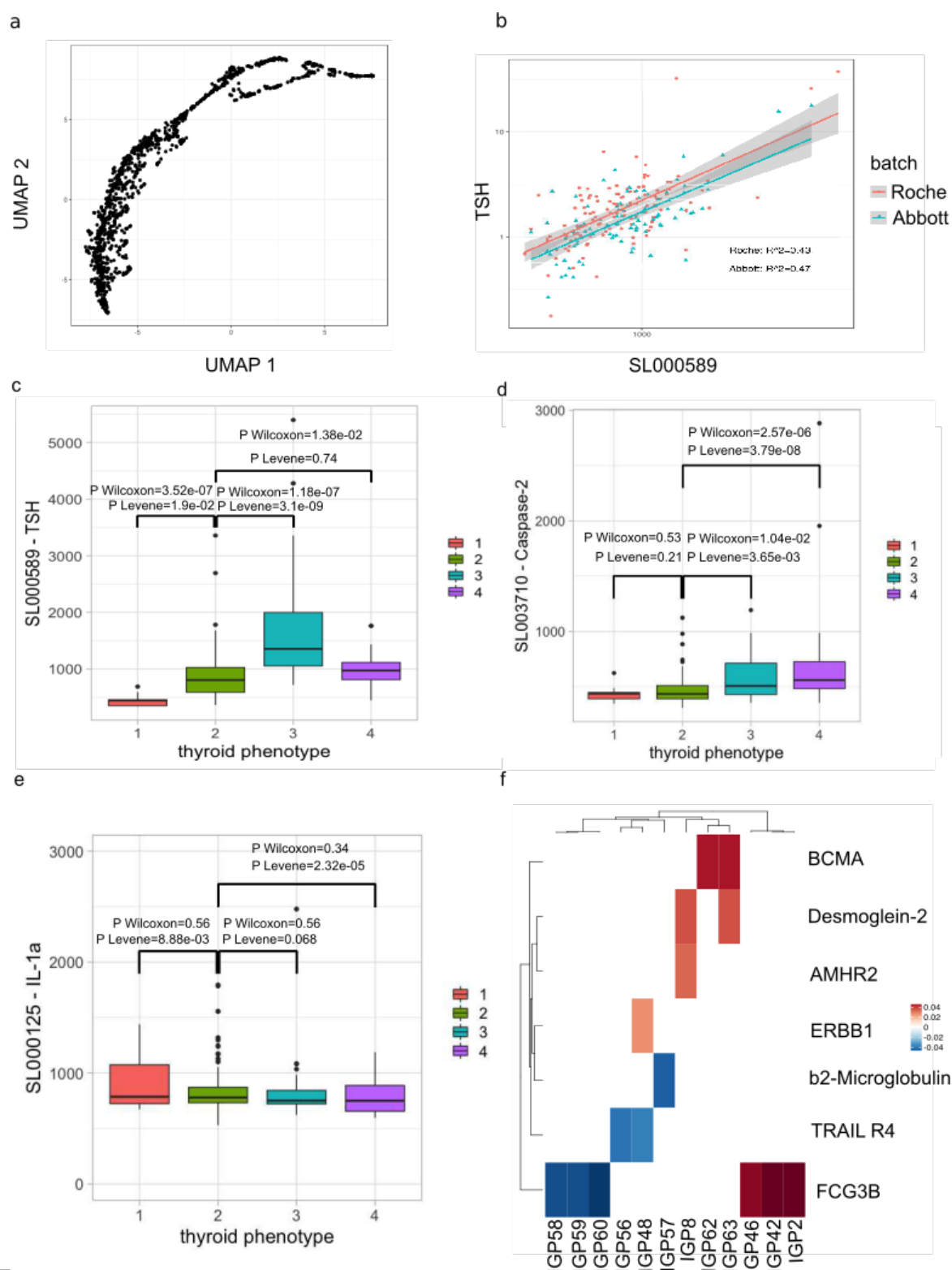


Figure 4. Association of circulating protein abundances with thyroid diseases and with AITD-IgG N-glycan structures. (a) 1,113 circulating proteins were arranged in two dimensions based on the similarity of their secretion profiles in the serum by the dimensionality reduction technique UMAP [72] using R package umapr [73]. (b) Correlation of log₁₀-transformed TSH measurements between two clinical FDA approved clinical immunoassays (Roche and Abbott) and SOMAscan assay in 217 individuals (122 using Roche immunoassay and 95 using Abbott immunoassay). (c) Box plot of the level of circulating TSH measured by SOMAscan assay in the serum according to the group of thyroid status. (d) Box plot of the level of circulating Caspase-2 measured by SOMAscan assay in the serum according to the group of TSH. (e) Box plot of the level of circulating IL-1 α measured by SOMAscan

assay. An extreme outlier sample in the group 4 with an IL-1 α of 250,000mg/ml was discarded for the analysis. (f) Heatmap of circulating protein abundances associated with AITD-IgG N-glycan structures. In fig.2c-e, participants were assigned to 4 categories according to TSH level and TPOAb status: 1=hyperthyroidism (TSH \leq 0.1 mIU/L; 13 individuals), 2=euthyroidism/TPOAb-negative (0.4<TSH \leq 4 mIU/L & TPOAb < 6 IU/mL (Abbott) or TPOAb < 34 IU/mL (Roche); 196 healthy individuals), 3=hypothyroidism (TSH \geq 4 mIU/L; 21 individuals), and 4=euthyroidism/TPOAb-positive (0.4<TSH \leq 4 mIU/L & TPOAb \geq 6 IU/mL (Abbott) or TPOAb \geq 34 IU/mL (Roche); 28 individuals). Wilcoxon-Mann-Whitney's test has been performed between groups to estimate whether there are mean differences whereas Levene's test has been performed between groups to estimate whether there are variance differences.

3.4. Afucosylated IgG is associated with serum levels of several circulating proteins

When we studied the correlation between the level of secreted TSH, Caspase-2 and IL-1 α proteins and IgG N-glycan trait levels in 164 individuals of whom 27 have AITD, we found no significant associations (P-value $>8.3\times10^{-4}$, Bonferroni test considering 3 independent proteins and 20 independent IgG N-glycan traits) (Table S6, Fig. 4f). However, several AITD-IgG N-glycan traits appeared to be associated with 7 other circulating proteins (AMHR2, BCMA, β 2-microglobulin, ERBB1, Desmoglein-2, TRAILR4, and FCGR3B) (P-value $<3.67\times10^{-5}$, Bonferroni test in considering only 227 independent proteins and 6 independent IgG N-glycan traits) (Table S6, Fig. 4f). For example, three AITD-IgG N-glycan traits (IGP2, IGP42, and IGP46) were positively associated with circulating FCGR3B (Fc γ RIIIb or CD16b), an Fc receptor expressed by polymorphonuclear neutrophils (PMN), whereas three AITD-IgG N-glycan traits (IGP58, IGP59, and IGP60) were negatively associated with the antibody Fc receptor FCGR3B. Also, IGP56 and IGP48 were negatively associated with β 2-microglobulin, involved in the presentation of intracellular antigens through the MHC class I complex; and IGP48 was positively associated with ERBB1, the epidermal growth factor receptor (EGFR), a checkpoint molecule associated with cellular proliferation and differentiation.

Overall, 12 AITD-IgG N-glycan traits (IGP2, IGP8, IGP42, IGP46, IGP48, IGP56, IGP57, IGP58, IGP59, IGP60, IGP62, and IGP63) were associated with serum levels of 7 circulating proteins (AMHR2, BCMA, β 2-microglobulin, ERBB1, Desmoglein-2, TRAILR4, and FCGR3B) in the TwinsUK cohort.

3.5. Free-soluble plasma Desmoglein-2 protein is associated with AITD genetic variants and two AITD-IgG N-glycan traits

We evaluated several GWAS on secreted proteins (protein quantification locus traits, pQTL) [31], to determine whether the secretion of proteins associated with AITD or with AITD-IgG N-glycan traits are driven by AITD genetic variants. We found no genetic variants associated with any of 17 AITD-IgG N-glycan structures that are also pQTL. However, four genetic variants associated with thyroid phenotypes published in the GWAS catalog (rs3761959, rs7528684, rs505922, and rs3184504) were also associated in *cis* and *trans* with nine circulating protein abundances (BGAT, CHSTB, DC-SIGN, Desmoglein-2, DYR, FCRL3, GP1BA, MBL, and VCAM-1) (Table S7). None of these proteins were associated directly with AITD or TPOAb levels in our study. However, we found that Desmoglein-2 was associated with two AITD-IgG N-glycan traits, IGP8, and IGP63 [4] (Fig. S3). Desmoglein-2 is highly expressed in epithelial cells including thyrocytes and cardiomyocytes and plays a role in the cell-cell junctions between epithelial, myocardial, and certain other cell types and is thought to be a regulator of apoptosis [74].

Therefore, four genetic variants associated with thyroid phenotypes are also associated with nine secreted protein abundances, including the apoptosis regulator Desmoglein-2 in blood. Desmoglein-2 was also associated with two AITD-IgG N-glycan traits.

4. Discussion

The dysregulation of the immune system may affect several biological structures and processes in AITD, such as antigen/antibody/Fc receptor complex formation, possibly driven by

genetic and environmental factors [75]. Little is known about the key players and the genetic variants identified in previous GWASs of patients with AITD. Targeting of self-antigen expressing tissues by immune cells may depend on the formation of antigen/antibody/Fc receptor complexes featuring substantial affinity or avidity properties. In the peripheral blood of individuals with AITD, we previously detected depletion in IgG core fucose that is known to enhance such interactions and may influence immune effector cell engagement of target cells by antibodies. We proposed that this signature is associated with TPOAb levels and with immune effector cell activation in patients with AITD [4]. Here, we reveal immune and genetic features pointing to activated NK cell subsets, thyroid cell-derived ligands for immunoreceptors on NK cells, alongside secreted mediators of apoptosis and immune activation, all signals of heightened antibody and innate effector cell responses in AITD.

We applied an *in silico* multi-omic approach on peripheral blood specimens from individuals from the TwinsUK cohort to investigate any association between immune features and genetic variants in AITD. In AITD patient samples, we observed increased levels of three circulating proteins (TSH, Caspase-2, and Interleukin-1 α) and a decreased level of IgG core fucosylation associated with an activated subpopulation of NK cells defined primarily by the expression of CD335, CD134, and CD158b receptors. Our data confirms the previously reported association of plasma TSH level with AITD status and also reveals previously unknown potential biomarkers for AITD, which are highly associated with immunological activation functions, such as ADCC, apoptosis and pro-inflammatory pathways. Furthermore, several genetic variants previously associated with AITD appear to alter thyrocyte gene expression of several ligands of NK immunoreceptors and abundance of plasma circulating proteins. This suggest that the genetic background may also play potential roles in NK cell activation likely focused on thyroid cells in individuals with AITD.

To our knowledge, no other cohorts have large datasets that are available to interrogate and feature the same diversity of *-omics* data with AITD phenotype or TPOAb levels. In our studies, we note an imbalance in the sample sizes between control individual groups and AITD groups. This is because the dataset comes from unselected twins and reflects the general European population [37], where approximately 5% of the population, but 5-15% for women, present individuals with AITD [5–7]. To overcome such imbalances in our sample sizes and low samples sizes with large *-omics* data, we applied machine learning and non-parametric methods with correction for multiple testing. Another limitation in our present study is the absence of AITD clinical diagnosis confirmed by clinicians for all individuals. We consequently applied more stringent criteria to define patients with AITD versus control individuals, by using TSH and TPOAb levels (see Section 2.3 of our Materials & Methods). We also performed analysis on TPOAb levels, as this is considered the main clinical quantitative biomarker of AITD status [38–40]. Replication and meta-analysis studies on larger *-omic* datasets incorporating clinical features will help to confirm our present findings.

Two secreted proteins (Caspase-2 and IL-1 α), which play roles in apoptosis and the inflammatory response, were positively associated with AITD. TPOAb have been proposed to target thyroid cells by engaging effector cells via their Fc receptors [4,14,15,76,77], and the apoptosis protein Caspase-2 may represent a marker potentially signifying antibody-mediated destruction of thyroid cells [78]. In concordance, IL-1 α , produced by activated immune, epithelial and endothelial cells in response to cell injury and apoptosis, is considered an apoptosis index of the target cell [79] and proportional to the degree of lymphoid infiltration in thyroid disorders [80]. IL-1 α seems to reduce the thyroid epithelial barrier, even in the absence of any other signs of cytotoxicity [81]. In concordance, in our study we found higher levels of secreted IL-1 α in AITD blood compared with levels in healthy individuals, and its variance was greater in euthyroidism/TPOAb-positive blood and in hyperthyroidism. This may signify dysregulation in cellular structures in the thyroid gland. Overall, Caspase-2 and IL-1 α may reflect the degree of thyroid cell death or apoptosis and of lymphoid infiltration towards the thyroid gland.

A subpopulation of NK cells expressing combinations of immunoreceptors (CD2, CD158a, CD158b, CD314, CD335, R7) was associated with the depletion of IgG core fucose in individuals with AITD. These included an activating NK receptor (CD314) and a differentiation receptor (CD335); whilst, fucose-depleted IgG was also positively associated with a subpopulation of NK cells with an

inhibitory NK receptor (CD158b) [54–59,82]. The combination of potentially autoreactive antibodies with enhanced Fc domains and activated effector cells such as NK cells may signal increased inflammation and susceptibility to autoimmune disease [83]. Previous studies showed that afucosylated antibodies have a much higher affinity (100-fold) for FcγRIIIa (CD16a) and may thus have enhanced ADCC [84]. Moreover, ADCC via FcγRIIIa may require NK cells, but not monocytes or polymorphonuclear cells, and activity levels of the antigen/antibody/effector cell complexes have been correlated only with the NK cell numbers present in the PBMC [20]. Our associations between the levels of IgG core fucose and of a subpopulation of NK cells reinforce the notion that there is a complementarity between IgG core fucose levels and NK cells, that could influence effector cell potency, potentially against a range of antigens including self-antigens.

It has been previously estimated that AITD are highly heritable (55–75%) and that most of IgG N-glycan traits and the immune cell traits associated with AITD are moderately heritable (**Table S8**) [4,27]. By estimating the proportion of genetic and environmental variance of 1,129 proteins in our study using the Structural Equation Modeling and twin structures present in the TwinsUK cohort (**Table S9**), we found a small proportion of proteins having additive genetic variances in their heritability, in concordance with previous findings on a smaller dataset [85]. As the best model of heritability in AITD is only with dominant genetic variance, the shared genetic variance between AITD and proteins as well as with IgG N-glycan traits and immune cell traits could not be estimated with accuracy. However, in our study, we identified several genetic variants previously associated with thyroid phenotypes to be also associated with the secretion of proteins and gene expression of ligands of two NK cell immunoreceptors. Specifically, genetic variants, rs1521 and rs3094228, associated with Graves' disease and TPOAb-positivity, alter the expression of thyroid cell-expressed ligands, *MIC-A*, *MIC-B*, and *HLA-C*, known to recognize CD314 and CD158b immunoreceptors expressed on NK cells. Moreover, rs3094228 falls in the same European haplotype as three genetic variants associated with higher abundance of the activated CD158b⁺CD314⁺CD335⁺ NK cell subset. Thus, individuals having the AITD-risk allele of rs1521 variant have reduced expression of *HLA-C* gene and, at a lesser extent, expression of *MIC-A* in thyrocytes, whereas the carriers of AITD-risk allele of rs3094228 genetic variant associated with TPOAb-positivity showed increased expression of *MIC-B* gene in thyrocytes and potentially higher abundance of the highly active CD158b⁺CD314⁺CD335⁺ NK cells. Consequently, if the thyrocytes in carriers of AITD-risk alleles for CD158b and CD314 ligands crosstalk with the subpopulation of NK cells with CD158b and CD314 immunoreceptors with the help of the antibodies, they could trigger the production of cytokines and cytotoxicity against thyrocytes by these NK cells.

Our findings thus highlight different immune features (glycan structures on antibodies, a subpopulation of immunoactive NK cells, the secretion of Caspase-2 and IL-1α) as potential signals of AITD status detectable in the bloodstream in addition to TSH and TPOAb levels. Moreover, if one speculates that active antibodies with low core-fucose might be thyroid autoantibodies (e.g., TPOAb) [86] and target cells are thyroid cells, it is conceivable (**Fig. 5**) that that immune cell-antibody-target cell interactions may lead to cytotoxicity functions targeting thyroid tissues [76,86,87]. Together, these may form part of a dysregulated autoimmune response in AITD. Further replication studies and validation studies of real-time functional evaluations associated with these immune features and genetic analyses are needed to confirm this model. These features could also be tested in the context of thyroid cancer immunotherapy [77] in future studies.

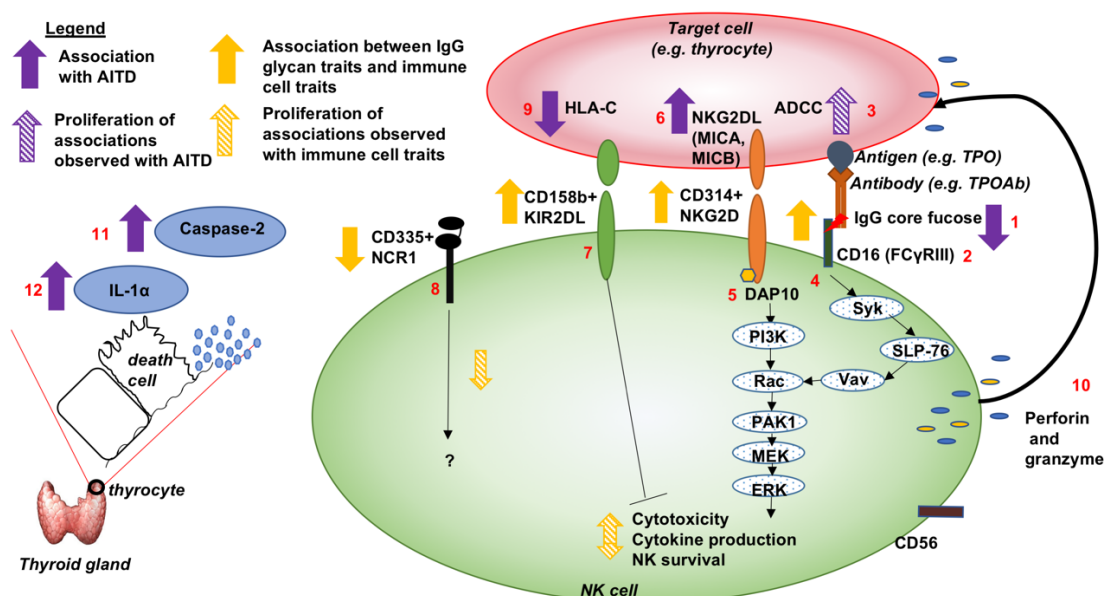


Figure 5. Model of different potential contributing players and their pathways activated in proposed antibody-dependent NK cell-mediated cytotoxicity in the thyroid gland of AITD patients. 1) The depletion of IgG core fucose was associated with TPOAb level and AITD status [4]. 2) The IgG N-glycan traits associated with AITD were also associated with a subpopulation of NK cells in our current study; for example, the depletion of IgG core fucose is associated positively with NK cells with the patterns of co-receptors CD335- or CD335-CD158b+CD314+. 3) Previous studies showed that afucosylated antibodies had increased affinity for binding to CD16 (FcγRIIIa), cell receptors of NK cells, and to enhance ADCC [18–21] via 4) protein tyrosine kinase-dependent pathways, through crosstalk with 5) NKG2D receptor (CD314) [88,89]. 6) Two SNPs, rs3094228 and rs1521, were associated with GD and TPOAb-positivity [60–62] and fall in gene regulatory regions of the *MIC-A* and *MIC-B* genes and increase their expression in thyroid cells [32]. These two genes encode heavily glycosylated proteins that are ligands for the NKG2D type II receptor (CD314). 7) The KIR2DL (CD158b) receptor is known to regulate the cytotoxicity of NK cells by unknown pathways, whereas 8) the NCR1 (CD335) receptor can contribute to the increased potency of activated NK cells to mediate cell lysis by unknown pathway [54,55]. 9) The SNP, rs1521 associated with GD[60], is also shown to reduce the expression of HLA-C gene, producing the ligand of CD158b, in thyroid cells [32,33,58,59]. 10) All together (the binding of NK cells with target cells through antibodies and their ligands), these lead to the activation of NK cells, which release cytotoxic granules containing perforin and granzymes. This release mediates ADCC of target cells (3), which are thyrocytes in AITD. Also, 11) a positive association between the circulation abundance of Caspase-2 protein and AITD were found in this study that could be associated with the destruction of thyrocytes. 12) A positive correlation of circulating abundance of IL-1α with AITD was also found in the bloodstream that could be a marker of lymphocyte infiltration in the thyroid gland of individuals with AITD, and thus of inflammation [80,81].

Supplementary Materials: The following are available online at www.mdpi.com/xxx/s1,

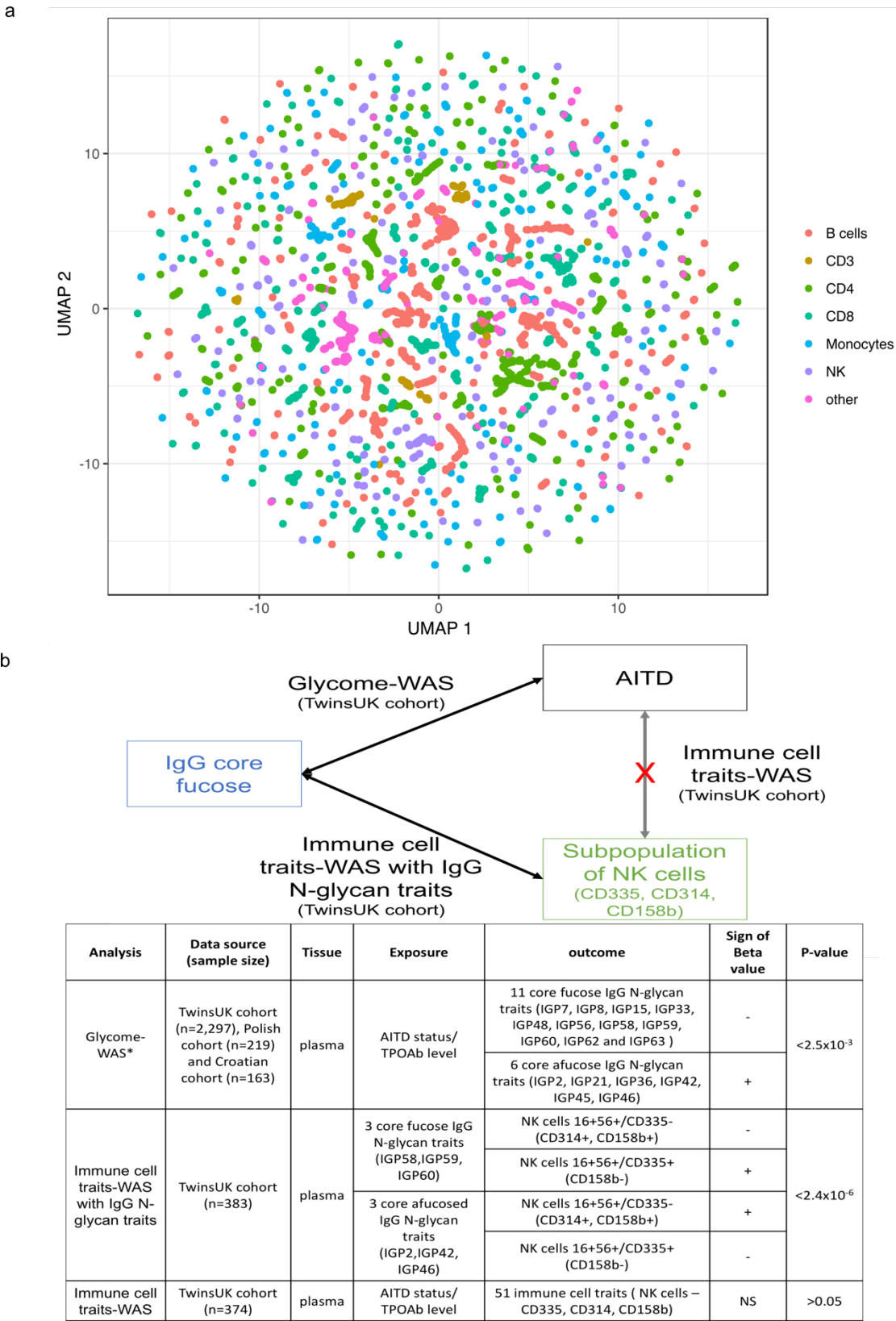


Figure S1: Immune cell traits and AITD status. (a) Immune cell traits were arranged in two dimensions based on the similarity of their quantification profiles by the dimensionality reduction technique UMAP [72] using R package umap [73]. Some clusters that emerge spontaneously can be associated with specific immune cell types (colors). (b) Overview of associations observed between IgG core-fucose, a subpopulation of NK cells and AITD status in the TwinsUK cohort. *Glycome-wide association studies of AITD and TPOAb levels were previously performed [4].

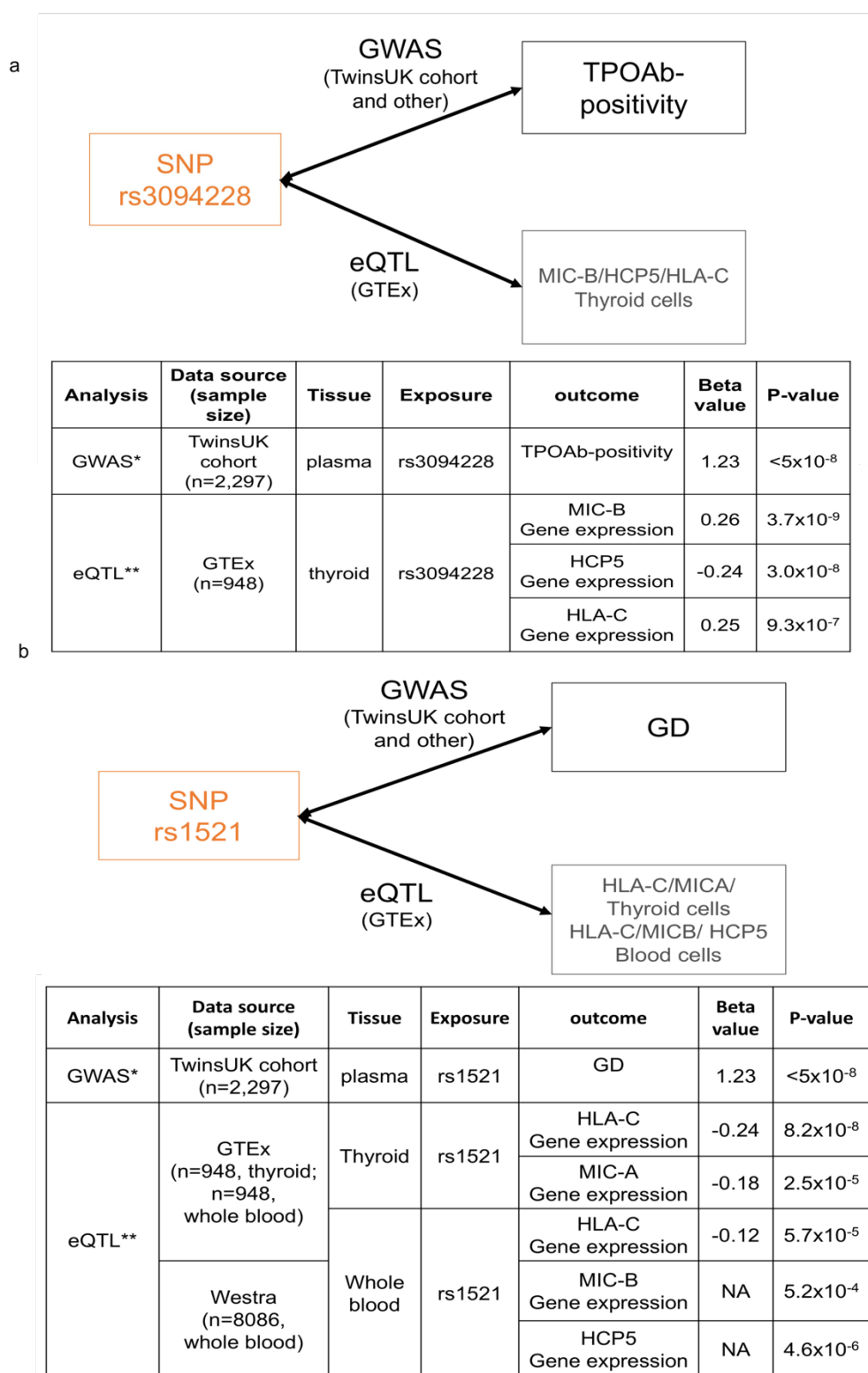
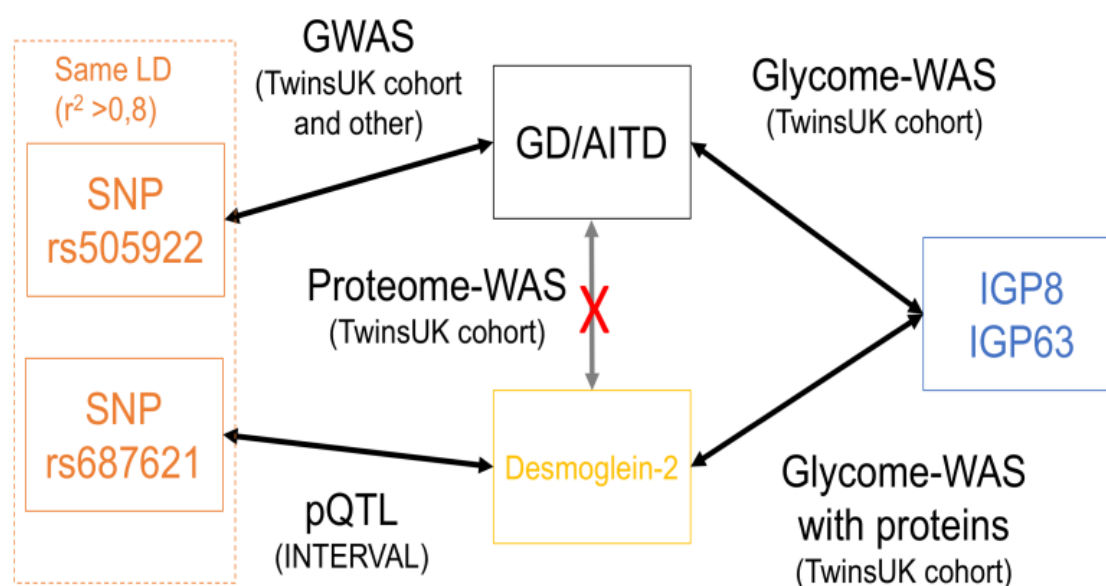


Figure S2. Overview of associations between AITD-SNP and eQTL in thyroid and blood cells. *Genome-wide association studies of AITD and TPOAb-positivity were previously performed, and the findings are available via GWAS catalog [30] whereas **eQTLs come from GTEx project [34] and Westra and al. [35]. (a) Associations between AITD-SNP and eQTL in thyroid and blood cells for the genetic variant rs3094228. (b) Associations between AITD-SNP and eQTL in thyroid and blood cells for the genetic variant rs1521.



Analysis	Data source (sample size)	Tissue	Exposure	outcome	Beta value/ OR	P-value
GWAS*	TwinsUK cohort (n=2,297)	plasma	rs505922	GD	1.13	<2.5x10 ⁻³
Protein-quantitative trait loci**	INTERVAL (n=3,301)	plasma	rs687621	Desmoglein-2	0.20	1.9x10 ⁻¹¹
Protein-WAS	TwinsUK cohort (n= up to 348)	plasma	AITD status/ TPOAb level	Desmoglein-2	NS	>1.9x10 ⁻⁴
Glycome-WAS***	TwinsUK cohort (n=2,297), Polish cohort (n=219) and Croatian cohort (n=163)	plasma	AITD status/ TPOAb level	IGP8	-6.93	2.1x10 ⁻³
				IGP63	-7.23	1.2x10 ⁻³
Protein-WAS	TwinsUK cohort (n= 164)	plasma	IGP8	Desmoglein-2	0.032	2.2x10 ⁻⁶
			IGP63	Desmoglein-2	0.032	8.0x10 ⁻⁶

545

546 Figure S3. Overview of multi-omic findings associated with Desmoglein-2 in individuals with AITD status and
 547 general population. We highlighted a locus with high LD having SNPs and two IgG glycan traits that are both
 548 associated with GD and the abundance of secreted plasma Desmoglein-2 in plasma. However, no direct
 549 association of AITD status with the abundance of secreted plasma Desmoglein-2. We previously performed
 550 glycome-wide association studies of AITD and TPOAb levels [4]. Genome-wide association studies of AITD and
 551 TPOAb-positivity were previously performed, and the findings are available via GWAS catalog [30] whereas
 552 pQTLs come from INTERVAL project[31]. IGP8 = the percentage of FA2[3]G1 glycan in total IgG glycans. IGP63
 553 = The percentage of fucosylation (without bisecting GlcNAc) of agalactosylated structures.

554 Table S1: Description of TwinsUK cohort used for different analysis performed here

555 Table S2: Significant glycome associations with immune cell traits in the TwinsUK cohort

556 Table S3: Associations of 51 immune cell traits with AITD and TPOAb level in the TwinsUK cohort

557 Table S4: Hits from selected eQTL studies for two SNPs, rs1521 and rs3094228 in the thyroid cells and whole
 558 blood

559 Table S5: Genes reported for genetic variants associated with thyroid phenotypes and immune cell traits

Table S6: Glycome-wide associations studies of 17 AITD-IgG N-glycan traits with 1,113 circulating proteins. Only significant ones were put here.

Table S7: Genetic variants associated with thyroid phenotypes and AITD-IgG N-glycan traits overlapping pQTL identified in INTERVAL project (LD $r^2 > 0.8$)

Table S8: Heritability of AITD, 17 IgG N-glycan traits and 51 immune cell traits in the TwinsUK cohort

Table S9: Heritability of 1,113 proteins in the TwinsUK cohort

Abbreviations. ADCC (Antibody-Dependent Cell-mediated Cytotoxicity), AITD (AutoImmune Thyroid Diseases), CDC (Complement-Dependent Cytotoxicity), CD158b (KIR2DL2/L3 - Killer Cell Immunoglobulin Like Receptor, Two Ig Domains And Long Cytoplasmic Tail 2 or 3 - or NKAT6 - Natural Killer-Associated Transcript 6), CD314 (NKG2D or KLRK1 - Killer Cell Lectin Like Receptor K1), eQTL (gene Expression Quantitative Trait Loci), FcγR (Fc gamma Receptor), GD (Graves' Diseases), GlcNAc (N-acetylglucosamine), GWAS (Genome-wide Association Study), HT (Hashimoto's thyroiditis), IgG (Immunoglobulin G), MIC-A (MHC Class I Polypeptide-Related Sequence A), MIC-B (MHC Class I Polypeptide-Related Sequence B), NK (Natural Killer cell), PBMC (Peripheral Blood Mononuclear Cells), pQTL (Protein expression Quantitative Trait Loci), Tg (Thyroglobulin), TSH (Thyroid-Stimulating Hormone), TSH-R (Thyroid-Stimulating Hormone Receptor), TPO (Thyroid Peroxidase), TPOAb (TPO Antibody).

Author Contributions: Conceptualization, T.C.M.; methodology, T.C.M.; software, T.C.M.; formal analysis, T.C.M.; investigation, T.C.M.; resources, M.R., G.L., S.J.K, R.J.B.D, C.S, E.M.L., J.P.W., and S.G.W.; data curation, T.C.M., A.V., and M.M.; project administration, T.C.M. and T.D.S.; writing—original draft preparation, T.C.M.; writing—review and editing, T.C.M., S.N.K., T.D.S., J.P.W., K.I., M.B., M.P., and J.T.B.; validation, T.C.M. and T.D.S.; visualization, T.C.M.; supervision, S.N.K. and T.D.S.; funding acquisition, T.D.S., S.G.W., G.L., and S.N.K.. All authors have read and agreed to the published version of the manuscript.

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Conflicts of Interest: G.L. is the founder and CEO of Genos Ltd, a private research organization that specializes in high-throughput glycomic analysis and has several patents in this field. M.P. is an employee of Genos Ltd. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

Appendix A

A.1. Detection of TSH and TPOAb in human sera

Sera to assess TPOAb and TSH levels were collected by a trained nurse or phlebotomist using venepuncture and a SafetyLok™ Blood Collection Kit (21G3/4 Needles) and plain 10 ml serum-separating tube vacutainer (no additives) between February 1994 and May 2007. After collection from the study subject, whole blood was held at 22°C for 50 min at room temperature for a clot to form and serum separated within 60 minutes of collection. Processing of blood was performed using a refrigerated (4°C) clinical centrifuge at 3000xg for 10 minutes with the serum supernatant subsequently collected, transferred to a 2ml screw capped Nunc Cryotubes and immediately frozen at -80°C and kept frozen in 2ml screw capped Nunc Cryotube at -80°C until use. Quantitative determination of TSH and TPOAb (only IgG class) levels was performed on the sera either by a chemiluminescent microparticle immunoassay (CMIA) [ARCHITECT® Anti-TPO or TSH (ABBOTT Diagnostics Division, Wiesbaden, Germany, 2005)] (TPOAb titer>6 mIU/L considered positive; reference range for TSH level 0.4-4.0 mIU/L) or by an electrochemiluminescence immunoassay "ECLIA" [Elecsys and Cobas e analyzers, (Roche Diagnostics, Indianapolis, IN, USA, 2010)] (TPOAb titer>34 IU/mL considered positive; reference range for TSH 0.4-4.0 mIU/L).

A.2. Detection of IgG glycosylation profiling

For IgG glycosylation analysis, using UPLC analysis of 2AB-labelled glycans, chromatograms were separated in the same manner into 24 peaks, and the amount of glycans in each peak was expressed as a percentage of the total integrated area. One glycan was excluded before any transformation and standardization of data because of its co-elution with a contaminant that significantly affected its values in some samples whereas two glycan peaks (GP) GP20 and GP21 (Zagreb code) were combined into a single trait called GP2021 (Zagreb code) because of difficulty in distinguishing between these peaks in some samples. A global normalization and natural logarithm transformation were applied to 22 directly measured glycan structures. As many of these structures share the same structural features (galactose, sialic acid, core-fucose, bisecting N-acetylglucosamine (GlcNAc)), 55 additional derived traits were calculated that average these features across multiple glycans from the 22 normalized and non-transformed directly measured glycans. Technical confounders (batch and run-day effects) were addressed using R package ComBat. The 22 directly measured glycans and 55 derived glycan traits were centered and scaled to have a mean of 0 and standard deviation (SD) of 1. Samples being more than 6 SD from the mean were considered as outliers and excluded from the analysis.

A.3. Detection of protein profiling in plasma

Plasma protein profiling was conducted using SOMAscan v2 (SomaLogic Inc, Boulder, CO) as previously described [29,30]. Briefly, hemolyzed samples were first excluded. Proteins were then measured using a SOMAmer-based capture array called "SOMAscan." Quality control was performed at the sample and SOMAmer level and involves the use of control SOMAmers on the microarray and calibration samples. At the sample level, hybridization controls on the microarray are used to monitor sample-by-sample variability in hybridization, while the median signal over all SOMAmers is used to monitor overall technical variability. The resulting hybridization scale factor and median scale factor are used to normalize data across samples. The acceptance criteria for these values are 0.4–2.5, based on historical trends in these values. Somamer-by-somamer calibration occurs through the repeated measurement of calibration samples; these samples are of the same

matrix as the study samples and are used to monitor repeatability and batch to batch variability. Historical values for these calibrator samples for each SOMAmer are used to generate a calibration scale factor. The acceptance criteria for calibrator scale factors is that 95% of SOMAmers must have a calibration scale factor within ± 0.4 of the median. For the current analysis, only 1,113 proteins were then studied.

A.4. Selection of SNPs associated with immune cell traits

To define the list of SNPs associated with immune cell traits regardless to any specific phenotypes in the TwinsUK cohort, we extracted SNPs for each immune cell traits that have a P-value under GWAS P-value threshold ($P\text{-value} < 5 \times 10^{-8}$) from previous published GWASs on these immune cell traits [27,28]. To define the list of SNPs associated with protein abundance found in this study, we extracted the significant SNPs reported in INTERVAL project [31]. To define the list of SNPs associated with gene expression (eQTL), we extracted the eQTLs reported significant by GTEx and previous papers present in HaploReg V4.1 [47]. To define the list of SNPs associated with AITD and thyroid functions, we selected to SNPs listed in the NHGRI GWAS catalog [52] with words “thyroid” or “Graves” or “Hashimoto.”

A.5. Determination of effective number of independent tests for different -omic data

Due to high and partial correlations within glycans, proteins and immune cell traits, we decided to use the equation 5 proposed by Li & Ji ((2005) [45] to define an effective number (M_{eff}) of independent tests. We then used this number to define the effective Bonferroni P-value threshold such as $0.05/M_{\text{eff}}$ instead of $0.05/M$, with M the actual number of tests. 20 independent tests were estimated for 76 glycans. Consequently, to account for multiple testing in the discovery cohort, we present results surpassing a conservative Bonferroni correction assuming 20 independent tests, thus giving a significant threshold of ($P\text{-value} < 2.5 \times 10^{-3} = 0.05/20$). 1,357 independent tests were estimated for 23,485 immune cell traits, thus giving a significant threshold of 3.68×10^{-5} ($0.05/1,357$). 227 independent tests were estimated among 1,113 proteins ($P < 0.05/227 = 1.9 \times 10^{-4}$).

A.6. Association studies between -omics features and thyroid phenotypes

To examine whether one of the 17 AITD-IgG N-glycan traits was significantly associated with one of the 23,485 immune cell traits, we compared the fitted model in equation (2) with a model that did not include the residual of glycan in equation (1):

$$\text{Model null: } Y_i \sim a + h \text{ (fixe intercepts)} + g \text{ (random intercepts)} + \varepsilon_{ij} \quad (1)$$

$$\text{Model 1: } Y_i \sim a + \mathbf{bG}_{ij} + h \text{ (fixe intercepts)} + g \text{ (random intercepts)} + \varepsilon_{ij} \quad (2)$$

Where Y_i represents the quantification of immune cell traits for individual i and G_{ij} is glycan structure of type j among 75 N-glycans for the same individual i . If biological covariates (age, sex) have not been adjusted before association analysis, they have been added in the model. A random intercept was added only in the discovery cohort in order to model the family-relatedness.

To examine whether an immune cell trait was significantly associated with TPOAb level and AITD status, we compared the fitted model in equation (2) with a model that did not include the immune cell traits in equation (1) where G_{ij} become the immune cell trait of type j among 23,485 in discovery cohort for the same individual i . For the discovery and replication cohorts in TwinsUK, we added a random intercept in order to model the family-relatedness.

To examine whether one of the 1,113 protein was significantly associated with TPOAb level and AITD status, we compared the fitted model in equation (2) with a model that did not include the protein in equation (1): where G_{ij} become the protein of type j among 1,129 in discovery cohort for the same individual i . We added a random intercept in order to model the family-relatedness. To examine whether one of 1,113 proteins was significantly associated with one of 17 significant glycans, we compared the fitted model in equation (2) with a model that did not include the protein in equation (1): where G_{ij} become the protein of type j among 1,129 in discovery cohort for the same individual i . We added a random intercept in order to model the family-relatedness.

11.7. Heritability analysis for proteins

Using twin data and ADCE models (additive genetics (A), dominante genetics (D), shared environment (C) and non-shared environment (E)), heritability of glycosylation structures, immune cell traits and AITD were estimated using the R package called mets that allows us to run the analysis with monozygotic and dizygotic twins as well as unrelated individuals. The significance of variance components A, D, and C was assessed by dropping each component sequentially from the full model (ADCE) and comparing the sub-model fit to the full model. Sub-models were compared to full models by hierarchical χ^2 tests. The difference between log-likelihood values between sub-model and full model is asymptotically distributed as χ^2 with degrees of freedom (df) equal to the difference in df of sub-model and the full model. A statistical indicator of goodness-of-fit is the Akaike information criterion (AIC), computed as $\chi^2 - 2df$; sub-models are accepted as the best-fitting model if there is no significant loss of fit when a latent variable (A, C, D, or E) is fixed to equal zero. When two sub-models have the same AIC compared to the full model, we decide to keep the model the most likely (with additive genetic variance) or with the lowest P-value for different components.

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Autoimmune Thyroid Diseases

Healthy vs AITD
or
TPOAb level

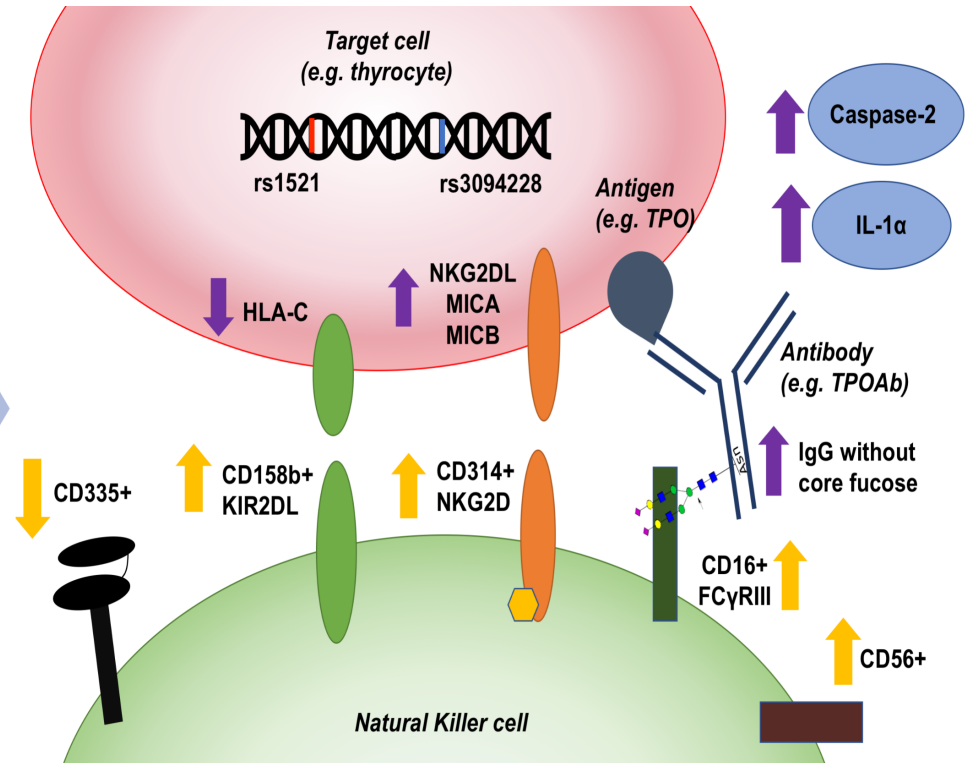
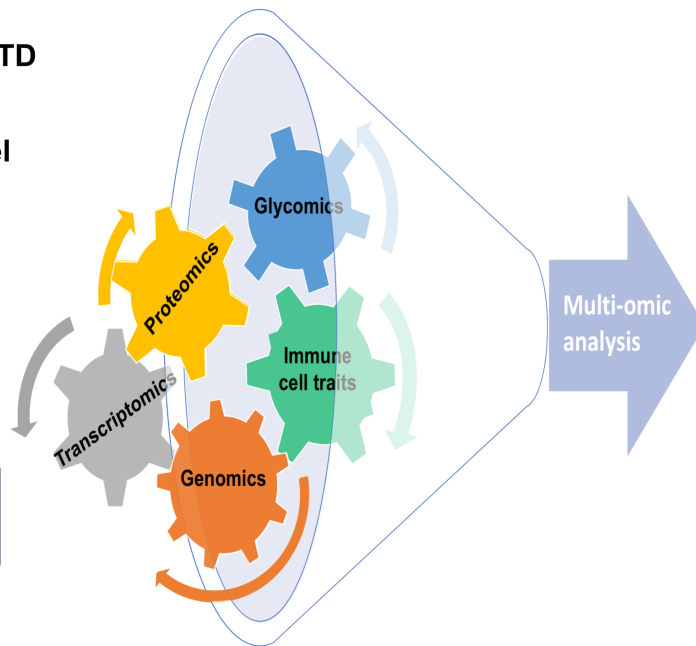


Table S1: Description of TwinsUK cohort used for different analysis performed here
* Roederer, M et al., Cell, 2015, doi: 10.1016/j.cell.2015.02.046 and Mangino et al, Nature Communications, 2017, <https://doi.org/10.1038/ncomms13850>

Analysis	GWAS on immune cell traits*	Immune-wide association studies with thyroid phenotypes						immune cell traits-wide association studies with IgG N-glycan traits		Proteome-wide association studies with thyroid phenotypes						Glycome-wide association studies with proteins			
Phenotype		TPOAb	AITD					TPOAb	AITD					TPOAb	AITD				
Group		continuous	control		case			continuous	control		case			continuous	control		case		
TPOAb immunoassay threshold of TPOAb-positivity (UI/mL)	NA	Roche	Abbott	Roche	Abbott	Roche	NA	Roche	Abbott	Roche	Abbott	Roche	NA	Roche	Abbott	Roche	Abbott	Roche	NA
	NA	34	6	34	6	34	NA	34	6	34	6	34	NA	34	6	34	6	34	NA
Number of individuals	497	374	77	127	19	22	383	155	25	105	9	18	164	61.19 (6.96)	56.59 (8.10)	61.64 (7.8)	55.20 (4.65)	60.66 (6.73)	64.20 (7.27)
Age (mean/sd)	60 (8.2)	55.8 (8.4)	52.59 (7.20)	55.41 (8.4)	51.58 (7.49)	53.40 (8.02)	60 (8.2)	61.19 (6.96)	56.59 (8.10)	61.64 (7.8)	55.20 (4.65)	60.66 (6.73)	64.20 (7.27)	61.19 (6.96)	56.59 (8.10)	61.64 (7.8)	55.20 (4.65)	60.66 (6.73)	64.20 (7.27)
Sex (F/M)	497/0	374/0	77/0	127/0	19/0	22/0	383/0	155/0	25/0	105/0	9/0	18/0	164/0	61.19 (6.96)	56.59 (8.10)	61.64 (7.8)	55.20 (4.65)	60.66 (6.73)	64.20 (7.27)
TPOAb (mean/sd)	NA	50.5 (108.4)	1.42 (8.49)	10.50 (5.37)	463.86 (568.45)	312.68 (146.01)	NA	66.65 (36.19)	0.14 (0.37)	9.33 (5.05)	585.66 (1156.48)	324.81 (147.69)	NA	66.65 (36.19)	0.14 (0.37)	9.33 (5.05)	585.66 (1156.48)	324.81 (147.69)	NA

Table S2: Significant glycome associations with immune cell traits in the TwinsUK cohort
SE=standart error

Glycan ID	Description	immuneTrait ID	Trait ID	Canonical name	Lineage	Subset name	Pvalue	Beta	SE	Zscore
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64161	P4:2970	NK Activating 1	16+56	16+56/314+335-337-158b+R7-	9.22E-08	0.285	0.052	5.445
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64161	P4:2970	NK Activating 1	16+56	16+56/314+335-337-158b+R7-	5.10E-08	0.277	0.050	5.555
IGP46	The percentage of A2G1 glycan in total Neutral IgG glycans (GPn)	immuno_64161	P4:2970	NK Activating 1	16+56	16+56/314+335-337-158b+R7-	9.29E-08	0.237	0.044	5.443
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_64161	P4:2970	NK Activating 1	16+56	16+56/314+335-337-158b+R7-	7.88E-08	-0.228	0.042	-5.477
IGP59	The percentage of fucosylation of agalactosylated structures	immuno_64161	P4:2970	NK Activating 1	16+56	16+56/314+335-337-158b+R7-	8.11E-08	-0.234	0.043	-5.468
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64161	P4:2970	NK Activating 1	16+56	16+56/314+335-337-158b+R7-	2.14E-08	-0.242	0.042	-5.718
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64190	P4:2997	NK Activating 1	16+56	16+56/314+335-158b+R7-	8.47E-08	0.286	0.052	5.461
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64190	P4:2997	NK Activating 1	16+56	16+56/314+335-158b+R7-	4.69E-08	0.278	0.050	5.571
IGP46	The percentage of A2G1 glycan in total Neutral IgG glycans (GPn)	immuno_64190	P4:2997	NK Activating 1	16+56	16+56/314+335-158b+R7-	8.94E-08	0.237	0.044	5.451
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_64190	P4:2997	NK Activating 1	16+56	16+56/314+335-158b+R7-	7.49E-08	-0.228	0.042	-5.487
IGP59	The percentage of fucosylation of agalactosylated structures	immuno_64190	P4:2997	NK Activating 1	16+56	16+56/314+335-158b+R7-	7.62E-08	-0.234	0.043	-5.480
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64190	P4:2997	NK Activating 1	16+56	16+56/314+335-158b+R7-	2.03E-08	-0.243	0.042	-5.728
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64953	P4:3683	NK Activating 1	16+56	16+56/314+335-337-158b+	9.58E-08	0.285	0.052	5.437
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64953	P4:3683	NK Activating 1	16+56	16+56/314+335-337-158b+	5.24E-08	0.277	0.050	5.550
IGP46	The percentage of A2G1 glycan in total Neutral IgG glycans (GPn)	immuno_64953	P4:3683	NK Activating 1	16+56	16+56/314+335-337-158b+	9.70E-08	0.237	0.044	5.435
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_64953	P4:3683	NK Activating 1	16+56	16+56/314+335-337-158b+	8.37E-08	-0.227	0.042	-5.466
IGP59	The percentage of fucosylation of agalactosylated structures	immuno_64953	P4:3683	NK Activating 1	16+56	16+56/314+335-337-158b+	8.77E-08	-0.233	0.043	-5.454
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64953	P4:3683	NK Activating 1	16+56	16+56/314+335-337-158b+	2.28E-08	-0.242	0.042	-5.706
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64984	P4:3710	NK Activating 1	16+56	16+56/314+335-158b+	8.51E-08	0.286	0.052	5.460
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64984	P4:3710	NK Activating 1	16+56	16+56/314+335-158b+	4.64E-08	0.278	0.050	5.573
IGP46	The percentage of A2G1 glycan in total Neutral IgG glycans (GPn)	immuno_64984	P4:3710	NK Activating 1	16+56	16+56/314+335-158b+	8.68E-08	0.238	0.044	5.456
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_64984	P4:3710	NK Activating 1	16+56	16+56/314+335-158b+	7.57E-08	-0.228	0.042	-5.485
IGP59	The percentage of fucosylation of agalactosylated structures	immuno_64984	P4:3710	NK Activating 1	16+56	16+56/314+335-158b+	7.85E-08	-0.234	0.043	-5.475
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64984	P4:3710	NK Activating 1	16+56	16+56/314+335-158b+	2.06E-08	-0.243	0.042	-5.726
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_62860	P4:18	NK Activating 2	16+56	16+56/CD2+314+335+337-158a-158b-R	8.79E-07	-0.250	0.050	-4.999
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_62860	P4:18	NK Activating 2	16+56	16+56/CD2+314+335+337-158a-158b-R	8.06E-07	-0.240	0.048	-5.025
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_62860	P4:18	NK Activating 2	16+56	16+56/CD2+314+335+337-158a-158b-R	5.26E-07	0.208	0.040	5.154
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_63595	P4:2460	NK Activating 2	16+56	16+56/335+337-158a-158b-R7-	1.77E-08	-0.284	0.049	-5.752
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_63595	P4:2460	NK Activating 2	16+56	16+56/335+337-158a-158b-R7-	9.79E-09	-0.275	0.047	-5.863
IGP46	The percentage of A2G1 glycan in total Neutral IgG glycans (GPn)	immuno_63595	P4:2460	NK Activating 2	16+56	16+56/335+337-158a-158b-R7-	4.86E-07	-0.211	0.041	-5.131
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_63595	P4:2460	NK Activating 2	16+56	16+56/335+337-158a-158b-R7-	3.67E-07	0.202	0.039	5.183
IGP59	The percentage of fucosylation of agalactosylated structures	immuno_63595	P4:2460	NK Activating 2	16+56	16+56/335+337-158a-158b-R7-	7.23E-07	0.204	0.040	5.044
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_63595	P4:2460	NK Activating 2	16+56	16+56/335+337-158a-158b-R7-	6.14E-08	0.221	0.040	5.540
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_63596	P4:2461	NK Activating 2	16+56	16+56/CD2+335+337-158a-158b-R7-	4.61E-07	-0.256	0.050	-5.134
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_63596	P4:2461	NK Activating 2	16+56	16+56/CD2+335+337-158a-158b-R7-	3.39E-07	-0.248	0.048	-5.206
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_63596	P4:2461	NK Activating 2	16+56	16+56/CD2+335+337-158a-158b-R7-	4.44E-07	0.210	0.040	5.189
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_63597	P4:2462	NK Activating 2	16+56	16+56/314+335+337-158a-158b-R7-	5.91E-08	-0.276	0.050	-5.527
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_63597	P4:2462	NK Activating 2	16+56	16+56/314+335+337-158a-158b-R7-	4.89E-08	-0.264	0.047	-5.565
IGP46	The percentage of A2G1 glycan in total Neutral IgG glycans (GPn)	immuno_63597	P4:2462	NK Activating 2	16+56	16+56/314+335+337-158a-158b-R7-	1.61E-06	-0.203	0.042	-4.891
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_63597	P4:2462	NK Activating 2	16+56	16+56/314+335+337-158a-158b-R7-	2.99E-07	0.212	0.040	5.239
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_63622	P4:2485	NK Activating 2	16+56	16+56/335+158a-158b-R7-	4.19E-08	-0.278	0.050	-5.594
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_63622	P4:2485	NK Activating 2	16+56	16+56/335+158a-158b-R7-	2.47E-08	-0.269	0.047	-5.697
IGP46	The percentage of A2G1 glycan in total Neutral IgG glycans (GPn)	immuno_63622	P4:2485	NK Activating 2	16+56	16+56/335+158a-158b-R7-	1.23E-06	-0.205	0.041	-4.945
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_63622	P4:2485	NK Activating 2	16+56	16+56/335+158a-158b-R7-	1.19E-06	0.195	0.039	4.948
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_63622	P4:2485	NK Activating 2	16+56	16+56/335+158a-158b-R7-	2.07E-07	0.214	0.040	5.312
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_63623	P4:2486	NK Activating 2	16+56	16+56/CD2+335+158a-158b-R7-	8.58E-07	-0.250	0.050	-5.010
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_63623	P4:2486	NK Activating 2	16+56	16+56/CD2+335+158a-158b-R7-	6.33E-07	-0.242	0.048	-5.082
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_63623	P4:2486	NK Activating 2	16+56	16+56/CD2+335+158a-158b-R7-	7.60E-07	0.206	0.040	5.082
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_63625	P4:2488	NK Activating 2	16+56	16+56/314+335+158a-158b-R7-	9.03E-08	-0.272	0.050	-5.447
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_63625	P4:2488	NK Activating 2	16+56	16+56/314+335+158a-158b-R7-	7.32E-08	-0.261	0.048	-5.490
IGP46	The percentage of fucosylation of monogalactosylated structures	immuno_63625	P4:2488	NK Activating 2	16+56	16+56/314+335+158a-158b-R7-	4.19E-07	0.210	0.041	5.173
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_63626	P4:2489	NK Activating 2	16+56	16+56/CD2+314+335+158a-158b-R7-	1.54E-06	-0.245	0.050	-4.885
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_63626	P4:2489	NK Activating 2	16+56	16+56/CD2+314+335+158a-158b-R7-	1.34E-06	-0.236	0.048	-4.921
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_63626	P4:2489	NK Activating 2	16+56	16+56/CD2+314+335+158a-158b-R7-	8.03E-07	0.206	0.041	5.070
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_63674	P4:2531	NK Activating 2	16+56	16+56/335+337-158b-R7-	4.62E-09	-0.282	0.047	-5.994
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_63674	P4:2531	NK Activating 2	16+56	16+56/335+337-158b-R7-	2.85E-09	-0.272	0.045	-6.079
IGP46	The percentage of A2G1 glycan in total Neutral IgG glycans (GPn)	immuno_63674	P4:2531	NK Activating 2	16+56	16+56/335+337-158b-R7-	2.26E-07	-0.207	0.039	-5.276
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_63674	P4:2531	NK Activating 2	16+56	16+56/335+337-158b-R7-	1.79E-07	0.198	0.037	5.319
IGP59	The percentage of fucosylation of agalactosylated structures	immuno_63674	P4:2531	NK Activating 2	16+56	16+56/335+337-158b-R7-	4.57E-07	0.198	0.039	5.131
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_63674	P4:2531	NK Activating 2	16+56	16+56/335+337-158b-R7-	3.49E-08	0.215	0.038	5.638

IGP2	The percentage of A2 glycan in total IgG glycans	immuno_63675	P4:2532	NK Activating 2	16+56	16+56/CD2+335+337-158b-R7-	2.26E-07	-0.251	0.048	-5.270
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_63675	P4:2532	NK Activating 2	16+56	16+56/CD2+335+337-158b-R7-	1.72E-07	-0.241	0.045	-5.333
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_63675	P4:2532	NK Activating 2	16+56	16+56/CD2+335+337-158b-R7-	1.62E-06	0.184	0.038	4.902
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_63675	P4:2532	NK Activating 2	16+56	16+56/CD2+335+337-158b-R7-	4.20E-07	0.200	0.038	5.190
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_63677	P4:2534	NK Activating 2	16+56	16+56/314+335+337-158b-R7-	1.42E-08	-0.275	0.048	-5.793
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_63677	P4:2534	NK Activating 2	16+56	16+56/314+335+337-158b-R7-	1.37E-08	-0.262	0.045	-5.798
IGP46	The percentage of A2G1 glycan in total Neutral IgG glycans (GPn)	immuno_63677	P4:2534	NK Activating 2	16+56	16+56/314+335+337-158b-R7-	7.02E-07	-0.200	0.040	-5.052
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_63677	P4:2534	NK Activating 2	16+56	16+56/314+335+337-158b-R7-	8.60E-07	0.189	0.038	5.008
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_63677	P4:2534	NK Activating 2	16+56	16+56/314+335+337-158b-R7-	1.39E-07	0.207	0.039	5.377
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_63678	P4:2535	NK Activating 2	16+56	16+56/CD2+314+335+337-158b-R7-	3.64E-07	-0.248	0.048	-5.174
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_63678	P4:2535	NK Activating 2	16+56	16+56/CD2+314+335+337-158b-R7-	3.79E-07	-0.236	0.046	-5.172
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_63678	P4:2535	NK Activating 2	16+56	16+56/CD2+314+335+337-158b-R7-	5.68E-07	0.199	0.039	5.129
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_63703	P4:2558	NK Activating 2	16+56	16+56/335+158b-R7-	6.83E-09	-0.280	0.047	-5.924
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_63703	P4:2558	NK Activating 2	16+56	16+56/335+158b-R7-	4.04E-09	-0.270	0.045	-6.017
IGP46	The percentage of A2G1 glycan in total Neutral IgG glycans (GPn)	immuno_63703	P4:2558	NK Activating 2	16+56	16+56/335+158b-R7-	2.97E-07	-0.205	0.039	-5.224
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_63703	P4:2558	NK Activating 2	16+56	16+56/335+158b-R7-	2.41E-07	0.197	0.037	5.262
IGP59	The percentage of fucosylation of agalactosylated structures	immuno_63703	P4:2558	NK Activating 2	16+56	16+56/335+158b-R7-	6.17E-07	0.196	0.039	5.071
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_63703	P4:2558	NK Activating 2	16+56	16+56/335+158b-R7-	4.67E-08	0.214	0.038	5.585
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_63704	P4:2559	NK Activating 2	16+56	16+56/CD2+335+158b-R7-	3.13E-07	-0.248	0.048	-5.206
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_63704	P4:2559	NK Activating 2	16+56	16+56/CD2+335+158b-R7-	2.36E-07	-0.239	0.045	-5.271
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_63704	P4:2559	NK Activating 2	16+56	16+56/CD2+335+158b-R7-	5.59E-07	0.198	0.039	5.133
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_63707	P4:2561	NK Activating 2	16+56	16+56/314+335+158b-R7-	1.98E-08	-0.272	0.048	-5.731
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_63707	P4:2561	NK Activating 2	16+56	16+56/314+335+158b-R7-	1.94E-08	-0.259	0.045	-5.734
IGP46	The percentage of A2G1 glycan in total Neutral IgG glycans (GPn)	immuno_63707	P4:2561	NK Activating 2	16+56	16+56/314+335+158b-R7-	1.06E-06	-0.197	0.040	-4.969
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_63707	P4:2561	NK Activating 2	16+56	16+56/314+335+158b-R7-	1.29E-06	0.185	0.038	4.925
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_63707	P4:2561	NK Activating 2	16+56	16+56/314+335+158b-R7-	2.06E-07	0.204	0.039	5.303
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_63708	P4:2562	NK Activating 2	16+56	16+56/CD2+314+335+158b-R7-	4.66E-07	-0.246	0.048	-5.125
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_63708	P4:2562	NK Activating 2	16+56	16+56/CD2+314+335+158b-R7-	4.49E-07	-0.235	0.046	-5.138
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_63708	P4:2562	NK Activating 2	16+56	16+56/CD2+314+335+158b-R7-	6.39E-07	0.198	0.039	5.105
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_63835	P4:2677	NK Activating 2	16+56	16+56/335+337-158a-R7-	1.61E-06	-0.222	0.045	-4.881
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_63925	P4:2758	NK Activating 2	16+56	16+56/335+337-R7-	1.32E-06	-0.210	0.043	-4.916
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64341	P4:3131	NK Activating 2	16+56	16+56/335+337-158a-158b-	1.57E-08	-0.286	0.050	-5.775
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64341	P4:3131	NK Activating 2	16+56	16+56/335+337-158a-158b-	8.68E-09	-0.277	0.047	-5.885
IGP46	The percentage of A2G1 glycan in total Neutral IgG glycans (GPn)	immuno_64341	P4:3131	NK Activating 2	16+56	16+56/335+337-158a-158b-	5.85E-07	-0.210	0.041	-5.094
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_64341	P4:3131	NK Activating 2	16+56	16+56/335+337-158a-158b-	4.26E-07	0.202	0.039	5.154
IGP59	The percentage of fucosylation of agalactosylated structures	immuno_64341	P4:3131	NK Activating 2	16+56	16+56/335+337-158a-158b-	7.39E-07	0.205	0.041	5.040
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64341	P4:3131	NK Activating 2	16+56	16+56/335+337-158a-158b-	7.26E-08	0.221	0.040	5.509
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64342	P4:3132	NK Activating 2	16+56	16+56/CD2+335+337-158a-158b-	4.75E-07	-0.256	0.050	-5.128
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64342	P4:3132	NK Activating 2	16+56	16+56/CD2+335+337-158a-158b-	3.55E-07	-0.247	0.048	-5.196
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64342	P4:3132	NK Activating 2	16+56	16+56/CD2+335+337-158a-158b-	4.46E-07	0.210	0.040	5.186
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64344	P4:3134	NK Activating 2	16+56	16+56/314+335+337-158a-158b-	4.41E-08	-0.278	0.050	-5.582
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64344	P4:3134	NK Activating 2	16+56	16+56/314+335+337-158a-158b-	3.59E-08	-0.267	0.047	-5.623
IGP46	The percentage of A2G1 glycan in total Neutral IgG glycans (GPn)	immuno_64344	P4:3134	NK Activating 2	16+56	16+56/314+335+337-158a-158b-	1.38E-06	-0.205	0.042	-4.921
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_64344	P4:3134	NK Activating 2	16+56	16+56/314+335+337-158a-158b-	1.59E-06	0.194	0.040	4.887
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64344	P4:3134	NK Activating 2	16+56	16+56/314+335+337-158a-158b-	2.30E-07	0.214	0.040	5.290
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64345	P4:3135	NK Activating 2	16+56	16+56/CD2+314+335+337-158a-158b-	7.22E-07	-0.252	0.050	-5.040
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64345	P4:3135	NK Activating 2	16+56	16+56/CD2+314+335+337-158a-158b-	6.52E-07	-0.242	0.048	-5.068
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64345	P4:3135	NK Activating 2	16+56	16+56/CD2+314+335+337-158a-158b-	3.94E-07	0.211	0.040	5.211
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64370	P4:3158	NK Activating 2	16+56	16+56/335+158a-158b-	2.28E-08	-0.283	0.050	-5.706
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64370	P4:3158	NK Activating 2	16+56	16+56/335+158a-158b-	1.27E-08	-0.274	0.047	-5.817
IGP46	The percentage of A2G1 glycan in total Neutral IgG glycans (GPn)	immuno_64370	P4:3158	NK Activating 2	16+56	16+56/335+158a-158b-	8.59E-07	-0.208	0.041	-5.018
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_64370	P4:3158	NK Activating 2	16+56	16+56/335+158a-158b-	6.07E-07	0.200	0.039	5.084
IGP59	The percentage of fucosylation of agalactosylated structures	immuno_64370	P4:3158	NK Activating 2	16+56	16+56/335+158a-158b-	1.07E-06	0.202	0.041	4.965
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64370	P4:3158	NK Activating 2	16+56	16+56/335+158a-158b-	1.03E-07	0.219	0.040	5.444
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64371	P4:3159	NK Activating 2	16+56	16+56/CD2+335+158a-158b-	7.27E-07	-0.252	0.050	-5.043
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64371	P4:3159	NK Activating 2	16+56	16+56/CD2+335+158a-158b-	5.35E-07	-0.244	0.048	-5.115
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64371	P4:3159	NK Activating 2	16+56	16+56/CD2+335+158a-158b-	5.96E-07	0.208	0.040	5.130
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64374	P4:3161	NK Activating 2	16+56	16+56/314+335+158a-158b-	6.54E-08	-0.275	0.050	-5.508
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64374	P4:3161	NK Activating 2	16+56	16+56/314+335+158a-158b-	5.23E-08	-0.264	0.048	-5.553
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64374	P4:3161	NK Activating 2	16+56	16+56/314+335+158a-158b-	2.93E-07	0.213	0.041	5.243
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64375	P4:3162	NK Activating 2	16+56	16+56/CD2+314+335+158a-158b-	1.52E-06	-0.245	0.050	-4.887
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64375	P4:3162	NK Activating 2	16+56	16+56/CD2+314+335+158a-158b-	1.36E-06	-0.235	0.048	-4.918
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64375	P4:3162	NK Activating 2	16+56	16+56/CD2+314+335+158a-158b-	8.44E-07	0.205	0.041	5.060
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64431	P4:3212	NK Activating 2	16+56	16+56/335+337-158b-	4.09E-09	-0.284	0.047	-6.015
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64431	P4:3212	NK Activating 2	16+56	16+56/335+337-158b-	2.54E-09	-0.274	0.045	-6.099

IGP46	The percentage of A2G1 glycan in total neutral IgG glycans (GPn)	immuno_64431	P4:3212	NK Activating 2	16+56	16+56/335+337-158b-	2.24E-07	-0.207	0.039	-5.277
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_64431	P4:3212	NK Activating 2	16+56	16+56/335+337-158b-	1.80E-07	0.199	0.037	5.319
IGP59	The percentage of fucosylation of agalactosylated structures	immuno_64431	P4:3212	NK Activating 2	16+56	16+56/335+337-158b-	4.14E-07	0.199	0.039	5.151
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64431	P4:3212	NK Activating 2	16+56	16+56/335+337-158b-	3.60E-08	0.215	0.038	5.632
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64432	P4:3213	NK Activating 2	16+56	16+56/CD2+335+337-158b-	1.89E-07	-0.253	0.048	-5.305
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64432	P4:3213	NK Activating 2	16+56	16+56/CD2+335+337-158b-	1.46E-07	-0.243	0.045	-5.363
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_64432	P4:3213	NK Activating 2	16+56	16+56/CD2+335+337-158b-	1.46E-06	0.185	0.038	4.922
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64432	P4:3213	NK Activating 2	16+56	16+56/CD2+335+337-158b-	3.89E-07	0.201	0.039	5.204
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64434	P4:3215	NK Activating 2	16+56	16+56/314+335+337-158b-	1.37E-08	-0.275	0.047	-5.799
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64434	P4:3215	NK Activating 2	16+56	16+56/314+335+337-158b-	1.40E-08	-0.262	0.045	-5.794
IGP46	The percentage of A2G1 glycan in total neutral IgG glycans (GPn)	immuno_64434	P4:3215	NK Activating 2	16+56	16+56/314+335+337-158b-	7.93E-07	-0.199	0.040	-5.027
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_64434	P4:3215	NK Activating 2	16+56	16+56/314+335+337-158b-	9.69E-07	0.188	0.038	4.984
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64434	P4:3215	NK Activating 2	16+56	16+56/314+335+337-158b-	1.61E-07	0.206	0.039	5.348
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64435	P4:3216	NK Activating 2	16+56	16+56/CD2+314+335+337-158b-	3.63E-07	-0.248	0.048	-5.175
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64435	P4:3216	NK Activating 2	16+56	16+56/CD2+314+335+337-158b-	3.74E-07	-0.236	0.046	-5.174
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64435	P4:3216	NK Activating 2	16+56	16+56/CD2+314+335+337-158b-	5.36E-07	0.199	0.039	5.140
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64460	P4:3239	NK Activating 2	16+56	16+56/335+158b-	4.94E-09	-0.283	0.047	-5.982
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64460	P4:3239	NK Activating 2	16+56	16+56/335+158b-	2.97E-09	-0.273	0.045	-6.072
IGP46	The percentage of A2G1 glycan in total neutral IgG glycans (GPn)	immuno_64460	P4:3239	NK Activating 2	16+56	16+56/335+158b-	2.69E-07	-0.206	0.039	-5.243
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_64460	P4:3239	NK Activating 2	16+56	16+56/335+158b-	2.31E-07	0.197	0.037	5.271
IGP59	The percentage of fucosylation of agalactosylated structures	immuno_64460	P4:3239	NK Activating 2	16+56	16+56/335+158b-	4.83E-07	0.198	0.039	5.120
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64460	P4:3239	NK Activating 2	16+56	16+56/335+158b-	4.55E-08	0.214	0.038	5.488
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64462	P4:3240	NK Activating 2	16+56	16+56/CD2+335+158b-	2.95E-07	-0.249	0.048	-5.218
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64462	P4:3240	NK Activating 2	16+56	16+56/CD2+335+158b-	2.28E-07	-0.240	0.045	-5.277
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64462	P4:3240	NK Activating 2	16+56	16+56/CD2+335+158b-	5.76E-07	0.198	0.039	5.126
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64464	P4:3242	NK Activating 2	16+56	16+56/314+335+158b-	2.02E-08	-0.272	0.048	-5.727
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64464	P4:3242	NK Activating 2	16+56	16+56/314+335+158b-	2.01E-08	-0.259	0.045	-5.728
IGP46	The percentage of A2G1 glycan in total neutral IgG glycans (GPn)	immuno_64464	P4:3242	NK Activating 2	16+56	16+56/314+335+158b-	1.16E-06	-0.196	0.040	-4.951
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_64464	P4:3242	NK Activating 2	16+56	16+56/314+335+158b-	1.41E-06	0.185	0.038	4.908
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64464	P4:3242	NK Activating 2	16+56	16+56/314+335+158b-	2.28E-07	0.204	0.039	5.282
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64465	P4:3243	NK Activating 2	16+56	16+56/CD2+314+335+158b-	4.83E-07	-0.246	0.048	-5.118
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64465	P4:3243	NK Activating 2	16+56	16+56/CD2+314+335+158b-	4.53E-07	-0.235	0.046	-5.136
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64465	P4:3243	NK Activating 2	16+56	16+56/CD2+314+335+158b-	6.49E-07	0.199	0.039	5.102
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64610	P4:3374	NK Activating 2	16+56	16+56/335+337-158a-	1.18E-06	-0.225	0.045	-4.944
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64641	P4:3401	NK Activating 2	16+56	16+56/335+158a-	1.61E-06	-0.222	0.046	-4.880
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64700	P4:3455	NK Activating 2	16+56	16+56/335+337-	1.14E-06	-0.211	0.043	-4.945
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64730	P4:3482	NK Activating 2	16+56	16+56/335+	1.56E-06	-0.208	0.043	-4.881
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_63906	P4:2740	NK Effector	16+56	16+56/335-337-R7-	1.83E-06	0.224	0.046	4.852
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_63906	P4:2740	NK Effector	16+56	16+56/335-337-R7-	1.19E-06	0.217	0.044	4.943
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_63935	P4:2767	NK Effector	16+56	16+56/335-R7-	1.60E-06	0.225	0.046	4.880
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_63935	P4:2767	NK Effector	16+56	16+56/335-R7-	1.05E-06	0.218	0.044	4.969
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64084	P4:2900	NK Effector	16+56	16+56/335-337-158a-158b+R7-	1.67E-06	0.246	0.051	4.862
IGP46	The percentage of A2G1 glycan in total neutral IgG glycans (GPn)	immuno_64084	P4:2900	NK Effector	16+56	16+56/335-337-158a-158b+R7-	9.62E-07	0.219	0.044	4.982
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64084	P4:2900	NK Effector	16+56	16+56/335-337-158a-158b+R7-	8.06E-07	-0.219	0.044	-5.017
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64157	P4:2967	NK Effector	16+56	16+56/335-337-158b+R7-	1.61E-07	0.286	0.054	5.340
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64157	P4:2967	NK Effector	16+56	16+56/335-337-158b+R7-	1.68E-07	0.274	0.051	5.328
IGP46	The percentage of A2G1 glycan in total neutral IgG glycans (GPn)	immuno_64157	P4:2967	NK Effector	16+56	16+56/335-337-158b+R7-	1.60E-07	0.240	0.045	5.339
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_64157	P4:2967	NK Effector	16+56	16+56/335-337-158b+R7-	5.30E-07	-0.225	0.044	-5.104
IGP59	The percentage of fucosylation of agalactosylated structures	immuno_64157	P4:2967	NK Effector	16+56	16+56/335-337-158b+R7-	5.31E-07	-0.227	0.045	-5.100
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64157	P4:2967	NK Effector	16+56	16+56/335-337-158b+R7-	8.38E-08	-0.244	0.045	-5.463
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64187	P4:2994	NK Effector	16+56	16+56/335-158b+R7-	1.52E-07	0.287	0.054	5.351
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64187	P4:2994	NK Effector	16+56	16+56/335-158b+R7-	1.58E-07	0.274	0.051	5.339
IGP46	The percentage of A2G1 glycan in total neutral IgG glycans (GPn)	immuno_64187	P4:2994	NK Effector	16+56	16+56/335-158b+R7-	1.52E-07	0.240	0.045	5.349
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_64187	P4:2994	NK Effector	16+56	16+56/335-158b+R7-	5.15E-07	-0.225	0.044	-5.110
IGP59	The percentage of fucosylation of agalactosylated structures	immuno_64187	P4:2994	NK Effector	16+56	16+56/335-158b+R7-	5.10E-07	-0.227	0.045	-5.108
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64187	P4:2994	NK Effector	16+56	16+56/335-158b+R7-	8.11E-08	-0.244	0.045	-5.470
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64680	P4:3437	NK Effector	16+56	16+56/335-337-	1.80E-06	0.207	0.043	4.851
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64710	P4:3464	NK Effector	16+56	16+56/335-	1.68E-06	0.207	0.043	4.866
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64950	P4:3680	NK Effector	16+56	16+56/335-337-158b+	1.54E-07	0.286	0.054	5.349
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64950	P4:3680	NK Effector	16+56	16+56/335-337-158b+	1.58E-07	0.274	0.051	5.340
IGP46	The percentage of A2G1 glycan in total neutral IgG glycans (GPn)	immuno_64950	P4:3680	NK Effector	16+56	16+56/335-337-158b+	1.47E-07	0.240	0.045	5.356
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_64950	P4:3680	NK Effector	16+56	16+56/335-337-158b+	4.96E-07	-0.226	0.044	-5.118
IGP59	The percentage of fucosylation of agalactosylated structures	immuno_64950	P4:3680	NK Effector	16+56	16+56/335-337-158b+	4.98E-07	-0.228	0.045	-5.112
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64950	P4:3680	NK Effector	16+56	16+56/335-337-158b+	7.72E-08	-0.244	0.045	-5.479
IGP2	The percentage of A2 glycan in total IgG glycans	immuno_64980	P4:3707	NK Effector	16+56	16+56/335-158b+	1.51E-07	0.287	0.054	5.353

IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	immuno_64980	P4:3707 NK Effector	16+56	16+56/335-158b+	1.54E-07	0.275	0.051	5.345
IGP46	The percentage of A2G1 glycan in total neutral IgG glycans (GPn)	immuno_64980	P4:3707 NK Effector	16+56	16+56/335-158b+	1.42E-07	0.240	0.045	5.362
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	immuno_64980	P4:3707 NK Effector	16+56	16+56/335-158b+	4.85E-07	-0.226	0.044	-5.122
IGP59	The percentage of fucosylation of agalactosylated structures	immuno_64980	P4:3707 NK Effector	16+56	16+56/335-158b+	4.90E-07	-0.228	0.045	-5.116
IGP60	The percentage of fucosylation of monogalactosylated structures	immuno_64980	P4:3707 NK Effector	16+56	16+56/335-158b+	7.54E-08	-0.244	0.045	-5.483

Table S3: Associations of 51 immune cell traits with AITD and TPOAb level in the TwinsUK cohort

immuneTrait ID	Trait ID	Canonical name	Lineage	Subset name	AITD				TPOAb level			
					Pvalue	Beta	SE	Zscore	Pvalue	Beta	SE	Zscore
immuno_64464	P4:3242	NK Activating 2	16+56	16+56/314+335+158b-	0.043	-0.049	0.024	-2.060	0.768	0.000	0.001	-0.297
immuno_63707	P4:2561	NK Activating 2	16+56	16+56/314+335+158b-R7-	0.045	-0.049	0.024	-2.041	0.769	0.000	0.001	-0.296
immuno_64374	P4:3161	NK Activating 2	16+56	16+56/314+335+158a-158b-	0.049	-0.046	0.023	-1.998	0.709	0.000	0.001	-0.377
immuno_63625	P4:2488	NK Activating 2	16+56	16+56/314+335+158a-158b-R7-	0.050	-0.046	0.023	-1.990	0.723	0.000	0.001	-0.357
immuno_64434	P4:3215	NK Activating 2	16+56	16+56/314+335+337-158b-	0.051	-0.047	0.024	-1.986	0.850	0.000	0.001	-0.191
immuno_63677	P4:2534	NK Activating 2	16+56	16+56/314+335+337-158b-R7-	0.051	-0.047	0.024	-1.985	0.817	0.000	0.001	-0.233
immuno_63703	P4:2558	NK Activating 2	16+56	16+56/335+158b-R7-	0.051	-0.048	0.024	-1.980	0.949	0.000	0.001	-0.065
immuno_64460	P4:3239	NK Activating 2	16+56	16+56/335+158b-	0.052	-0.047	0.024	-1.969	0.951	0.000	0.001	-0.062
immuno_64344	P4:3134	NK Activating 2	16+56	16+56/314+335+337-158a-158b-	0.055	-0.045	0.023	-1.951	0.756	0.000	0.001	-0.314
immuno_63597	P4:2462	NK Activating 2	16+56	16+56/314+335+337-158a-158b-R7-	0.056	-0.045	0.023	-1.945	0.753	0.000	0.001	-0.317
immuno_63674	P4:2531	NK Activating 2	16+56	16+56/335+337-158b-R7-	0.057	-0.047	0.024	-1.930	0.994	0.000	0.001	-0.008
immuno_64431	P4:3212	NK Activating 2	16+56	16+56/335+337-158b-	0.059	-0.046	0.024	-1.914	0.984	0.000	0.001	0.021
immuno_64370	P4:3158	NK Activating 2	16+56	16+56/335+158a-158b-	0.065	-0.044	0.023	-1.870	0.851	0.000	0.001	-0.190
immuno_63595	P4:2460	NK Activating 2	16+56	16+56/335+337-158a-158b-R7-	0.066	-0.044	0.023	-1.867	0.925	0.000	0.001	-0.095
immuno_63622	P4:2485	NK Activating 2	16+56	16+56/335+158a-158b-R7-	0.066	-0.044	0.023	-1.865	0.844	0.000	0.001	-0.198
immuno_64341	P4:3131	NK Activating 2	16+56	16+56/335+337-158a-158b-	0.072	-0.043	0.023	-1.826	0.945	0.000	0.001	-0.070
immuno_64641	P4:3401	NK Activating 2	16+56	16+56/335+158a-	0.086	-0.041	0.024	-1.729	0.750	0.000	0.001	0.324
immuno_63906	P4:2740	NK Effector	16+56	16+56/335-337-R7-	0.086	0.043	0.025	1.726	0.351	0.000	0.001	-0.954
immuno_64680	P4:3437	NK Effector	16+56	16+56/335-337-	0.087	0.043	0.025	1.727	0.365	0.000	0.001	-0.926
immuno_63935	P4:2767	NK Effector	16+56	16+56/335-R7-	0.087	0.043	0.025	1.720	0.335	-0.001	0.001	-0.987
immuno_64730	P4:3482	NK Activating 2	16+56	16+56/335+	0.088	-0.043	0.025	-1.719	0.358	0.000	0.001	0.941
immuno_64710	P4:3464	NK Effector	16+56	16+56/335-	0.088	0.043	0.025	1.718	0.355	0.000	0.001	-0.947
immuno_64610	P4:3374	NK Activating 2	16+56	16+56/335+337-158a-	0.094	-0.040	0.024	-1.688	0.701	0.000	0.001	0.389
immuno_63835	P4:2677	NK Activating 2	16+56	16+56/335+337-158a-R7-	0.096	-0.040	0.024	-1.675	0.710	0.000	0.001	0.378
immuno_64700	P4:3455	NK Activating 2	16+56	16+56/335+337-	0.104	-0.041	0.025	-1.637	0.313	0.001	0.001	1.035
immuno_63925	P4:2758	NK Activating 2	16+56	16+56/335+337-R7-	0.104	-0.041	0.025	-1.636	0.322	0.001	0.001	1.016
immuno_64465	P4:3243	NK Activating 2	16+56	16+56/CD2+314+335+158b-	0.153	-0.033	0.023	-1.443	0.988	0.000	0.001	-0.016
immuno_63708	P4:2562	NK Activating 2	16+56	16+56/CD2+314+335+158b-R7-	0.155	-0.033	0.023	-1.434	0.995	0.000	0.001	0.006
immuno_64375	P4:3162	NK Activating 2	16+56	16+56/CD2+314+335+158a-158b-	0.161	-0.032	0.023	-1.414	0.838	0.000	0.001	-0.211
immuno_63626	P4:2489	NK Activating 2	16+56	16+56/CD2+314+335+158a-158b-R7-	0.162	-0.032	0.023	-1.410	0.855	0.000	0.001	-0.188
immuno_64435	P4:3216	NK Activating 2	16+56	16+56/CD2+314+335+337-158b-	0.164	-0.033	0.023	-1.404	0.916	0.000	0.001	0.109
immuno_63678	P4:2535	NK Activating 2	16+56	16+56/CD2+314+335+337-158b-R7-	0.165	-0.033	0.023	-1.400	0.896	0.000	0.001	0.135
immuno_62860	P4:18	NK Activating 2	16+56	16+56/CD2+314+335+337-158a-158b-R7-	0.175	-0.031	0.023	-1.368	0.963	0.000	0.001	-0.048
immuno_63623	P4:2486	NK Activating 2	16+56	16+56/CD2+335+158a-158b-R7-	0.178	-0.031	0.023	-1.358	0.875	0.000	0.001	-0.162
immuno_63704	P4:2559	NK Activating 2	16+56	16+56/CD2+335+158b-R7-	0.179	-0.032	0.024	-1.354	0.939	0.000	0.001	0.079
immuno_64371	P4:3159	NK Activating 2	16+56	16+56/CD2+335+158a-158b-	0.179	-0.031	0.023	-1.356	0.856	0.000	0.001	-0.187
immuno_64462	P4:3240	NK Activating 2	16+56	16+56/CD2+335+158b-	0.180	-0.032	0.024	-1.354	0.940	0.000	0.001	0.078
immuno_64345	P4:3135	NK Activating 2	16+56	16+56/CD2+314+335+337-158a-158b-	0.180	-0.031	0.023	-1.351	0.923	0.000	0.001	-0.100
immuno_63675	P4:2532	NK Activating 2	16+56	16+56/CD2+335+337-158b-R7-	0.189	-0.031	0.024	-1.323	0.864	0.000	0.001	0.177
immuno_64432	P4:3213	NK Activating 2	16+56	16+56/CD2+335+337-158b-	0.189	-0.031	0.024	-1.326	0.896	0.000	0.001	0.135
immuno_64342	P4:3132	NK Activating 2	16+56	16+56/CD2+335+337-158a-158b-	0.197	-0.030	0.023	-1.300	0.981	0.000	0.001	-0.024
immuno_63596	P4:2461	NK Activating 2	16+56	16+56/CD2+335+337-158a-158b-R7-	0.202	-0.030	0.023	-1.286	0.978	0.000	0.001	-0.028
immuno_64157	P4:2967	NK Effector	16+56	16+56/335-337-158b-R7-	0.805	0.006	0.023	0.248	0.464	-0.001	0.001	-0.758
immuno_64187	P4:2994	NK Effector	16+56	16+56/335-158b+R7-	0.808	0.006	0.023	0.244	0.466	-0.001	0.001	-0.754
immuno_64980	P4:3707	NK Effector	16+56	16+56/335-158b+	0.816	0.005	0.023	0.233	0.459	-0.001	0.001	-0.765
immuno_64950	P4:3680	NK Effector	16+56	16+56/335-337-158b+	0.816	0.005	0.023	0.233	0.457	-0.001	0.001	-0.769
immuno_64161	P4:2970	NK Activating 1	16+56	16+56/314+335-337-158b+R7-	0.893	0.003	0.023	0.135	0.454	-0.001	0.001	-0.778
immuno_64953	P4:3683	NK Activating 1	16+56	16+56/314+335-337-158b+	0.896	0.003	0.023	0.130	0.453	-0.001	0.001	-0.778
immuno_64190	P4:2997	NK Activating 1	16+56	16+56/314+335-158b+R7-	0.904	0.003	0.023	0.121	0.453	-0.001	0.001	-0.780
immuno_64984	P4:3710	NK Activating 1	16+56	16+56/314+335-158b+	0.905	0.003	0.023	0.120	0.453	-0.001	0.001	-0.778
immuno_64084	P4:2900	NK Effector	16+56	16+56/335-337-158a-158b+R7-	0.970	0.001	0.024	0.038	0.528	-0.001	0.001	-0.648

Table S4: Hits from selected eQTL studies for two SNPs, rs1521 and rs3094228 in the thyroid cells and whole blood

* GWAS catalog for the genetic variants associated with thyroid phenotypes(Data downloaded from GWAS Catalog on 17/06/2015)

** eQTLs from Haploreg v4.1 (<https://pubs.broadinstitute.org/mammals/haploreg/haploreg.php>) and GTEx v6,v7, v8 (<https://www.gtexportal.org/home/>)

Lead SNP- risk allele	Thyroid phenotypes (from GWAS catalog)*				Study ID	Paper Title	eQTL** PMID	ref allele	Tissue	Correlated gene	NES	p-value					
rs1521-T	6	31382927	31382927	Graves' disease	GTEx2019_v8 (dbGaP Accession phs000424.v8.p2)	The GTEx Consortium atlas of genetic regulatory effects across human tissues	10.1101/290225	C	Thyroid	VARS2	ENSG00000137411.16	0.29	5.00E-12				
									Thyroid	MIR6891	ENSG00000277402.1	0.35	6.20E-11				
									Thyroid	HCG27	ENSG00000206344.7	0.21	1.40E-09				
									Thyroid	HLA-C	ENSG00000204525.16	-0.24	8.20E-08				
									Thyroid	PSORS1C3	ENSG00000204528.3	0.33	7.50E-08				
									Thyroid	XXbac-BPG181B23.7	ENSG00000272221.1	-0.25	1.40E-06				
									Thyroid	MICA	ENSG00000204520.12	-0.18	2.50E-05				
									Thyroid	PRRT1	ENSG00000204314.10	0.12	4.20E-05				
									Thyroid	FLOT1	ENSG00000137312.14	0.11	6.50E-05				
									Whole_Blood	XXbac-BPG181B23.7	ENSG00000272221.1	-0.34	2.20E-17				
									Whole_Blood	MIR6891	ENSG00000277402.1	0.28	1.40E-09				
									Whole_Blood	PSORS1C3	ENSG00000204528.3	0.21	2.30E-08				
									Whole_Blood	XXbac-BPG299F13.17	ENSG00000272501.1	-0.12	2.20E-07				
									Whole_Blood	HCG27	ENSG00000206344.7	0.088	1.20E-05				
									Whole_Blood	VARS2	ENSG00000137411.16	0.1	2.20E-05				
									Whole_Blood	DDR1	ENSG00000204580.13	0.17	2.80E-05				
									Whole_Blood	ZBTB12	ENSG00000204366.3	0.2	3.20E-05				
									Whole_Blood	HLA-C	ENSG00000204525.16	-0.12	5.70E-05				
									Whole_Blood	C4A	ENSG00000244731.7	-0.2	1.30E-04				
									Whole_Blood	FLOT1	ENSG00000137312.14	0.045	1.70E-04				
									Thyroid	VARS2	ENSG00000137411.12	0.31	3.40E-09				
									Thyroid	HCG27	ENSG00000206344.6	0.23	6.90E-08				
					GTEx2017_v7 (dbGaP Accession phs000424.v7.p2)	The Genotype-Tissue Expression (GTEx) pilot analysis: Multitissue gene regulation in humans	290225	C	Thyroid	HLA-C	ENSG00000204525.10	-0.25	8.30E-06				
									Thyroid	PSORS1C3	ENSG00000204528.3	0.31	2.00E-05				
									Whole_Blood	XXbac-BPG181B23.7	ENSG00000272221.1	-0.33	7.90E-08				
									Thyroid	HLA-C	ENSG00000204525.10	NA	1.28E-08				
									Thyroid	XXbac-BPG248L24.12	NA	1.03E-08					
									Whole_Blood	HLA-C	ENSG00000204525.10	NA	2.98E-08				
									Whole_Blood	XXbac-BPG181B23.7	ENSG00000272221.1	NA	2.86E-07				
									Whole_Blood	XXbac-BPG248L24.12	NA	8.29E-09					
									Whole_Blood	-	NA	1.11E-35	NA				
									Whole_Blood	-	NA	6.64E-38	NA				
									Whole_Blood	AIF1	ENSG00000204472	NA	2.51E-06				
									GTEx2015_v6 (dbGaP Accession phs000424.v6.p2)	The Genotype-Tissue Expression (GTEx) pilot analysis: Multitissue gene regulation in humans	259540	C	Whole_Blood	HCP5	ENSG00000206337.6	NA	4.62E-06
					Whole_Blood	LTA	ENSG00000226879	NA					8.46E-10				
					Whole_Blood	-	NA	1.11E-35					NA				
					Whole_Blood	-	NA	6.64E-38					NA				
					Whole_Blood	AIF1	ENSG00000204472	NA					2.51E-06				
					Whole_Blood	ATP6V1G2 BAT1	ENSG00000213760	NA					9.31E-16				
					rs3094228-C	6	31462150	31462150					Thyroid peroxidase antibody positivity (TPOAb-positivity)	GTEx2019_v8 (dbGaP Accession phs000424.v8.p2)	The GTEx Consortium atlas of genetic regulatory effects across human tissues	10.1101/290225	T
Thyroid	CYP21A1P	ENSG00000204338.8	-0.37	2.80E-14													
Thyroid	MICB	ENSG00000204516.9	0.26	3.70E-09													
Thyroid	HCP5	ENSG00000206337.10	-0.24	3.00E-08													
Thyroid	FLOT1	ENSG00000137312.14	0.17	5.00E-08													
Thyroid	PSORS1C1	ENSG00000204540.10	-0.37	7.00E-08													
Thyroid	CCHCR1	ENSG00000204536.13	0.2	2.00E-07													
Thyroid	HLA-C	ENSG00000204525.16	0.25	9.30E-07													
Thyroid	HLA-S	ENSG00000225851.1	-0.31	3.80E-06													
Thyroid	RNF5	ENSG00000204308.7	-0.14	2.70E-05													
Whole_Blood	C4A	ENSG00000244731.7	-0.48	1.50E-16													
Whole_Blood	C4B	ENSG00000224389.8	0.47	2.60E-13													
Whole_Blood	CYP21A1P	ENSG00000204338.8	-0.4	1.10E-12													
Whole_Blood	CYP21A2	ENSG00000231852.6	0.4	3.20E-10													
Whole_Blood	FLOT1	ENSG00000137312.14	0.058	2.20E-05													
Whole_Blood	HLA-S	ENSG00000225851.1	-0.27	3.70E-05													
Whole_Blood	CCHCR1	ENSG00000204536.13	0.13	8.10E-05													
GTEx2017_v7 (dbGaP Accession phs000424.v7.p2)	The Genotype-Tissue Expression (GTEx) pilot analysis: Multitissue gene regulation in humans	290225	T	Whole_Blood					XXbac-BPG248L24.12	ENSG00000271581.1	-0.24	1.50E-04					
				Thyroid					C4A	ENSG00000244731.3	-0.4	3.40E-13					
				Thyroid					CYP21A1P	ENSG00000204338.4	-0.37	1.10E-09					
				Thyroid					HCP5	ENSG00000206337.6	-0.32	1.30E-09					
				Thyroid					PSORS1C1	ENSG00000204540.6	-0.4	4.00E-06					
				Thyroid					HCG22	ENSG00000228789.2	-0.35	5.10E-06					
				Thyroid					MICB	ENSG00000204516.5	0.26	5.10E-06					
				Thyroid					CCHCR1	ENSG00000204536.9	0.21	1.70E-05					
				Whole_Blood					C4A	ENSG00000244731.3	-0.48	4.00E-09					
				Whole_Blood					CYP21A1P	ENSG00000204338.4	-0.41	3.80E-07					
				Whole_Blood					C4B	ENSG00000224389.4	0.41	9.90E-06					
				Whole_Blood					CYP21A2	ENSG00000231852.2	0.38	2.90E-05					
rs2596460	6	31449483	31449483	16+56 CD2-314+335-337-158a+158b+	GTEx2019_v8 (dbGaP Accession phs000424.v8.p2)	The GTEx Consortium atlas of genetic regulatory effects across human tissues	10.1101/290225	A	Thyroid	ATP6V1G2	ENSG00000213760.10	0.28	1.10E-05				
									Thyroid	SKIV2L	ENSG00000204351.11	0.23	2.20E-05				
									Thyroid	CCHCR1	ENSG00000204536.13	-0.22	1.30E-04				
					GTEx2017_v7 (dbGaP Accession phs000424.v7.p2)	The Genotype-Tissue Expression (GTEx) pilot analysis: Multitissue gene regulation in humans	290225	A	Thyroid	VARS2	ENSG00000137411.16	-0.27	2.00E-04				
									Whole_Blood	MICA	ENSG00000204520.12	0.22	3.40E-04				
									Whole_Blood	HLA-C	ENSG00000204525.10	-0.3	5.60E-05				
rs2596457	6	31449995	31449995	16+56 CD2-314+335-337-158a+158b+	GTEx2019_v8 (dbGaP Accession phs000424.v8.p2)	The GTEx Consortium atlas of genetic regulatory effects across human tissues	10.1101/290225	A	Thyroid	CCHCR1	ENSG00000204536.9	-0.29	2.40E-05				
									Thyroid	ATP6V1G2	ENSG00000213760.10	0.3	2.10E-06				
									Thyroid	SKIV2L	ENSG00000204351.11	0.24	8.70E-06				
					GTEx2017_v7 (dbGaP Accession phs000424.v7.p2)	The Genotype-Tissue Expression (GTEx) pilot analysis: Multitissue gene regulation in humans	290225										
rs2523691	6	31452660	31452660	16+56 CD2-314+335-337-158a+158b+	GTEx2019_v8 (dbGaP Accession phs000424.v8.p2)	The GTEx Consortium atlas of genetic regulatory effects across human tissues	10.1101/290225	G	Thyroid	ATP6V1G2	ENSG00000213760.10	0.28	1.10E-05				
									Thyroid	SKIV2L	ENSG00000204351.11	0.23	2.20E-05				
									Thyroid	CCHCR1	ENSG00000204536.13	-0.22	1.30E-04				
					GTEx2017_v7 (dbGaP Accession phs000424.v7.p2)	The Genotype-Tissue Expression (GTEx) pilot analysis: Multitissue gene regulation in humans	290225										

Table S5 : Genes reported for genetic variants associated with thyroid phenotypes and immune cell traits

* the closest genes reported in GWAS catalog for the genetic variants associated with thyroid phenotypes and immune cell traits (Data downloaded from GWAS Catalog on 17/06/2015)

** Roederer, M et al., Cell, 2015, doi: 10.1016/j.cell.2015.02.046 and Mangino et al, Nature Communications, 2017, <https://doi.org/10.1038/ncomms13850>

Name of gene reported in GWAS catalog* and Roederer's paper **	Thyroid phenotypes (from GWAS catalog)*					A1TD status or biomarkers associated with A1TD	LD (r2)	distance lead SNP(thyroid)-lead SNP(immune cell traits)	Immune cell traits**						Phenotypes		
	Lead SNP	Chromosome	start (hg38)	end (hg38)	Phenotype				Lead SNP	Chromosome	start (hg38)	end (hg38)	start (hg19)	end (hg19)		Immune Name in TwinsUK	
HCP5	rs3094228	6	31462150	31462150	Thyroid peroxidase antibody positivity	Yes	<0.2	12667	rs2596460	6	31449483	31449483	31449733	31449733	P4:3763	NK Activating 1	16+56/CD2-314+335-337-158a+158b+
							<0.2	12155	rs2596457	6	31449995	31449995	31450245	31450245	P4:3763	NK Activating 1	16+56/CD2-314+335-337-158a+158b+
							<0.2	9490	rs2523691	6	31452660	31452660	31452910	31452910	P4:3763	NK Activating 1	16+56/CD2-314+335-337-158a+158b+
MICA	rs1521	6	31382927	31382927	Graves' disease	Yes	<0.2	-66556	rs2596460	6	31449483	31449483	31449733	31449733	P4:3763	NK Activating 1	16+56/CD2-314+335-337-158a+158b+
ACCN1	rs9901756	17	34137135	34137135	Hypothyroidism	No	<0.2	365816	rs12603968	17	33771319	33771319	33771319	33771319	P4:3551	NK Effector	16+56/314-158a+
							<0.2	366131	rs12602135	17	33771004	33771004	33771004	33771004	P4:3551	NK Effector	16+56/314-158a+
DIRC3	rs6759952 rs966423	2	217406996	217406996	Thyroid cancer	No	<0.2	-317240	rs744564	2	217724236	217724236	217724236	217724236	P4:3551	NK Effector	16+56/314-158a+
							<0.2	-278619									
GLIS3	rs1571583	9	4267209	4267209	Thyroid hormone levels	No	<0.2	492850	rs10973456	9	3774359	3774359	3774359	3774359	P7:110	DC mDC imDC	11c+ (nodim)/1c-/16+/32+
HACE1	rs9322817	6	104784358	104784358	Thyroid stimulating hormone level	No	<0.2	378569	rs156205	6	104405789	104405789	104405789	104405789	P4:3763	NK Activating 1	16+56/CD2-314+335-337-158a+158b+
L3MBTL4	rs948426	18	6567183	6567183	Hypothyroidism	No	<0.2	438743	rs17486103	18	6128440	6128440	6128440	6128440	P7:110	DC mDC imDC	11c+ (nodim)/1c-/16+/32+
NFIA	rs334699	1	61154824	61154824	Thyroid hormone levels	No	<0.2	-240528	rs11581697	1	61395352	61395352	61395352	61395352	P4:3551	NK Effector	16+56/314-158a+
							<0.2	-240632	rs12072379	1	61395456	61395456	61395456	61395456	P4:3551	NK Effector	16+56/314-158a+
NFIB	rs10961534	9	14470835	14470835	Hypothyroidism	No	<0.2	31760	rs11787815	9	14439075	14439075	14439075	14439075	P4:3551	NK Effector	16+56/314-158a+
NR3C2	rs10028213	4	148731458	148731458	Thyroid stimulating hormone level	No	<0.2	441933	rs3910047	4	148289525	148289525	148289525	148289525	P4:3551	NK Effector	16+56/314-158a+
NRG1	rs7825175	8	32558756	32558756	Thyroid hormone levels	No	<0.2	5271541	rs4279551	8	27287215	27287215	27287215	27287215	P7:110	DC mDC imDC	11c+ (nodim)/1c-/16+/32+
							<0.2	510838	rs2881470	8	32047918	32047918	32047918	32047918	P7:110	DC mDC imDC	11c+ (nodim)/1c-/16+/32+
							<0.2	488634	rs1462900	8	32070122	32070122	32070122	32070122	P7:110	DC mDC imDC	11c+ (nodim)/1c-/16+/32+
							<0.2	480970	rs1381871	8	32077786	32077786	32077786	32077786	P7:110	DC mDC imDC	11c+ (nodim)/1c-/16+/32+
							<0.2	496755	rs1471387	8	32078096	32078096	32078096	32078096	P7:110	DC mDC imDC	11c+ (nodim)/1c-/16+/32+
	rs2439302	8	32574851	32574851	Thyroid cancer	No	<0.2	494464	rs1947734	8	32080387	32080387	32080387	32080387	P7:110	DC mDC imDC	11c+ (nodim)/1c-/16+/32+
							<0.2	488995	rs7818326	8	32085856	32085856	32085856	32085856	P7:110	DC mDC imDC	11c+ (nodim)/1c-/16+/32+
							<0.2	488541	rs16878780	8	32086310	32086310	32086310	32086310	P7:110	DC mDC imDC	11c+ (nodim)/1c-/16+/32+
							<0.2	488339	rs9297185	8	32086512	32086512	32086512	32086512	P7:110	DC mDC imDC	11c+ (nodim)/1c-/16+/32+
							<0.2	152041	rs551901	6	165480954	165480954	165480954	165480954	P4:3763	NK Activating 1	16+56/CD2-314+335-337-158a+158b+
PDE10A	rs753760	6	165632995	165632995	Thyroid hormone levels	No	<0.2	151764	rs684847	6	165481231	165481231	165481231	165481231	P4:3763	NK Activating 1	16+56/CD2-314+335-337-158a+158b+
SOX9	rs9915657	17	72131395	72131395	Thyroid hormone levels	No	<0.2	691727	rs9302936	17	71439668	71439668	71439668	71439668	P4:3763	NK Activating 1	16+56/CD2-314+335-337-158a+158b+
							<0.2	686259	rs1990222	17	71445136	71445136	71445136	71445136	P4:3763	NK Activating 1	16+56/CD2-314+335-337-158a+158b+
VAV3	rs12126655	1	107814198	107814198	Plasma thyroid-stimulating hormone levels	No	<0.2	-121433	rs10494086	1	107935631	107935631	107935631	107935631	P4:3551	NK Effector	16+56/314-158a+
							<0.2	-127812	rs1020812	1	107942010	107942010	107942010	107942010	P4:3551	NK Effector	16+56/314-158a+
	rs4915077	1	107823394	107823394	Hypothyroidism	No	<0.2	-123692	rs6696531	1	107947086	107947086	107947086	107947086	P4:3551	NK Effector	16+56/314-158a+

Table S6: Glycome-wide associations studies of 17 AITD-IgG N-glycan traits with 1,113 circulating proteins. Only significant ones were put here.

Glycan ID	Description	Soma ID	Target	Uniprot	P-value	Beta	SE
IGP2	The percentage of A2 glycan in total IgG glycans	SL008609	FCG3B	O75015	1.46E-06	0.051	0.010
IGP8	The percentage of FA2[3]G1 glycan in total IgG glycans	SL007464	AMHR2	Q16671	1.50E-07	0.029	0.005
IGP8	The percentage of FA2[3]G1 glycan in total IgG glycans	SL004857	Desmoglein-2	Q14126	2.15E-06	0.032	0.007
IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	SL008609	FCG3B	O75015	2.63E-06	0.050	0.010
IGP46	The percentage of A2G1 glycan in total neutral IgG glycans (GPn)	SL008609	FCG3B	O75015	3.88E-06	0.045	0.009
IGP48	The percentage of FA2[3]G1 glycan in total neutral IgG glycans (GPn)	SL002644	ERBB1	P00533	3.04E-06	0.023	0.005
IGP48	The percentage of FA2[3]G1 glycan in total neutral IgG glycans (GPn)	SL000283	b2-Microglobulin	P61769	4.16E-06	-0.034	0.007
IGP56	The percentage of monogalactosylated structures in total neutral IgG glycans	SL000283	b2-Microglobulin	P61769	5.10E-07	-0.037	0.007
IGP57	The percentage of digalactosylated structures in total neutral IgG glycans	SL004159	TRAIL R4	Q9UBN6	2.95E-06	-0.041	0.008
IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	SL008609	FCG3B	O75015	5.21E-06	-0.044	0.009
IGP59	The percentage of fucosylation of agalactosylated structures	SL008609	FCG3B	O75015	9.33E-06	-0.044	0.010
IGP60	The percentage of fucosylation of monogalactosylated structures	SL008609	FCG3B	O75015	4.06E-07	-0.048	0.009
IGP62	The percentage of fucosylated (without bisecting GlcNAc) structures in total neutral IgG glycans	SL004672	BCMA	Q02223	5.44E-08	0.041	0.007
IGP63	The percentage of fucosylation (without bisecting GlcNAc) of agalactosylated structures	SL004672	BCMA	Q02223	1.47E-07	0.041	0.007
IGP63	The percentage of fucosylation (without bisecting GlcNAc) of agalactosylated structures	SL004857	Desmoglein-2	Q14126	8.02E-06	0.032	0.007

Table S7: Genetic variants associated with thyroid phenotypes and AITD-IgG N-glycan traits overlapping pQTL identified in INTERVAL project (LD r2>0.8)
 *Sun et al, Nature, 2018, doi: 10.1038/s41586-018-0175-2

GWAS Catalog (Data downloaded from GWAS Catalog on 17/06/2015)											SOMAmer ID (version 2) used here	pQTL*							
chr	Start (hg38)	Stop (hg38)	Strongest SNPs	SNPs	Disease traits	Region	Reported Genes	Mapped Genes	Context	Extra Information about GWAS findings		SOMAmer ID (version 4)	Target	Target fullname	UniProt	cis/ trans	Mapped gene	Sentinel variant	LD with sentinel variant (r2)
1	157699488	157699488	rs3761959-A, rs3761959-G	rs3761959	Graves disease , Multiple sclerosis	1q23.1	FCRL3	FCRL3	intron		SL014088	FCRL3.4440.15.2	FCRL3	Fc receptor-like protein 3	Q96P31	cis	FCRL3	rs7528684	1
1	157701026	157701026	rs7528684-T	rs7528684	Type 1 diabetes autoantibodies, Rheumatoid arthritis, Graves disease	1q23.1	FCRL3	FCRL3	nearGene-5	I-A2A									
9	133273813	133273813	rs505922-C, rs505922-T, rs505922-7	rs505922	Protein quantitative trait loci, Venous thromboembolism, End-stage coagulation, Pancreatic cancer, Graves disease , Duodenal ulcer	9q34.2	ABO	ABO	intron	Recessive, TNFA, vWF	No detected	DHFR.9823.2.3	DYR	Dihydrofolate reductase	P00374	trans	ABO	rs676457	0.99
											SL004516	MBL2.3000.66.1	MBL	Mannose-binding protein C	P11226	trans	ABO	rs139840563	0.98
											No detected	ABO.9253.52.3	BGAT	Histo-blood group ABO system transferase	P16442	cis	ABO	rs505922	1
											SL004857	DSG2.9484.75.3	Desmoglein-2	Desmoglein-2	Q14126	trans	ABO	rs687621	0.97
											SL005157	CD209.3029.52.2	DC-SIGN	CD209 antigen	Q9NNX6	trans	ABO	rs505922	1
12	111446804	111446804	rs3184504-C, rs3184504-T, rs3184504-T	rs3184504	Type 1 diabetes, Platelet counts, Blood metabolite levels, Beta-2 microglobulin plasma levels, Diastolic blood pressure, Coronary artery disease, Eosinophil counts, Systolic blood pressure, Autoimmune hepatitis type-1, Red blood cell traits, Rheumatoid arthritis, Type 1 diabetes autoantibodies, Hypothyroidism , Coronary heart disease	12q24.12	SH2B3, NAA25, C12orf51, ATXN2, BRAP, LOC100101246, PTPN11	SH2B3	missense	T1D, kynurenine, Hgb, EA	SL006460	GP1BA.4990.87.1	GP1BA	Platelet glycoprotein Ib alpha chain	P07359	trans	BRAP	rs11065979	0.81
												VCAM1.2967.8.1	VCAM-1	Vascular cell adhesion protein 1	P19320	trans	SH2B3	rs3184504	1

Table S8: Heritability of AITD, 17 IgG N-glycan traits and 51 immune cell traits in the TwinsUK cohort

* Martin et al, 2018

** Mangino et al, Nature Communications, 2017, <https://doi.org/10.1038/ncomms13850>

type	features	general info	best model	H2 [95%CI]	A [95%CI]	D [95%CI]	C [95%CI]	E [95%CI]
thyroid phenotype*	AITD TPOAb-positivity		DE	0.63 [0.59-0.67]	NS	0.63 [0.59-0.67]	NS	0.36 [0.32-0.40]
			DE	0.57 [0.50-0.65]	NS	0.57 [0.50-0.65]	NS	0.42 [0.34-0.49]
IgG N-glycan traits*	IGP2	The percentage of A2 glycan in total IgG glycans	AE	0.731 [0.747;0.716]	0.731 [0.697;0.764]	NS	NS	0.269 [0.236;0.303]
	IGP7	The percentage of FA2[6]G1 glycan in total IgG glycans	AE	0.557 [0.563;0.552]	0.557 [0.509;0.605]	NS	NS	0.443 [0.395;0.491]
	IGP8	The percentage of FA2[3]G1 glycan in total IgG glycans	AE	0.662 [0.676;0.65]	0.662 [0.621;0.702]	NS	NS	0.338 [0.298;0.379]
	IGP15	The percentage of FA2G1S1 glycan in total IgG glycans	AE	0.704 [0.72;0.691]	0.704 [0.669;0.739]	NS	NS	0.296 [0.261;0.331]
	IGP21	The percentage of A2B32S2 glycan in total IgG glycans	AE	0.347 [0.327;0.363]	0.347 [0.288;0.406]	NS	NS	0.653 [0.594;0.712]
	IGP33	Ratio of all fucosylated (+/- bisecting GlyNAc) monosialylated and disialylated structures in total IgG glycans	DCE	0.241 [0.183;0.277]	NS	0.241 [0.142;0.34]	0.246 [0.175;0.316]	0.513 [0.457;0.57]
	IGP36	Ratio of all fucosylated sialylated structures with and without bisecting GlcNAc	AE	0.704 [0.72;0.691]	0.704 [0.669;0.739]	NS	NS	0.296 [0.261;0.331]
	IGP42	The percentage of A2 glycan in total neutral IgG glycans (GPn)	AE	0.739 [0.755;0.724]	0.739 [0.706;0.771]	NS	NS	0.261 [0.229;0.294]
	IGP45	The percentage of FA2B glycan in total neutral IgG glycans (GP ⁿ)	DCE	0.41 [0.407;0.413]	NS	0.41 [0.346;0.475]	0.361 [0.305;0.416]	0.229 [0.199;0.259]
	IGP46	The percentage of A2G1 glycan in total neutral IgG glycans (GP ⁿ)	AE	0.725 [0.742;0.711]	0.725 [0.691;0.76]	NS	NS	0.275 [0.24;0.309]
	IGP48	The percentage of FA2[3]G1 glycan in total neutral IgG glycans (GPn)	AE	0.776 [0.793;0.761]	0.776 [0.747;0.805]	NS	NS	0.224 [0.195;0.253]
	IGP56	The percentage of monogalactosylated structures in total neutral IgG glycans	ADE	0.678 [0.492;0.749]	0.475 [0.277;0.674]	0.203 [0;0.411]	NS	0.322 [0.281;0.363]
	IGP58	The percentage of all fucosylated (+/- bisecting GlcNAc) structures in total neutral IgG glycans	DCE	0.57 [0.599;0.55]	NS	0.57 [0.494;0.646]	0.162 [0.098;0.226]	0.268 [0.232;0.303]
	IGP59	The percentage of fucosylation of agalactosylated structures	ADE	0.735 [0.591;0.791]	0.333 [0.136;0.531]	0.491 [0.107;0.805]	NS	0.265 [0.231;0.3]
	IGP60	The percentage of fucosylation of monogalactosylated structures	AE	0.714 [0.731;0.7]	0.714 [0.677;0.751]	NS	NS	0.286 [0.249;0.323]
	IGP62	The percentage of fucosylated (without bisecting GlcNAc) structures in total neutral IgG glycans	AE	0.76 [0.777;0.745]	0.76 [0.729;0.79]	NS	NS	0.24 [0.21;0.271]
	IGP63	The percentage of fucosylation (without bisecting GlcNAc) of agalactosylated structures	AE	0.737 [0.754;0.722]	0.737 [0.703;0.771]	NS	NS	0.263 [0.229;0.297]
immune cell traits**	P4-3242	16+56/314+335+158b-	ACE	0.427 [0.20;0.66]	0.427 [0.20;0.66]	NS	0.356[0.14;0.54]	0.217 [0.16;0.31]
	P4-2561	16+56/314+335+158b-R7-	ACE	0.424 [0.19;0.66]	0.424 [0.19;0.66]	NS	0.358 [0.14;0.54]	0.218 [0.16;0.31]
	P4-3161	16+56/314+335+158a-158b-	AE	0.792 [0.71;0.81]	0.792 [0.71;0.81]	NS	NS	0.208 [0.15;0.29]
	P4-2488	16+56/314+335+158a-158b-R7-	AE	0.792 [0.71;0.81]	0.792 [0.71;0.81]	NS	NS	0.208 [0.15;0.29]
	P4-3215	16+56/314+335+337-158b-	ACE	0.422 [0.19;0.66]	0.422 [0.19;0.66]	NS	0.358 [0.14;0.54]	0.220[0.16;0.31]
	P4-2534	16+56/314+335+337-158b-R7-	ACE	0.423 [0.19;0.66]	0.423 [0.19;0.66]	NS	0.356 [0.14;0.54]	0.221[0.16;0.31]
	P4-2558	16+56/335+158b-R7-	ACE	0.439 [0.21;0.68]	0.439 [0.21;0.68]	NS	0.344 [0.12;0.53]	0.217[0.16;0.31]
	P4-3239	16+56/335+158b-	ACE	0.437 [0.20;0.68]	0.437 [0.20;0.68]	NS	0.344 [0.12;0.53]	0.220[0.16;0.31]
	P4-3134	16+56/314+335+337-158a-158b-	AE	0.791 [0.71;0.85]	0.792 [0.71;0.85]	NS	NS	0.209 [0.15;0.29]
	P4-2462	16+56/314+335+337-158b-R7-	AE	0.783 [0.70;0.84]	0.783 [0.70;0.84]	NS	NS	0.217 [0.16;0.30]
	P4-2531	16+56/335+337-158b-R7-	ACE	0.433 [0.20;0.67]	0.433 [0.20;0.67]	NS	0.347 [0.13;0.53]	0.217[0.16;0.31]
	P4-3212	16+56/335+337-158b-	ACE	0.428 [0.19;0.67]	0.428 [0.19;0.67]	NS	0.347[0.12;0.53]	0.225 [0.16;0.32]
	P4-3158	16+56/335+158a-158b-	AE	0.792 [0.71;0.81]	0.792 [0.71;0.81]	NS	NS	0.208 [0.15;0.29]
	P4-2460	16+56/335+337-158a-158b-R7-	AE	0.782 [0.70;0.84]	0.782 [0.70;0.84]	NS	NS	0.218 [0.16;0.30]
	P4-2485	16+56/335+158a-158b-R7-	AE	0.783 [0.70;0.84]	0.783 [0.70;0.84]	NS	NS	0.217 [0.16;0.30]
	P4-3131	16+56/335+337-158a-158b-	AE	0.781 [0.70;0.84]	0.781 [0.70;0.84]	NS	NS	0.219 [0.16;0.30]
	P4-3401	16+56/335+158a-	ACE	0.475 [0.26;0.70]	0.475 [0.26;0.70]	NS	0.337 [0.12;0.51]	0.188[0.13;0.27]
	P4-2740	16+56/335-337-R7-	ACE	0.349 [0.15;0.55]	0.349 [0.15;0.55]	NS	0.461 [0.27;0.62]	0.19 [0.14;0.27]
	P4-3437	16+56/335-337-	ACE	0.303 [0.11;0.50]	0.303 [0.11;0.50]	NS	0.509 [0.33;0.66]	0.188 [0.13;0.27]
	P4-2767	16+56/335-R7-	ACE	0.352 [0.15;0.56]	0.352 [0.15;0.56]	NS	0.46 [0.27;0.62]	0.188 [0.13;0.27]
	P4-3482	16+56/335+	ACE	0.303 [0.11;0.50]	0.303 [0.11;0.50]	NS	0.51 [0.33;0.66]	0.187[0.13;0.27]
	P4-3464	16+56/335-	ACE	0.305 [0.11;0.50]	0.305 [0.11;0.50]	NS	0.508 [0.33;0.66]	0.187[0.13;0.27]
	P4-3374	16+56/335+337-158a-	ACE	0.475 [0.26;0.70]	0.475 [0.26;0.70]	NS	0.336 [0.12;0.51]	0.188[0.14;0.27]
	P4-2677	16+56/335+337-158a-R7-	ACE	0.472 [0.26;0.70]	0.472 [0.26;0.70]	NS	0.339 [0.13;0.51]	0.189 [0.14;0.27]
	P4-3455	16+56/335+337-	ACE	0.309 [0.12;0.50]	0.309 [0.12;0.50]	NS	0.505 [0.33;0.65]	0.186 [0.13;0.27]
	P4-2758	16+56/335+337-R7-	ACE	0.306 [0.11;0.50]	0.309 [0.12;0.50]	NS	0.506 [0.33;0.66]	0.187 [0.13;0.27]
	P4-3243	16+56/CD2+314+335+158b-	ACE	0.461 [0.23;0.71]	0.461 [0.23;0.71]	NS	0.322 [0.10;0.51]	0.217 [0.16;0.31]
	P4-2562	16+56/CD2+314+335+158b-R7-	ACE	0.465 [0.23;0.71]	0.465 [0.23;0.71]	NS	0.32 [0.09;0.51]	0.215 [0.15;0.30]
	P4-3162	16+56/CD2+314+335+158a-158b-	AE	0.788 [0.71;0.84]	0.788 [0.71;0.84]	NS	NS	0.212 [0.16;0.29]
	P4-2489	16+56/CD2+314+335+158a-158b-R7-	AE	0.79 [0.71;0.85]	0.79 [0.71;0.85]	NS	NS	0.21 [0.15;0.29]
	P4-3216	16+56/CD2+314+335+337-158b-	ACE	0.459 [0.22;0.70]	0.459 [0.22;0.70]	NS	0.323 [0.10;0.51]	0.218 [0.16;0.31]
	P4-2535	16+56/CD2+314+335+337-158b-R7-	ACE	0.46 [0.23;0.70]	0.46 [0.23;0.70]	NS	0.324 [0.10;0.51]	0.216 [0.16;0.31]
	P4-18	16+56/CD2+314+335+337-158a-158b-R7-	AE	0.788 [0.71;0.84]	0.788 [0.71;0.84]	NS	NS	0.212 [0.16;0.29]
	P4-2486	16+56/CD2+335+158a-158b-R7-	AE	0.782 [0.70;0.84]	0.782 [0.70;0.84]	NS	NS	0.218 [0.16;0.30]
	P4-2559	16+56/CD2+335+158b-R7-	ACE	0.471 [0.24;0.72]	0.471 [0.24;0.72]	NS	0.312 [0.08;0.50]	0.217 [0.16;0.31]
	P4-3159	16+56/CD2+335+158a-158b-	AE	0.78 [0.70;0.84]	0.78 [0.70;0.84]	NS	NS	0.22 [0.16;0.30]
	P4-3240	16+56/CD2+335+158b-	ACE	0.466 [0.23;0.71]	0.466 [0.23;0.71]	NS	0.314 [0.09;0.51]	0.219 [0.15;0.30]
	P4-3135	16+56/CD2+314+335+337-158a-158b-	AE	0.786 [0.71;0.84]	0.786 [0.71;0.84]	NS	NS	0.214 [0.16;0.29]
	P4-2532	16+56/CD2+335+337-158b-R7-	ACE	0.47 [0.24;0.72]	0.47 [0.24;0.72]	NS	0.313 [0.08;0.50]	0.217 [0.16;0.31]
	P4-3213	16+56/CD2+335+337-158b-	ACE	0.467 [0.23;0.71]	0.467 [0.23;0.71]	NS	0.314 [0.08;0.51]	0.22 [0.16;0.31]
	P4-3132	16+56/CD2+335+337-158a-158b-	AE	0.78 [0.70;0.84]	0.78 [0.70;0.84]	NS	NS	0.22 [0.16;0.30]
	P4-2461	16+56/CD2+335+337-158a-158b-R7-	AE	0.781 [0.70;0.84]	0.781 [0.70;0.84]	NS	NS	0.219 [0.16;0.30]
	P4-2967	16+56/335-337-158b+R7-	AE	0.772 [0.68;0.84]	0.772 [0.68;0.84]	NS	NS	0.228 [0.16;0.32]
	P4-2994	16+56/335-158b+R7-	AE	0.772 [0.68;0.84]	0.772 [0.68;0.84]	NS	NS	0.228 [0.16;0.32]
	P4-3707	16+56/335-158b+	AE	0.775 [0.68;0.84]	0.775 [0.68;0.84]	NS	NS	0.225 [0.16;0.32]
	P4-3680	16+56/335-337-158b+	AE	0.775 [0.68;0.84]	0.775 [0.68;0.84]	NS	NS	0.225 [0.16;0.32]
	P4-2970	16+56/314+335-337-158b+R7-	AE	0.775 [0.68;0.84]	0.775 [0.68;0.84]	NS	NS	0.225 [0.16;0.32]
	P4-3683	16+56/314+335-337-158b+	AE	0.776 [0.68;0.84]	0.776 [0.68;0.84]	NS	NS	0.224 [0.16;0.31]
	P4-2997	16+56/314+335-158b+R7-	AE	0.775 [0.68;0.84]	0.775 [0.68;0.84]	NS	NS	0.225 [0.16;0.32]
	P4-3710	16+56/314+335-158b+	AE	0.777 [0.69;0.84]	0.777 [0.69;0.84]	NS	NS	0.223 [0.16;0.31]
	P4-2900	16+56/335-337-158a-158b+R7-	AE	0.756 [0.66;0.82]	0.756 [0.66;0.82]	NS	NS	0.244 [0.18;0.34]

Table S9: Heritability of 1,113 proteins in the TwinsUK cohort

h2= heritability; CI_2.5=confidence interval at 2.5; CI_97.5=confidence interval at 97.5; A=additive genetic variance; D= dominance genetic variance; C= common environmental variance; E= unique environmental variance

SomaID	Target	UniProt	Entrez Gene ID	Entrez Gene Symbol	Units	SomaQC	Associated with AITD	Associated with one of 17 AITD-IgG N-glycan structures	best model	h2 estimated	h2 CI_2.50	h2 CI_97.50	A estimation	A CI_2.50	A CI_97.50	D estimation	D CI_2.50	D CI_97.50	C estimation	C CI_2.50	C CI_97.50	E estimation	E CI_2.50	E CI_97.50
SL000125	IL-1a	P01583	3552	IL1A	RFU		Yes	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL000589	TSH	P01215	P011081	TSHB	RFU		Yes	No	DE	0.356	0.229	0.402	0	0	0	0.356	0.122	0.591	0	0	0	0.644	0.409	0.878
SL003710	Caspase-2	P42575	835	CASP2	RFU		Yes	No	DE	0.470	0.443	0.479	0	0	0	0.470	0.238	0.701	0	0	0	0.530	0.299	0.762
SL002283	b2-Microglobulin	P01769	567	B2M	RFU		No	Yes	CE	0	0	0	0	0	0	0	0	0	0.509	0.356	0.662	0.491	0.338	0.644
SL002644	ERBB1	P00533	1956	EGFR	RFU		No	Yes	CE	0	0	0	0	0	0	0	0	0	0.389	0.208	0.569	0.611	0.431	0.792
SL008609	FCG3B	O75015	2215	FCGR3B	RFU		No	Yes	AE	0.479	0.468	0.485	0.479	0.303	0.655	0	0	0	0	0	0	0.521	0.345	0.697
SL000002	VEGF	P15692	7422	VEGFA	RFU		No	No	DE	0.500	0.499	0.500	0	0	0	0.500	0.302	0.697	0	0	0	0.500	0.303	0.698
SL000003	Angiogenin	P03950	283	ANG	RFU		No	No	AE	0.718	0.782	0.677	0.718	0.603	0.832	0	0	0	0	0	0	0.282	0.168	0.397
SL000004	bFGF	P09038	2247	FGF2	RFU		No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL000006	PAI-1	P05121	5054	SERPINE1	RFU		No	No	CE	0	0	0	0	0	0	0	0	0	0.497	0.343	0.650	0.503	0.350	0.657
SL000007	ER	P03372	2099	ESR1	RFU		No	No	E	0	0	0	0	0	0	0.742	0.613	0.872	0	0	0	0.258	0.128	0.387
SL000009	ERBB2	P04626	2064	ERBB2	RFU		No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL000017	vWF	P04275	7450	VWF	RFU		No	No	AE	0.700	0.776	0.657	0.700	0.563	0.838	0	0	0	0	0	0	0.300	0.162	0.437
SL000019	Apo A-I	P02647	335	APOA1	RFU	Calibration Scale	No	No	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
SL000020	Apo B	P04114	338	APOB	RFU		No	No	CE	0	0	0	0	0	0	0	0	0	0.617	0.491	0.744	0.383	0.256	0.509
SL000021	Insulin	P01308	3630	INS	RFU		No	No	CE	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL000022	D-dimer	P02671	P022243	2244	FGA FGB FGG	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.279	0.086	0.471	0.721	0.529	0.914
SL000024	TF	P13726	2152	F3	RFU		No	No	CE	0	0	0	0	0	0	0	0	0	0.383	0.208	0.558	0.617	0.442	0.792
SL000027	COX-2	P35354	5743	PTGS2	RFU		No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL000038	MCP-1	P13500	6347	CCL2	RFU		No	No	CE	0	0	0	0	0	0	0	0	0	0.220	0.026	0.414	0.780	0.586	0.974
SL000039	IL-8	P10145	3576	IL8	RFU		No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL000045	IGFBP-3	P17936	3486	IGFBP3	RFU		No	No	CE	0	0	0	0	0	0	0	0	0	0.423	0.253	0.594	0.577	0.406	0.747
SL000047	IGF-I	P05019	3479	IGF1	RFU		No	No	DE	0.296	0.114	0.361	0	0	0	0.296	0.060	0.532	0	0	0	0.704	0.468	0.940
SL000048	Protein C	P04070	5624	PROC	RFU		No	No	AE	0.444	0.405	0.460	0.444	0.238	0.650	0	0	0	0	0	0	0.556	0.350	0.762
SL000049	Protein S	P07225	5627	PROS1	RFU		No	No	E	0.296	0.145	0.357	0.296	0.083	0.509	0	0	0	0	0	0	0.704	0.491	0.917
SL000051	CRP	P02741	1401	CRP	RFU		No	No	AE	0.413	0.358	0.437	0.413	0.219	0.606	0	0	0	0	0	0	0.587	0.394	0.781
SL000053	IPA	P00750	5327	PLAT	RFU		No	No	AE	0.531	0.550	0.522	0.531	0.342	0.720	0	0	0	0	0	0	0.469	0.280	0.658
SL000055	Cadherin E	P12830	999	CDH1	RFU		No	No	CE	0	0	0	0	0	0	0	0	0	0.452	0.284	0.621	0.548	0.379	0.716
SL000057	Thymidine kase	P04183	7083	TK1	RFU		No	No	CE	0	0	0	0	0	0	0	0	0	0.275	0.079	0.471	0.725	0.529	0.921
SL000062	PSA	P07288	354	KLK3	RFU		No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL000064	Kallikrein 7	P49862	5650	KLK7	RFU		No	No	CE	0	0	0	0	0	0	0	0	0	0.573	0.435	0.711	0.427	0.289	0.565
SL000070	Glypican 3	P51854	2719	GPC3	RFU		No	No	DE	0	0	0	0	0	0	0.581	0.411	0.752	0	0	0	0.419	0.248	0.589
SL000076	p27Kip1	P46527	1027	CDKN1B	RFU		No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL000087	IL-6	P05231	3569	IL6	RFU		No	No	CE	0	0	0	0	0	0	0	0	0	0.270	0.081	0.460	0.730	0.540	0.919
SL000088	TGF-b2	P61812	7042	TGFB2	RFU		No	No	DE	0.437	0.364	0.459	0	0	0	0.437	0.168	0.706	0	0	0	0.563	0.294	0.832
SL000089	TGF-b3	P10600	7043	TGFB3	RFU		No	No	DE	0.250	-0.083	0.341	0	0	0	0.250	-0.036	0.536	0	0	0	0.750	0.464	1.036
SL000091	PSA-ACT	P07288	P0354	12	KLK3 SERP3	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.323	0.135	0.510	0.677	0.490	0.865
SL000104	Bcl-2	P10415	596	BCL2	RFU		No	No	AE	0.190	-0.116	0.293	0.190	-0.058	0.439	0	0	0	0	0	0	0.810	0.561	1.058
SL000124	MMP-2	P08253	4313	MMP2	RFU		No	No	AE	0.593	0.640	0.570	0.593	0.425	0.762	0	0	0	0	0	0	0.407	0.238	0.575
SL000130	Cyclin B1	P14635	891	CCNB1	RFU		No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL000131	PC	P12004	5111	PC	RFU		No	No	CE	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL000133	MIP-3a	P78556	6364	CCL20	RFU		No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL000134	Met	P08581	4233	MET	RFU		No	No	AE	0.563	0.596	0.547	0.563	0.395	0.732	0	0	0	0	0	0	0.437	0.268	0.605
SL000136	AREG	P15514	374	AREG	RFU		No	No	CE	0	0	0	0	0	0	0	0	0	0.713	0.612	0.813	0.287	0.187	0.388
SL000138	HB-EGF	Q99075	1839	HBEFG	RFU		No	No	E	0.587	0.632	0.565	0.587	0.416	0.758	0	0	0	0	0	0	0.413	0.242	0.584
SL000139	EPI	O14944	2069	EREG	RFU		No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL000142	TS	P04818	7298	TYMS	RFU		No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL000158	PSMA	Q04609	2346	FOLH1	RFU		No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL000164	Myoglobin	P02144	4151	MB	RFU		No	No	AE	0.542	0.564	0.531	0.542	0.370	0.714	0	0	0	0	0	0	0.458	0.286	0.630
SL000247	6-Phosphogluconate dehydratase	P52209	5226	PGD	RFU		No	No	AE	0.817	0.878	0.773	0.817	0.737	0.898	0	0	0	0	0	0	0.183	0.102	0.263
SL000248	a1-Antitrypsin	P01011	12	ALB	RFU		No	No	CE	0	0	0	0	0	0	0	0	0	0.278	0.083	0.473	0.722	0.527	0.917
SL000249	a1-Antitrypsin	P01009	5265	SERP11	RFU		No	No	CE	0	0	0	0	0	0	0	0	0	0.346	0.165	0.527	0.654	0.473	0.835
SL000250	a2-Antiplasmin	P08697	5345	SERPINF2	RFU		No	No	DE	0.679	0.795	0.628	0	0	0	0.679	0.482	0.876	0	0	0	0.321	0.124	0.518
SL000251	a2-HS-Glycoprotein	P02765	197	AHSG	RFU		No	No	AE	0.442	0.401	0.459	0.442	0.235	0.649	0	0	0	0	0	0	0.558	0.351	0.765
SL000252	a2-Macroglobulin	P01023	2	A2M	RFU		No	No	CE	0	0	0	0	0	0	0	0	0	0.602	0.471	0.733	0.398	0.267	0.529
SL000254	Albumin	P02768	213	ALB	RFU		No	No	CE	0	0	0	0	0	0	0	0	0	0.562	0.419	0.704	0.438	0.296	0.581
SL000268	Angiostatin	P00747	5340	PLG	RFU		No	No	AE	0.733	0.795	0.693	0.733	0.628	0.838	0	0	0	0	0	0	0.267	0.162	0.372
SL000271	Angiotensinogen	P01019	183	AGT	RFU		No	No	CE	0	0	0	0	0	0	0	0	0	0.395	0.220	0.571	0.605	0.429	0.780
SL000272	Antithrombin III	P01008	462	SERPINC1	RFU		No	No	CE	0	0	0	0	0	0	0	0	0	0.350	0.149	0.552	0.650	0.448	0.851
SL000276	Apo E	P02649	348	APOE	RFU	Calibration Scale	No	No	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
SL000277	Apo E2	P02649	348	APOE	RFU		No	No	CE	0	0	0	0	0	0	0	0	0	0.675	0.564	0.787	0.325	0.213	0.436
SL000280	GOT1	P17174	2805	GOT1	RFU		No	No	DE	0	0	0	0	0	0	0.588	0.412	0.763	0	0	0	0.412	0.237	0.588
SL000299	b-ECGF	P05230	2246	FGF1	RFU		No	No	E	0														

SL000343	Cathepsin B	P07858	1508	CTSB	RFU	No	No	AE	0.494	0.491	0.496	0.494	0.321	0.668	0	0	0	0	0	0	0.506	0.332	0.679
SL000344	Cathepsin D	P07339	1509	CTSD	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL000345	Cathepsin G	P08311	1511	CTSG	RFU	No	No	DE	0.415	0.139	0.452	0	0	0	0.415	0.033	0.797	0	0	0.585	0.203	0.967	
SL000346	Cathepsin H	P09688	1512	CTSH	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.557	0.414	0.443	0.299	0.586	
SL000347	CEBG	P08185	865	SERP6	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL000357	Coagulation Factor IX	P00740	2158	F9	RFU	No	No	FE	0.515	0.524	0.511	0.515	0.327	0.704	0	0	0	0	0	0.485	0.296	0.673	
SL000358	Coagulation Factor VII	P08709	2155	F7	RFU	No	No	DE	0.730	0.803	0.686	0	0	0	0.730	0.610	0.850	0	0	0.270	0.150	0.390	
SL000360	Coagulation Factor X	P00742	2159	F10	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.396	0.218	0.573	0.604	0.427	
SL000377	CK-BB	P12277	1152	CKB	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL000382	CK-MB	P12277	P061152 1158	CKB CKM	RFU	No	No	AE	0.550	0.577	0.537	0.550	0.376	0.725	0	0	0	0	0.450	0.275	0.624		
SL000383	CK-MM	P06732	1158	CKM	RFU	No	No	AE	0.546	0.573	0.534	0.546	0.364	0.728	0	0	0	0	0.454	0.272	0.636		
SL000384	CTLA-4	P16410	1493	CTLA4	RFU	No	No	DE	0.463	0.377	0.478	0	0	0	0.463	0.114	0.812	0	0	0.537	0.188	0.886	
SL000396	Cytochrome c	P99999	54205	CYCS	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL000398	Cytochrome P450 3A4	P08684	1576	CYP3A4	RFU	No	No	AE	0.721	0.789	0.679	0.721	0.604	0.839	0	0	0	0	0.279	0.161	0.396		
SL000401	Elastase	P08246	1991	ELANE	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL000403	Endostatin	P39060	80781	COL18A1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.687	0.578	0.797	0.313	0.203	
SL000406	Eotaxin	P51671	6356	CCL11	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.275	0.085	0.465	0.725	0.535	
SL000408	Epo	P01588	2056	EPO	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL000409	ERK-1	P27361	5595	MAPK3	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.513	0.363	0.863	0.487	0.337	
SL000414	Factor B	P00751	629	CFB	RFU	No	No	DE	0.405	0.325	0.435	0	0	0	0.405	0.177	0.633	0	0	0.595	0.367	0.823	
SL000415	Factor H	P08603	3075	CFH	RFU	No	No	DE	0.371	0.252	0.413	0	0	0	0.371	0.131	0.612	0	0	0.629	0.388	0.869	
SL000420	Ferritin	P02794	P022495 2512	FTH1 FTL	RFU	No	No	AE	0.623	0.673	0.595	0.623	0.479	0.768	0	0	0	0	0.377	0.232	0.521		
SL000424	Fibrinogen	P02671	P022243 2244	FGB FGB FGG	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.226	0.024	0.428	0.774	0.572	
SL000426	Fibronectin	P02751	2335	FN1	RFU	No	No	AE	0.627	0.683	0.598	0.627	0.476	0.779	0	0	0	0	0.373	0.221	0.524		
SL000427	Fractalkine/CX3CL-1	P78423	6376	CX3CL1	RFU	No	No	AE	0.405	0.349	0.430	0.405	0.220	0.589	0	0	0	0	0.595	0.411	0.780		
SL000428	FSH	P01215	P01081 2488	CGA FSHB	RFU	No	No	DE	0.298	0.157	0.357	0	0	0	0.298	0.157	0.357	0	0	0.702	0.497	0.908	
SL000433	Glucagon	P01275	2641	GCG	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.539	0.394	0.683	0.461	0.317	
SL000437	Haptoglobin Mixed Ty	P00738	3240	HP	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.370	0.195	0.546	0.630	0.454	
SL000440	Hemopexin	P02790	3263	HPX	RFU	No	No	AE	0.905	0.944	0.872	0.905	0.860	0.949	0	0	0	0	0.095	0.051	0.140		
SL000441	HGF	P14210	3082	HGF	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL000445	HIV-2 Rev	P18093	1724716	Human-virus	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL000449	HSP 40	P25685	3337	DBJ1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.211	0.013	0.408	0.789	0.592	
SL000450	HSP 60	P10809	3329	HSPD1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.546	0.403	0.689	0.454	0.311	
SL000451	HSP 70	P08107	3303	HSPA1A	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.345	0.163	0.527	0.655	0.473	
SL000456	IC3b	P01024	718	CS	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.496	0.339	0.653	0.504	0.347	
SL000458	IFN-g R1	P15260	3459	IFNGR1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL000460	IGD	P01880	3495 5080	IGHD IGK@ IGL@RFU	RFU	No	No	DE	0.454	0.423	0.467	0	0	0	0.454	0.256	0.652	0	0	0.546	0.348	0.744	
SL000461	IGe	P01854	3497 5080	IGHGe IGK@ IGL@RFU	RFU	No	No	AE	0.557	0.586	0.542	0.557	0.384	0.729	0	0	0	0	0.443	0.271	0.616		
SL000462	IGFBP-1	P08833	3484	IGFBP1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.567	0.429	0.705	0.433	0.295	
SL000466	IGFBP-2	P18065	3485	IGFBP2	RFU	No	No	AE	0.527	0.541	0.520	0.527	0.352	0.701	0	0	0	0	0.473	0.299	0.648		
SL000467	IGg	P01857	3500 3501	IGHG1 IGHG2 IGRF@	RFU	No	No	DE	0.502	0.503	0.501	0	0	0	0.502	0.316	0.688	0	0	0.498	0.312	0.684	
SL000468	IgM	P01871	3507 3512	IGHM IGJ IGK@ IGRF@	RFU	No	No	AE	0.432	0.392	0.451	0.432	0.247	0.617	0	0	0	0	0.568	0.383	0.753		
SL000470	IL-11	P20809	3589	IL11	RFU	No	No	DE	0.344	0.196	0.395	0	0	0	0.344	0.101	0.587	0	0	0.656	0.413	0.899	
SL000474	IL-16	Q14005	3603	IL16	RFU	No	No	AE	0.658	0.723	0.622	0.658	0.512	0.804	0	0	0	0	0.342	0.196	0.488		
SL000478	IL-2	P00568	3558	IL2	RFU	No	No	CE	0	0	0	0	0	0	0.584	0.449	0.719	0	0.416	0.281	0.551		
SL000479	IL-3	P08700	3562	IL3	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL000480	IL-4	P05112	3565	IL4	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL000481	IL-5	P05113	3567	IL5	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.416	0.245	0.587	0.584	0.413	
SL000483	IL-7	P13232	3574	IL7	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL000493	LDH-H 1	P07195	3945	LDHB	RFU	No	No	AE	0.709	0.772	0.669	0.709	0.593	0.824	0	0	0	0	0.291	0.176	0.407		
SL000496	Lactoferrin	P02788	4057	LTF	RFU	No	No	DE	0.605	0.650	0.581	0	0	0	0.605	0.455	0.755	0	0	0.395	0.245	0.545	
SL000497	Laminin	P25391	P01284217 39	LAMA1 LAMB1	RFU	No	No	AE	0.616	0.668	0.589	0.616	0.461	0.771	0	0	0	0	0.384	0.229	0.539		
SL000498	Leptin	P41159	3952	LEP	RFU	No	No	AE	0.622	0.680	0.593	0.622	0.462	0.783	0	0	0	0	0.378	0.217	0.538		
SL000506	Luteinizing hormone	P01215	P01081 3972	GGA LHB	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.305	0.119	0.490	0.695	0.510	
SL000507	Lymphotoxin a1/b2	P01374	Q014049 4050	LTA LTB	RFU	No	No	DE	0.538	0.568	0.526	0	0	0	0.538	0.317	0.759	0	0	0.462	0.241	0.683	
SL000508	Lymphotoxin a2/b1	P01374	Q014049 4050	LTA LTB	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.296	0.103	0.489	0.704	0.511	
SL000509	Lymphotoxin b R	P36941	4055	LTBR	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.274	0.080	0.468	0.726	0.532	
SL000510	Lysozyme	P61626	4069	LYZ	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.682	0.570	0.795	0.318	0.205	
SL000515	MCP-2	P80075	6355	CCL8	RFU	No	No	DE	0.388	0.300	0.422	0	0	0	0.388	0.168	0.608	0	0	0.612	0.392	0.832	
SL000516	MCP-3	P80098	6354	CCL7	RFU	No	No	DE	0.760	0.833	0.713	0	0	0	0.760	0.650	0.870	0	0	0.240	0.130	0.350	
SL000517	MCP-4	Q98616	6357	CCL13	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.172	-0.027	0.372	0.828	0.628	
SL000519	MIP-1a	P01047	6348	CCL3	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.154	-0.045	0.354	0.846	0.646	
SL000521	MMP-1	P03956	4312	MMP1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.367	0.191	0.543	0.633	0.457	
SL000522	MMP-12	P39900	4321	MMP12	RFU	No	No	AE	0.595	0.639	0.572	0.595	0.435	0.755	0	0	0	0	0	0.405	0.245	0.565	
SL000523	MMP-13	P45452	4322	MMP13	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL000524	MMP-3	P08254	4314	MMP3	RFU	No	No	AE	0.203	-0.032	0.294	0.203	-0.018	0.424	0	0	0	0	0	0.797	0.576	1.018	
SL000525	MMP-7	P09237	4316	MMP7	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.570	0.431	0.708	0.430	0.292	
SL000526	MMP-8	P22894	4317	MMP8	RFU	No	No	CE	0	0	0	0											

SL000573		P02743	325	APCS	RFU	No	No	AE	0.475	0.459	0.482	0.475	0.284	0.666	0	0	0	0	0	0	0.525	0.334	0.716
SL000581	SOD	P00441	6647	SOD1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.249	0.056	0.441	0.751	0.559	0.944
SL000582	Survivin	O15392	332	BIRC5	RFU	No	No	DE	0.759	0.852	0.705	0	0	0	0.759	0.627	0.891	0	0	0	0.241	0.109	0.373
SL000584	TGF-b1	P01137	7040	TGFB1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.227	0.032	0.423	0.773	0.577	0.968
SL000586	Thrombin	P00734	2147	F2	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.596	0.465	0.728	0.404	0.272	0.535
SL000587	Thyroglobulin	P01266	7038	TG	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL000588	TMA	P07202	7173	TPO	RFU	No	No	DE	0.513	0.537	0.508	0	0	0	0.513	0.180	0.845	0	0	0	0.487	0.155	0.820
SL000590	Thyroxine-Binding Glo	P05543	6906	SERP17	RFU	No	No	DE	0.556	0.585	0.542	0	0	0	0.556	0.388	0.725	0	0	0	0.444	0.275	0.612
SL000591	TIMP-1	P01033	7076	TIMP1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.261	0.065	0.457	0.739	0.543	0.935
SL000592	TIMP-2	P16035	7077	TIMP2	RFU	No	No	AE	0.472	0.453	0.480	0.472	0.273	0.671	0	0	0	0	0	0	0.528	0.329	0.727
SL000597	TNF-b	P01374	4049	LTA	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL000601	Transferrin	P02787	7018	TF	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.260	0.062	0.457	0.740	0.543	0.938
SL000603	Trypsin	P07477	5644	PRSS1	RFU	No	No	AE	0.600	0.649	0.575	0.600	0.434	0.765	0	0	0	0	0	0	0.400	0.235	0.566
SL000605	Ubiquitin+1	P62979	6233	RPS27A	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.449	0.284	0.614	0.551	0.386	0.716
SL000613	uPA	P00749	5328	PLAU	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.478	0.319	0.637	0.522	0.363	0.681
SL000615	Vasoaactive Intestil Pep	P01282	7432	VIP	RFU	No	No	DE	0.546	0.575	0.533	0	0	0	0.546	0.349	0.743	0	0	0	0.454	0.257	0.651
SL000617	ALT	P24298	2875	GPT	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.340	0.158	0.522	0.660	0.478	0.842
SL000622	Coagulation Factor V	P12259	2153	F5	RFU	No	No	AE	0.701	0.764	0.662	0.701	0.581	0.821	0	0	0	0	0	0	0.299	0.179	0.419
SL000633	Fas ligand soluble	P48023	356	FASLG	RFU	No	No	DE	0.394	0.320	0.425	0	0	0	0.394	0.189	0.599	0	0	0	0.606	0.401	0.811
SL000638	Cadherin-2	P19022	1000	CDH2	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL000640	Nidogen	P14543	4811	NID1	RFU	No	No	AE	0.678	0.755	0.637	0.678	0.528	0.828	0	0	0	0	0	0	0.322	0.172	0.472
SL000645	MMP-10	P09238	4319	MMP10	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL000655	Keratin 18	P05783	3875	KRT18	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL000658	GAS1	P54826	2619	GAS1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL000668	GSTA6 ANTIGEN	P10671	948	CD36	RFU	No	No	DE	0.584	0.629	0.562	0	0	0	0.584	0.408	0.759	0	0	0	0.416	0.241	0.592
SL000670	GSTA3	Q16772	2940	GSTA3	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL000674	FST	P19883	10468	FST	RFU	No	No	AE	0.213	-0.141	0.315	0.213	-0.063	0.488	0	0	0	0	0	0	0.787	0.512	1.063
SL000678	Granulysin	P22749	10578	GNLY	RFU	No	No	AE	0.679	0.740	0.643	0.679	0.552	0.806	0	0	0	0	0	0	0.321	0.194	0.448
SL000695	Lipocalin 2	P80188	3934	LCN2	RFU	No	No	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
SL000836	Hemoglobin	P69905	P613039	3043 HBA1 HBB	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.397	0.225	0.569	0.603	0.431	0.775
SL001691	FGF7	P21781	2252	FGF7	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.602	0.470	0.733	0.398	0.267	0.530
SL001713	IL-17	Q16552	3605	IL17A	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.241	0.044	0.437	0.759	0.563	0.956
SL001716	IL-12	P29459	P23592	3593 IL12A IL12B	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL001717	IL-10	P22301	3586	IL10	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL001718	IL-13	P35225	3596	IL13	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL001720	VCAM-1	P19320	7412	VCAM1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.388	0.212	0.564	0.612	0.436	0.788
SL001721	PECAM-1	P16284	5175	PECAM1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL001726	GM-CSF	P04141	1437	CSF2	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL001729	G-CSF	P09919	1440	CSF3	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.223	0.029	0.416	0.777	0.584	0.971
SL001737	STRATIFIN	P31947	2810	SFN	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.475	0.317	0.634	0.525	0.366	0.683
SL001753	Sialoadhesin	Q9BZ22	6614	SIGLEC1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL001766	HCG	P01215	P011081	1082 CGA CGB	RFU	No	No	DE	0.502	0.504	0.502	0	0	0	0.502	0.311	0.694	0	0	0	0.498	0.306	0.689
SL001774	FABP	P05413	2170	FABP3	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.422	0.253	0.591	0.578	0.409	0.747
SL001777	Cystatin C	P01034	1471	CST3	RFU	No	No	DE	0.593	0.635	0.571	0	0	0	0.593	0.439	0.747	0	0	0	0.407	0.253	0.561
SL001795	IL-1b	P01584	3553	IL1B	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.169	-0.029	0.367	0.181	0.533	1.029
SL001796	Myoeloperoxidase	P05164	4353	MPO	RFU	No	No	AE	0.600	0.644	0.576	0.600	0.447	0.753	0	0	0	0	0	0	0.400	0.247	0.553
SL001797	Kallikrein 6	Q92876	5653	KLK6	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.202	0.002	0.402	0.798	0.598	0.998
SL001800	TNF sR-II	P20333	7133	TNFRSF1B	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.493	0.337	0.650	0.507	0.350	0.663
SL001802	IFN-g	P01579	3458	IFNG	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.697	0.592	0.802	0.303	0.198	0.408
SL001815	Mn SOD	P04179	6648	SOD2	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.488	0.330	0.646	0.512	0.354	0.670
SL001868	CASA	P47710	1446	CSN1S1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL001888	SLPI	P03973	6590	SLPI	RFU	No	No	AE	0.529	0.543	0.521	0.529	0.362	0.695	0	0	0	0	0	0	0.471	0.305	0.638
SL001890	GA733-1 protein	P09758	4070	TACSTD2	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL001896	Clusterin	P10909	1191	CLU	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.363	0.184	0.542	0.637	0.458	0.816
SL001897	SPINT2	Q43291	10653	SPINT2	RFU	No	No	AE	0.685	0.751	0.646	0.685	0.553	0.816	0	0	0	0	0	0	0.315	0.184	0.447
SL001902	BCAM	P50895	4059	BCAM	RFU	No	No	AE	0.523	0.543	0.516	0.523	0.290	0.757	0	0	0	0	0	0	0.477	0.243	0.710
SL001905	Mesothelin	Q13421	10232	MSLN	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL001938	Activin A	P08476	3624	INHBA	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.318	0.134	0.501	0.682	0.499	0.866
SL001943	IL-6 sRa	P08887	3570	IL6R	RFU	No	No	AE	0.845	0.899	0.804	0.845	0.777	0.913	0	0	0	0	0	0	0.155	0.087	0.223
SL001945	sE-Selectin	P16581	6401	SELE	RFU	No	No	AE	0.723	0.792	0.680	0.723	0.605	0.841	0	0	0	0	0	0	0.277	0.159	0.395
SL001947	MIA	Q16674	8190	MIA	RFU	No	No	DE	0.669	0.730	0.634	0	0	0	0.669	0.538	0.801	0	0	0	0.331	0.199	0.462
SL001973	Mammaglobin 2	O75556	4246	SCGB2A1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL001992	TNF sR-I	P19438	7132	TNFRSF1A	RFU	No	No	AE	0.502	0.504	0.502	0.502	0.313	0.692	0	0	0	0	0	0	0.498	0.308	0.687
SL001995	Angiopietin-1	Q15389	284	ANGPT1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.293	0.107	0.480	0.707	0.520	0.893
SL001996	Angiopietin-2	Q15123	285	ANGPT2	RFU	No	No	AE	0.666	0.726	0.632	0.666	0.535	0.798	0	0	0	0	0	0	0.334	0.202	0.465
SL001997	IL-1 sRI	P14778	3554	IL1R1	RFU	No	No	AE	0.539	0.560	0.529	0.539	0.362	0.715	0	0	0	0	0	0	0.461	0.285	0.638
SL001998	TFPI	P10646	7035	TFPI	RFU	No	No	CE															

SL002662	Coagulation Factor XI	P03951	2160	F11	RFU	No	No	AE	0.514	0.523	0.511	0.514	0.334	0.695	0	0	0	0	0	0	0.486	0.305	0.666
SL002684	CSF-1	P09603	1435	CSF1	RFU	No	No	DE	0.434	0.335	0.459	0	0	0	0.434	0.133	0.736	0	0	0	0.566	0.264	0.867
SL002695	Glutamate carboxypep	Q96KP4	55748	CNDP2	RFU	No	No	DE	0.534	0.566	0.523	0	0	0	0.534	0.291	0.777	0	0	0	0.466	0.223	0.709
SL002702	PM1	P11309	5292	PM1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
SL002704	PTN	P21246	5764	PTN	RFU	No	No	DE	0.336	0.185	0.389	0	0	0	0.336	0.086	0.576	0	0	0	0.664	0.424	0.904
SL002705	Thrombospondin-1	P07966	7057	THBS1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.369	0.193	0.545	0.631	0.455	0.807
SL002706	CD23	P06734	2208	FCER2	RFU	No	No	AE	0.521	0.533	0.516	0.521	0.346	0.696	0	0	0	0	0	0	0.479	0.304	0.654
SL002755	PAPP-A	Q13219	5069	PAPPA	RFU	No	No	DE	0.724	0.792	0.681	0	0	0	0.724	0.607	0.841	0	0	0	0.276	0.159	0.393
SL002756	hnRNP K	P61978	3190	HLRNPK	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL002763	Kallikrein 11	Q9UBX7	11012	KLK11	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.595	0.462	0.728	0.405	0.272	0.538
SL002783	Cardiotrophin-1	Q16619	1489	CTF1	RFU	No	No	DE	0.366	0.194	0.414	0	0	0	0.366	0.085	0.647	0	0	0	0.634	0.353	0.915
SL002792	BARK1	P25098	156	ADRBK1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.595	0.463	0.727	0.405	0.273	0.537
SL002803	PGP9.5	P09936	7345	UCHL1	RFU	No	No	AE	0.742	0.818	0.695	0.742	0.623	0.861	0	0	0	0	0	0	0.258	0.139	0.377
SL002823	sl-Selectin	P14151	6402	SELL	RFU	No	No	DE	0.301	0.136	0.363	0	0	0	0.301	0.074	0.528	0	0	0	0.699	0.472	0.926
SL002922	sICAM-1	P05362	3383	ICAM1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.573	0.435	0.710	0.427	0.290	0.565
SL003041	PF-4	P02776	5196	PF4	RFU	No	No	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
SL003043	TIMP-3	P35625	7078	TIMP3	RFU	No	No	AE	0.569	0.603	0.552	0.569	0.407	0.732	0	0	0	0	0	0	0.431	0.268	0.593
SL003060	bFGF-R	P11362	2260	FGFR1	RFU	No	No	DE	0.524	0.538	0.518	0	0	0	0.524	0.342	0.707	0	0	0	0.476	0.293	0.658
SL003080	MIF	P14174	4282	MIF	RFU	No	No	AE	0.621	0.678	0.591	0.621	0.460	0.782	0	0	0	0	0	0	0.379	0.218	0.540
SL003104	Eotaxin-2	O00175	6369	CCL24	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL003166	ALCAM	Q13740	214	ALCAM	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.446	0.274	0.618	0.554	0.382	0.726
SL003167	BLC	O43927	10563	CXCL13	RFU	No	No	AE	0.380	0.291	0.416	0.380	0.166	0.595	0	0	0	0	0	0	0.620	0.405	0.834
SL003168	CTACK	Q8Y4X3	10850	CCL27	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.170	-0.035	0.376	0.830	0.624	1.035
SL003169	E-78	P42830	6374	CXCL5	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.266	0.071	0.461	0.734	0.539	0.929
SL003171	FGF-4	P09620	2249	FGF4	RFU	No	No	DE	0.540	0.579	0.527	0	0	0	0.540	0.295	0.785	0	0	0	0.460	0.215	0.705
SL003172	GCP-2	P80162	6372	CXCL6	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.417	0.249	0.586	0.583	0.414	0.751
SL003173	Gro-a	P09341	2919	CXCL1	RFU	No	No	AE	0.568	0.606	0.550	0.568	0.388	0.748	0	0	0	0	0	0	0.432	0.252	0.612
SL003176	I-309	P22362	6346	CCL1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.279	0.087	0.472	0.721	0.528	0.913
SL003177	sICAM-2	P13598	3384	ICAM2	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.162	-0.045	0.368	0.838	0.632	1.045
SL003178	sICAM-3	P32942	3385	ICAM3	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL003179	Integrin a1b1	P56199	P013672 3688	ITGA1 ITGB1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.529	0.383	0.676	0.471	0.324	0.617
SL003182	Integrin aVb5	P06756	P113685 3693	ITGA5 ITGB5	RFU	No	No	DE	0.510	0.517	0.507	0	0	0	0.510	0.300	0.719	0	0	0	0.490	0.281	0.700
SL003183	IP-10	P02778	3627	CXCL10	RFU	No	No	AE	0.528	0.541	0.521	0.528	0.366	0.690	0	0	0	0	0	0	0.472	0.310	0.634
SL003184	sLeptin R	P48357	3953	LEPR	RFU	No	No	AE	0.958	0.976	0.941	0.958	0.939	0.977	0	0	0	0	0	0	0.042	0.023	0.061
SL003186	Lymphotactin	P47982	6375	XCL1	RFU	No	No	DE	0.764	0.992	0.74	0	0	0	0.607	0.387	0.828	0	0	0	0.393	0.172	0.613
SL003187	MDC	O00626	6387	CCL22	RFU	No	No	DE	0.661	0.725	0.625	0	0	0	0.661	0.519	0.803	0	0	0	0.359	0.197	0.481
SL003189	MIP-3b	Q09731	6363	CCL19	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL003190	MIP-5	Q16663	6359	CCL15	RFU	No	No	DE	0.636	0.692	0.605	0	0	0	0.636	0.490	0.782	0	0	0	0.364	0.218	0.510
SL003191	P-2	P02775	5473	PPBP	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.345	0.165	0.524	0.655	0.476	0.835
SL003192	Properdin	P27918	5199	CFP	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.420	0.249	0.590	0.580	0.410	0.751
SL003193	6CKine	O00585	6366	CCL21	RFU	No	No	AE	0.701	0.771	0.660	0.701	0.573	0.829	0	0	0	0	0	0	0.299	0.171	0.427
SL003196	TARC	Q92583	6361	CCL17	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.412	0.242	0.582	0.588	0.418	0.758
SL003197	TECK	O15444	6370	CCL25	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.632	0.510	0.755	0.368	0.245	0.490
SL003198	Tesin	P24821	3371	TNC	RFU	No	No	AE	0.806	0.869	0.762	0.806	0.721	0.891	0	0	0	0	0	0	0.194	0.109	0.279
SL003199	sTie-1	P35590	7075	TIE1	RFU	No	No	DE	0.843	0.843	0.715	0	0	0	0.764	0.650	0.879	0	0	0	0.236	0.121	0.350
SL003200	sTie-2	Q02763	7010	TEK	RFU	No	No	AE	0.538	0.556	0.528	0.538	0.361	0.714	0	0	0	0	0	0	0.462	0.286	0.639
SL003201	VEGF sR2	P35968	3791	KDR	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.560	0.415	0.704	0.440	0.296	0.585
SL003220	C3adesArg	P01024	718	C3	RFU	No	No	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
SL003280	HMG-1	P09429	3146	HMGB1	RFU	No	No	DE	0.750	0.833	0.700	0	0	0	0.750	0.624	0.875	0	0	0	0.250	0.125	0.376
SL003300	HCC-4	O15467	6360	CCL16	RFU	No	No	DCE	0.165	-0.077	0.249	0	0	0	0.165	-0.040	0.370	0.646	0.451	0.841	0.189	0.104	0.273
SL003301	Ck-b-8-1	P55773	6368	CCL23	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.303	0.116	0.490	0.697	0.510	0.884
SL003302	MPHF-1	P55773	6368	CCL23	RFU	No	No	AE	0.749	0.822	0.703	0.749	0.636	0.862	0	0	0	0	0	0	0.251	0.138	0.364
SL003303	CCL28	Q8NRJ3	56477	CCL28	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.360	0.182	0.538	0.640	0.462	0.818
SL003304	IGF-I sR	P08069	3480	IGF1R	RFU	No	No	DE	0.529	0.546	0.521	0	0	0	0.529	0.342	0.716	0	0	0	0.471	0.284	0.658
SL003305	IL-2 sRn	P01589	3559	IL2RA	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL003307	IL-2 sRg	P31785	3561	IL2RG	RFU	No	No	ACE	0.423	0.209	0.438	0.423	0.028	0.819	0	0	0	0.371	-0.011	0.752	0.206	0.116	0.297
SL003308	IL-4 sR	P24394	3566	IL4R	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL003309	LBP	P18428	3929	LBP	RFU	No	No	AE	0.603	0.647	0.579	0.603	0.451	0.755	0	0	0	0	0	0	0.397	0.245	0.549
SL003310	VEGF121	P15692	7422	VEGFA	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.156	-0.044	0.355	0.844	0.645	1.044
SL003322	VEGF sR3	P35916	2324	FLT4	RFU	No	No	DE	0.849	0.905	0.807	0	0	0	0.849	0.780	0.918	0	0	0	0.151	0.082	0.220
SL003323	PARC	P55774	6362	CCL18	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.641	0.521	0.762	0.359	0.238	0.479
SL003324	Coagulation Factor Xa	P00742	2159	F10	RFU	No	No	DE	0.623	0.685	0.592	0	0	0	0.623	0.455	0.791	0	0	0	0.377	0.209	0.545
SL003326	I-TAC	O14625	6373	CXCL11	RFU	No	No	DE	0.525	0.537	0.519	0	0	0	0.525	0.358	0.691	0	0	0	0.475	0.309	0.642
SL003327	Factor D	P00746	1675	CFD	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
SL003328	Factor I	P05156	3426	CFI	RFU	No	No	AE	0.461	0.437	0.												

[illegible]

SL004327	BAFF	Q9Y275	10673	TNFSF13B	RFU	No	No	CE	0	0	0	0	0	0	0	0.299	0.110	0.487	0.701	0.513	0.890		
SL004329	BMP-14	P43026	8200	GDF5	RFU	No	No	DE	0.445	0.370	0.465	0	0	0	0.445	0.158	0.731	0.555	0.269	0.842			
SL004330	CD22	P20273	933	CD22	RFU	No	No	E	0	0	0	0	0	0	0	0	0	1	1	1			
SL004331	CNTF	P26441	1270	CNTF	RFU	No	No	DE	0.499	0.498	0.500	0	0	0	0.499	0.182	0.816	0	0.501	0.184	0.818		
SL004332	EG-VEGF	P58284	84432	PDCK1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	1	1	1			
SL004333	FGF-10	O15620	2255	FGF10	RFU	No	No	E	0	0	0	0	0	0	0	0	0	1	1	1			
SL004334	FGF-16	O43320	8823	FGF16	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.585	0.450	0.719	0.415	0.281	0.550
SL004335	FGF-17	O60258	8822	FGF17	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1		
SL004336	FGF-18	O76093	8817	FGF18	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1		
SL004337	FGF-19	O95750	9965	FGF19	RFU	No	No	AE	0.554	0.583	0.540	0.554	0.381	0.727	0	0	0	0.446	0.273	0.619			
SL004338	FGF-20	Q9NP95	26281	FGF20	RFU	No	No	AE	0.707	0.789	0.661	0.707	0.565	0.849	0	0	0	0.293	0.151	0.435			
SL004339	FGF-5	P12034	2250	FGF5	RFU	No	No	E	0	0	0	0	0	0	0	0	0	1	1	1			
SL004340	FGF-6	P10767	2251	FGF6	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.401	0.228	0.573	0.599	0.427	0.772
SL004342	FGF-8B	P55075	2253	FGF8	RFU	No	No	E	0	0	0	0	0	0	0	0	0	1	1	1			
SL004343	FG ligand	P49771	2323	FLT3LG	RFU	No	No	E	0	0	0	0	0	0	0	0	0	1	1	1			
SL004345	GDF-11	O95390	10220	GDF11	RFU	No	No	E	0	0	0	0	0	0	0	0	0	1	1	1			
SL004346	IL-20	Q9NYY1	50604	IL20	RFU	No	No	E	0	0	0	0	0	0	0	0	0	1	1	1			
SL004347	IL-22	Q9GZX6	50616	IL22	RFU	No	No	DE	0.474	0.451	0.482	0	0	0	0.474	0.239	0.708	0	0.526	0.292	0.761		
SL004348	IFN-lambda 1	Q8UJ54	282618	IL29	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.529	0.381	0.677	0.471	0.323	0.619
SL004349	IFN-lambda 2	Q8LZJ0	282616	IL28A	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1		
SL004350	IL-17B	Q9UHF5	27190	IL17B	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1		
SL004351	IL-17E	Q9H293	64806	IL25	RFU	No	No	DE	0.434	0.348	0.458	0	0	0	0.434	0.150	0.719	0	0.566	0.281	0.850		
SL004352	IL-17F	Q96PD4	112744	IL17F	RFU	No	No	DE	0.628	0.726	0.589	0	0	0	0.628	0.410	0.845	0	0.372	0.155	0.590		
SL004353	IL-17D	Q8TAD2	53342	IL17D	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1		
SL004354	IL-19	Q9UJH0	29949	IL19	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.543	0.396	0.690	0.457	0.310	0.644
SL004355	LD78-beta	P16619	414062	CCL3L1	RFU	No	No	DE	0.595	0.679	0.565	0	0	0	0.595	0.360	0.830	0	0.405	0.170	0.640		
SL004356	LAG-1	Q8NHV4	388372	CCL4L1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.188	-0.009	0.384	0.812	0.616	1.009
SL004359	Neurotrophin-3	P20783	4908	NTF3	RFU	No	No	DE	0.640	0.725	0.602	0	0	0	0.640	0.451	0.829	0	0.360	0.171	0.549		
SL004360	Neurotrophin-5	P34130	4909	NTF4	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1		
SL004362	SCGF-beta	Q9Y240	6320	CLEC11A	RFU	No	No	AE	0.613	0.661	0.587	0.613	0.464	0.763	0	0	0	0.387	0.237	0.536			
SL004363	SCGF-alpha	Q9Y240	6320	CLEC11A	RFU	No	No	AE	0.737	0.805	0.693	0.737	0.625	0.849	0	0	0	0.263	0.151	0.375			
SL004364	TACI	O14836	23495	TNFRSF13B	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.246	0.047	0.445	0.754	0.555	0.953
SL004365	TWEAK	O43508	8742	TNFSF12	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1		
SL004366	TWEAKR	Q8NP84	51330	TNFRSF12A	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1		
SL004367	TKK1	O94907	22943	DKK1	RFU	No	No	AE	0.547	0.577	0.534	0.547	0.355	0.739	0	0	0	0.453	0.261	0.645			
SL004400	Coagulation Factor IXa	P00740	2158	CE9	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0.438	0.267	0.609	0.562	0.391	0.733
SL004415	ACE2	Q9BYF1	59272	ACE2	RFU	No	No	E	0	0	0	0	0	0	0	0	0	1	1	1			
SL004438	Cystatin M	Q15828	1474	CST6	RFU	No	No	AE	0.755	0.825	0.710	0.755	0.647	0.862	0	0	0	0.245	0.138	0.353			
SL004457	Protease nexin I	P07093	5270	SERPINE2	RFU	No	No	DE	0.325	0.171	0.381	0	0	0	0.325	0.091	0.559	0	0.675	0.441	0.909		
SL004458	Elafin	P19957	5266	PI3	RFU	No	No	AE	0.476	0.460	0.482	0.476	0.280	0.671	0	0	0	0.524	0.329	0.720			
SL004466	Heparin cofactor II	P05546	3053	SERPIND1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.458	0.293	0.623	0.542	0.377	0.707
SL004469	amyloid precursor prot	P05067	351	APP	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.388	0.214	0.561	0.612	0.439	0.786
SL004477	calgranulin B	P06702	6280	S100A9	RFU	No	No	DE	0.463	0.440	0.474	0	0	0	0.463	0.267	0.660	0	0.537	0.340	0.733		
SL004482	Endoglin	P17813	2022	ENG	RFU	No	No	AE	0.485	0.476	0.489	0.485	0.294	0.676	0	0	0	0.515	0.324	0.706			
SL004484	IL-1 R AcP	P35247	6441	SFRP3	RFU	No	No	DE	0.597	0.669	0.568	0	0	0	0.597	0.384	0.810	0.403	0.190	0.616			
SL004486	VEGF-C	P49787	7424	VEGFC	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.229	0.032	0.425	0.771	0.575	0.968
SL004492	TLR2	O60603	7097	TLR2	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.332	0.147	0.516	0.668	0.484	0.853
SL004511	BPI	P17213	671	BPI	RFU	No	No	DE	0.612	0.663	0.585	0	0	0	0.612	0.456	0.768	0	0.388	0.232	0.544		
SL004515	PGRP-S	O75594	8993	PGLYRP1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.313	0.130	0.497	0.687	0.503	0.870
SL004516	MBL	P11226	4153	MBL2	RFU	No	No	AE	0.813	0.873	0.770	0.813	0.733	0.893	0	0	0	0.187	0.107	0.267			
SL004536	LEAP-1	P81172	57817	HAMP	RFU	No	No	DE	0.224	-0.004	0.310	0	0	0	0.224	-0.002	0.450	0	0.776	0.550	1.002		
SL004556	DAF	P08174	1604	CD55	RFU	No	No	AE	0.590	0.628	0.569	0.590	0.441	0.739	0	0	0	0.410	0.261	0.559			
SL004579	Macrophage mannose	P22897	4360	MRC1	RFU	No	No	AE	0.784	0.852	0.738	0.784	0.687	0.881	0	0	0	0.216	0.119	0.313			
SL004580	Macrophage scavenger	P21757	4481	MSR1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1		
SL004588	IL-1 R AcP	Q9NPH3	3556	IL1RAP	RFU	No	No	AE	0.806	0.866	0.762	0.806	0.723	0.888	0	0	0	0.194	0.112	0.277			
SL004589	Azuricidin	P20160	566	AZU1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.542	0.397	0.687	0.458	0.313	0.603
SL004591	G-CSF-R	Q99062	1441	CSF3R	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.171	-0.027	0.369	0.829	0.631	1.027
SL004605	40S ribosomal protein	P08865	3921	RPSA	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1		
SL004610	LRP8	Q14114	7804	LRP8	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.414	0.243	0.586	0.586	0.414	0.757
SL004625	ADAMTS-4	O75173	9507	ADAMTS4	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.351	0.166	0.535	0.649	0.465	0.834
SL004626	ADAMTS-5	Q8U0	11096	ADAMTS5	RFU	No	No	DE	0.832	0.901	0.783	0	0	0	0.832	0.746	0.918	0	0.168	0.082	0.254		
SL004635	CD30 Ligand	P32971	944	TNFSF8	RFU	No	No	AE	0.690	0.760	0.649	0.690	0.555	0.825	0	0	0	0.310	0.175	0.445			
SL004636	Flt-3	P36888	2322	FLT3	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.625	0.500	0.750	0.375	0.250	0.500
SL004637	MSP R	Q04912	4486	MST1R	RFU	No	No	DE	0.482	0.443	0.490	0	0	0	0.482	0.138	0.827	0	0.518	0.173	0.862		
SL004639	Tnfr	Q16288	4916	TNFRK3	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.498	0.343	0.653	0.502	0.347	0.657
SL004642	ADAM 9	Q13443	8754	ADAM9	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1		
SL004643	Angiotensin-4	Q9Y264	51378	ANGPT4	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.361	0.182	0.540	0.639	0.460	0.818
SL004644	EDA	Q92838	1896	EDA	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1		
SL004645	HAL-1	Q43278	6692	SPINT1	RFU	No	No	DE	0.539	0.564	0.529	0	0	0	0.539	0.350	0.729	0	0.461	0.271	0.650		
SL004646	Layilin	Q6UX15	143903	LAYN	RFU	No	No	AE	0.337	0.168	0.392	0.337	0.083	0.591									

SL004708	CTAP-III	P02775	5473	PPBP	RFU	No	No	CE	0	0	0	0	0	0	0	0	0.367	0.191	0.543	0.633	0.457	0.809	
SL004712	SDF-1	P48061	6387	CXCL12	RFU	No	No	CE	0	0	0	0	0	0	0	0	0.409	0.233	0.584	0.591	0.416	0.767	
SL004714	LIF sR	P42702	3977	LIFR	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL004716	JNK2	P45984	5601	MAPK9	RFU	No	No	DE	0.696	0.784	0.650	0	0	0	0.696	0.541	0.851	0	0	0	0.304	0.149	0.459
SL004718	Karyopherin-α2	P52292	3838	KP2	RFU	No	No	AE	0.251	-0.009	0.335	0.251	-0.005	0.506	0	0	0	0	0	0	0.749	0.494	1.005
SL004720	Calcineurin B a	P63068	5534	PPP3R1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL004723	HDAC8	Q9BY41	55869	HDAC8	RFU	No	No	AE	0.518	0.530	0.513	0.518	0.323	0.713	0	0	0	0	0	0	0.482	0.287	0.677
SL004724	MOZ	Q92794	7994	KAT6A	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL004725	Hat1	O14929	8520	HAT1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL004726	CD97	P48960	976	CD97	RFU	No	No	DE	0.245	-0.121	0.340	0	0	0	0.245	-0.050	0.540	0	0	0	0.755	0.460	1.050
SL004737	Tropomyosin 1 alpha c	P09493	7168	TPM1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL004739	ITI heavy chain H4	Q14624	3700	ITI4	RFU	No	No	AE	0.779	0.844	0.735	0.779	0.685	0.873	0	0	0	0	0	0	0.221	0.127	0.315
SL004742	Aflamin	P43652	173	AFM	RFU	No	No	E	0.463	0.437	0.474	0.463	0.259	0.667	0	0	0	0	0	0	0.537	0.333	0.741
SL004750	DEAD-box protein 19E	Q8UMR2	11269	DDX19B	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
SL004751	HO-2	P30519	3163	HMOX2	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL004752	DRR1	Q95990	11170	FAM107A	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL004757	DRG-1	Q9NP79	51534	VT41	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0	0	0.569	0.431	0.707	
SL004759	eIF-5	P55010	1983	EIF5	RFU	No	No	DE	0.822	0.894	0.773	0	0	0	0.822	0.731	0.913	0	0	0	0.178	0.087	0.269
SL004760	PAFAH beta subunit	P68402	5049	PAFAH1B2	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0	0.642	0.521	0.763		
SL004765	MAPKAPK3	Q16644	7867	MAPKAPK3	RFU	No	No	AE	0.734	0.806	0.690	0.734	0.618	0.851	0	0	0	0	0	0	0.266	0.149	0.382
SL004768	AIF1	P55008	199	AIF1	RFU	No	No	AE	0.387	0.301	0.421	0.387	0.172	0.602	0	0	0	0	0	0	0.613	0.398	0.828
SL004771	Aurora kise A	O14965	6790	AURKA	RFU	No	No	DE	0.500	0.500	0.500	0	0	0	0.500	0.260	0.739	0	0	0	0.500	0.261	0.740
SL004781	CSK	P41240	1445	CSK	RFU	No	No	E	0.753	0.817	0.710	0.753	0.651	0.855	0	0	0	0	0	0	0.247	0.145	0.349
SL004782	TSG-6	P98066	7130	TNFAIP6	RFU	No	No	AE	0.656	0.714	0.623	0.656	0.522	0.791	0	0	0	0	0	0	0.344	0.209	0.478
SL004791	DR3	Q93038	8718	TNFRSF25	RFU	No	No	E	0.880	0.927	0.843	0	0	0	0.880	0.826	0.935	0	0	0	0.120	0.065	0.174
SL004795	ERAB	Q99714	3028	HSD17B10	RFU	No	No	DCE	0.230	-0.053	0.303	0	0	0	0.230	-0.022	0.481	0.539	0.305	0.772	0.232	0.129	0.335
SL004804	Nectin-like protein 1	Q8N126	57863	CADM3	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL004805	Nectin-like protein 2	Q9BY67	23705	CADM1	RFU	No	No	DE	0.383	0.278	0.421	0	0	0	0.383	0.146	0.620	0	0	0	0.617	0.380	0.854
SL004812	Triosephosphate isom	P60174	7167	TP1	RFU	No	No	ACE	0.374	-0.437	0.412	0.374	-0.039	0.787	0	0	0	0.371	-0.014	0.756	0.256	0.144	0.368
SL004814	Coactosin-like protein	Q14019	23406	COTL1	RFU	No	No	DE	0.545	0.587	0.530	0	0	0	0.545	0.304	0.787	0	0	0	0.455	0.213	0.696
SL004820	Phosphoglycerate mut	P18669	5223	PGAM1	RFU	No	No	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
SL004823	Cyclophilin A	P62937	5478	PIPA	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.601	0.470	0.732	0.399	0.268	0.530
SL004837	Actinin AB	P08476	P093624 3625	INHBA INHBB	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.389	0.215	0.563	0.611	0.437	0.785
SL004844	EphA5	P54756	2044	EPHA5	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL004845	EphB4	P54760	2050	EPHB4	RFU	No	No	DE	0.289	0.146	0.350	0	0	0	0.289	0.087	0.492	0	0	0	0.711	0.508	0.913
SL004849	IL-1 sR9	Q9NP60	26280	IL1RAPL2	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL004850	IL-17 sR	Q96F46	23765	IL17RA	RFU	No	No	DE	0.936	0.962	0.912	0	0	0	0.936	0.907	0.965	0	0	0	0.064	0.035	0.093
SL004851	ALK-1	P37023	94	ACVRL1	RFU	No	No	DE	0.563	0.668	0.539	0	0	0	0.563	0.252	0.875	0	0	0	0.437	0.125	0.748
SL004852	B7-H1	Q9NZ07	29126	CD274	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.152	-0.047	0.351	0.848	0.649	1.047
SL004853	B7-H2	O75144	23308	ICOSLG	RFU	No	No	AE	0.472	0.455	0.480	0.472	0.279	0.665	0	0	0	0	0	0	0.528	0.335	0.721
SL004855	contactin-1	Q12860	1272	CNTN1	RFU	No	No	AE	0.464	0.439	0.475	0.464	0.256	0.672	0	0	0	0	0	0	0.536	0.328	0.744
SL004856	Desmoglein-1	Q02413	1828	DSG1	RFU	No	No	AE	0.262	-0.182	0.356	0.262	-0.064	0.587	0	0	0	0	0	0	0.738	0.413	1.064
SL004857	Desmoglein-2	Q14126	1829	DSG2	RFU	No	No	DE	0.618	0.673	0.590	0	0	0	0.618	0.461	0.776	0	0	0	0.382	0.224	0.539
SL004858	GFR-1	P56159	2674	GFR1	RFU	No	No	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
SL004859	GTR	Q9V5U5	8784	TNFRSF18	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL004860	HTRA2	Q43464	27429	HTRA2	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.371	0.194	0.548	0.629	0.452	0.806
SL004861	IL-18 Rb	Q9S256	8807	IL18RAP	RFU	No	No	DE	0.794	0.880	0.740	0	0	0	0.794	0.680	0.908	0	0	0	0.206	0.092	0.320
SL004862	PD-12	Q9BQ51	80380	PDCD1LG2	RFU	No	No	DE	0.584	0.639	0.560	0	0	0	0.584	0.385	0.783	0	0	0	0.416	0.217	0.615
SL004863	TAJ	Q9NS68	55504	TNFRSF19	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL004864	Cadherin-12	P55289	1010	CDH12	RFU	No	No	DE	0.300	-0.176	0.383	0	0	0	0.300	-0.052	0.653	0	0	0	0.700	0.347	1.052
SL004865	Cadherin-6	P55285	1004	CDH6	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL004866	Carbonic anhydrase I	P00915	759	CA1	RFU	No	No	AE	0.551	0.578	0.538	0.551	0.379	0.723	0	0	0	0	0	0	0.449	0.277	0.621
SL004867	Carbonic anhydrase III	P07451	761	CA3	RFU	No	No	AE	0.591	0.633	0.569	0.591	0.433	0.748	0	0	0	0	0	0	0.409	0.252	0.567
SL004868	Carbonic anhydrase V	P43166	766	CA7	RFU	No	No	DE	0.389	0.111	0.435	0	0	0	0.389	0.032	0.746	0	0	0	0.811	0.254	0.968
SL004869	Carbonic anhydrase X	Q8N101	377677	CA13	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.498	0.344	0.651	0.502	0.349	0.656
SL004871	DR6	Q75509	27242	TNFRSF21	RFU	No	No	DE	0	0	0	0	0	0	0	0	0	0.538	0.390	0.686	0.462	0.314	0.610
SL004872	EDAR	Q9JNE0	10913	EDAR	RFU	No	No	AE	0.765	0.829	0.722	0.765	0.668	0.862	0	0	0	0	0	0	0.235	0.138	0.332
SL004875	IL-1Rrp2	Q9HB29	8808	IL1RL2	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.286	0.094	0.478	0.714	0.522	0.906
SL004876	Kallistatin	P29622	5267	SERP4	RFU	No	No	K	0	0	0	0	0	0	0	0	0	0.487	0.325	0.650	0.513	0.350	0.675
SL004891	hnRNP A2/B1	P22626	3181	HNRNPA2B1	RFU	No	No	DE	0.723	0.809	0.674	0	0	0	0.723	0.583	0.862	0	0	0	0.277	0.138	0.417
SL004899	HSP70 protein 8	P11142	3312	HSPA8	RFU	No	No	DE	0.548	0.613	0.530	0	0	0	0.548	0.261	0.835	0	0	0	0.452	0.165	0.739
SL004901	Protein disulfide-isom	P07237	5034	P4HB	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL004908	Tropomyosin 2	P07951	7169	TPM2	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL004914	PPase	Q15181	5464	PPA1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL004915	CDC27	Q00289	1192	CLC1	RFU	No	No	AE	0.745	0.809	0.702	0.745	0.640	0.849	0	0	0	0	0	0	0.255	0.151	0.360
SL004919	Peroxioredoxin-1	Q06830	5052	PRDX1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.517	0.367	0.667	0.483	0.333	0.633
SL0049																							

[illegible]

SL006119	TFF3	Q07654	7033	TFF3	RFU	No	No	CE	0	0	0	0	0	0	0	0.184	-0.015	0.383	0.816	0.617	1.015	
SL006132	Lamin-B1	P20700	4001	LMNB1	RFU	No	No	DE	0.546	0.599	0.530	0	0	0	0.546	0.278	0.814	0.454	0.186	0.722		
SL006189	KIF23	Q02241	9493	KIF23	RFU	No	No	DE	0.362	0.171	0.413	0	0	0	0.362	0.071	0.654	0.638	0.346	0.929		
SL006197	DJ homolog	Q86DA6	131118	DJC19	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1		
SL006268	NSF1C	Q9JNZ2	55968	NSFL1C	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.479	0.321	0.636		
SL006372	YES1	P07947	7525	YES1	RFU	No	No	E	0.704	0.770	0.664	0.704	0.581	0.826	0	0	0	0.521	0.364	0.679		
SL006374	BMX	P51813	660	BMX	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0.296	0.174	0.419		
SL006378	Esterase D	P10768	2098	ESD	RFU	No	No	CE	0	0	0	0	0	0	0	0.337	0.155	0.519	0.663	0.481	0.845	
SL006397	NRP1	O14786	8829	NRP1	RFU	No	No	AE	0.335	0.217	0.383	0.335	0.127	0.543	0	0	0	0.665	0.457	0.873		
SL006406	PLXC1	O60486	10154	PLXNC1	RFU	No	No	AE	0.692	0.759	0.653	0.692	0.564	0.821	0	0	0	0.308	0.179	0.436		
SL006448	HRG	P04196	3273	HRG	RFU	No	No	CE	0	0	0	0	0	0	0	0.467	0.306	0.627	0.533	0.373	0.694	
SL006460	GP1BA	P07359	2811	GP1BA	RFU	No	No	E	0	0	0	0	0	0	0	0	0	1	1	1		
SL006476	NMT1	P30419	4836	NMT1	RFU	No	No	CE	0	0	0	0	0	0	0	0.540	0.396	0.685	0.460	0.315	0.604	
SL006480	TRY3	P35030	5646	PRSS3	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	1	
SL006512	HGF	Q04756	3083	HGFAC	RFU	No	No	AE	0.826	0.881	0.785	0.826	0.755	0.898	0	0	0	0.174	0.102	0.245		
SL006522	LG3BP	Q08380	3959	LGALS3BP	RFU	No	No	E	0	0	0	0	0	0	0	0.739	0.646	0.832	0.261	0.168	0.354	
SL006523	MFGM	Q08431	4240	MFGE8	RFU	No	No	CE	0	0	0	0	0	0	0	0.494	0.340	0.649	0.506	0.351	0.660	
SL006528	SEPR	Q12884	2191	FAP	RFU	No	No	CE	0	0	0	0	0	0	0	0.255	0.061	0.450	0.745	0.550	0.939	
SL006542	FCN2	Q15485	2220	FCN2	RFU	No	No	DE	0.707	0.788	0.662	0	0	0	0.707	0.567	0.848	0	0	0.293	0.152	0.433
SL006544	BGH3	Q15582	7045	TGFB1	RFU	No	No	CE	0	0	0	0	0	0	0	0.385	0.208	0.563	0.615	0.437	0.792	
SL006550	ECM1	Q16610	1893	ECM1	RFU	No	No	AE	0.493	0.487	0.495	0.493	0.285	0.700	0	0	0	0.507	0.300	0.715		
SL006610	ATS13	Q76LX8	11093	ADAMTS13	RFU	No	No	CE	0	0	0	0	0	0	0	0.600	0.468	0.732	0.400	0.268	0.532	
SL006629	SIRT2	Q8IKJ6	22933	SIRT2	RFU	No	No	CE	0	0	0	0	0	0	0	0.431	0.265	0.598	0.569	0.402	0.735	
SL006675	CKAP2	Q8VWK9	26586	CKAP2	RFU	No	No	AE	0.286	0.110	0.353	0.286	0.060	0.512	0	0	0	0.714	0.488	0.940		
SL006694	CNDP1	Q09K42	84735	CNDP1	RFU	No	No	AE	0.670	0.730	0.635	0.670	0.540	0.801	0	0	0	0.330	0.199	0.460		
SL006698	transcription factor MLI	Q8N3X6	254251	LCORL	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL006705	PFD5	Q99471	5204	PFDN5	RFU	No	No	CE	0	0	0	0	0	0	0	0.341	0.154	0.528	0.659	0.472	0.846	
SL006713	Collectin Kidney 1	Q9BW98	78989	COLEC11	RFU	No	No	AE	0.621	0.670	0.594	0.621	0.476	0.765	0	0	0	0.379	0.235	0.524		
SL006777	FETUB	Q9UGM5	26998	FETUB	RFU	No	No	AE	0.638	0.690	0.609	0.638	0.502	0.775	0	0	0	0.362	0.225	0.498		
SL006803	ANGL3	Q9Y5C1	27329	ANGPTL3	RFU	No	No	DE	0.919	0.955	0.887	0	0	0	0.919	0.878	0.959	0.081	0.041	0.122		
SL006805	MRCKB	Q9Y552	9578	CDC42BPB	RFU	No	No	E	0	0	0	0	0	0	0	0	0	1	1	1		
SL006830	complement factor H-n	Q98XR6	81494	CFHR5	RFU	No	No	CE	0	0	0	0	0	0	0	0.385	0.210	0.560	0.615	0.440	0.790	
SL006892	ABL1	P00519	25	ABL1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL006910	Cathepsin V	Q60911	1515	CTSL2	RFU	No	No	AE	0.361	0.269	0.400	0.361	0.162	0.559	0	0	0	0.639	0.441	0.838		
SL006911	CHK1	Q14757	1111	CHEK1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL006912	FGF	P09769	2288	FGF	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL006913	FYN	P06241	2534	FYN	RFU	No	No	AE	0.672	0.729	0.638	0.672	0.548	0.797	0	0	0	0.328	0.203	0.452		
SL006914	Glucocorticoid recepto	P04150	2908	NR3C1	RFU	No	No	CE	0	0	0	0	0	0	0	0.363	0.185	0.540	0.637	0.460	0.815	
SL006915	IL-27	Q8NEV9	Q1246778	10 IL27 EB3	RFU	No	No	AE	0.264	-0.018	0.347	0.264	-0.008	0.536	0	0	0	0.736	0.464	1.008		
SL006916	LCK	P06239	3932	LCK	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL006917	LYN	P07948	4067	LYN	RFU	No	No	E	0	0	0	0	0	0	0	0.587	0.454	0.720	0.413	0.280	0.546	
SL006918	MK01	P28482	5594	MAPK1	RFU	No	No	ACE	0.426	0.230	0.440	0.426	0.030	0.821	0	0	0	0.364	-0.016	0.744		
SL006919	RSK-like protein kise	O75582	9252	RPS6KA5	RFU	No	No	CE	0	0	0	0	0	0	0	0.543	0.398	0.687	0.457	0.313	0.602	
SL006920	MAPK14	Q16539	1432	MAPK14	RFU	No	No	CE	0	0	0	0	0	0	0	0.569	0.431	0.707	0.431	0.293	0.569	
SL006921	PRK1	C15118	5163	EPH1	RFU	No	No	DE	0.264	-0.014	0.347	0	0.264	-0.007	0.534	0	0	0	0.736	0.466	1.027	
SL006922	RAD51	Q08609	5888	RAD51	RFU	No	No	AE	0.438	0.403	0.455	0.438	0.255	0.621	0	0	0	0.562	0.379	0.745		
SL006923	TBP	P20226	6908	TBP	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL006924	ART	O00253	181	AGRP	RFU	No	No	DE	0.505	0.509	0.504	0	0	0	0.505	0.308	0.702	0	0.495	0.298	0.692	
SL006970	DLL1	O00548	28514	DLL1	RFU	No	No	AE	0.257	0.054	0.333	0.257	0.030	0.485	0	0	0	0.743	0.515	0.970		
SL006992	MATN3	O15232	4148	MATN3	RFU	No	No	DE	0.553	0.613	0.535	0	0	0	0.553	0.287	0.819	0	0.447	0.181	0.713	
SL006993	MK13	O15264	5603	MAPK13	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL006998	PDPK1	O15530	5170	PDPK1	RFU	No	No	AE	0.730	0.794	0.689	0.730	0.622	0.838	0	0	0	0.270	0.162	0.378		
SL007003	DHH	O43323	50846	DHH	RFU	No	No	AE	0.604	0.649	0.579	0.604	0.452	0.756	0	0	0	0.396	0.244	0.548		
SL007022	HNRPO	O60506	10492	SYNCRIP	RFU	No	No	CE	0	0	0	0	0	0	0	0.259	0.063	0.456	0.741	0.544	0.937	
SL007024	GREM1	O60565	26595	GREM1	RFU	No	No	AE	0.628	0.700	0.594	0.628	0.448	0.808	0	0	0	0.372	0.192	0.552		
SL007025	JA2	O60674	3717	JA2	RFU	No	No	AE	0.779	0.841	0.736	0.779	0.687	0.870	0	0	0	0.221	0.130	0.313		
SL007049	CYT7	O76096	8530	CS7	RFU	No	No	DE	0.721	0.784	0.681	0	0	0	0.721	0.610	0.832	0	0.279	0.168	0.390	
SL007056	BMP10	O95393	27302	BMP10	RFU	No	No	CE	0	0	0	0	0	0	0	0.240	0.046	0.434	0.760	0.566	0.954	
SL007059	LY86	O95711	9450	LY86	RFU	No	No	CE	0	0	0	0	0	0	0	0.211	0.010	0.411	0.789	0.589	0.990	
SL007100	LKHA4	P09960	4048	LTA4H	RFU	No	No	CE	0	0	0	0	0	0	0	0.506	0.354	0.658	0.494	0.342	0.646	
SL007121	CATE	P14091	1510	CTSE	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL007122	IDE	P14735	3416	IDE	RFU	No	No	CE	0	0	0	0	0	0	0	0.182	-0.016	0.379	0.818	0.621	1.016	
SL007145	NR1D1	P20393	9572	NR1D1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL007153	PERL	P22079	4025	LPO	RFU	No	No	AE	0.550	0.579	0.537	0.550	0.366	0.734	0	0	0	0.450	0.266	0.634		
SL007173	GRN	P28799	2896	GRN	RFU	No	No	DE	0.707	0.772	0.667	0	0.707	0.588	0.826	0	0	0	0.293	0.174	0.412	
SL007179	EPHB2	P29323	2048	EPHB2	RFU	No	No	AE	0.533	0.549	0.525	0.533	0.371	0.695	0	0	0	0.467	0.305	0.629		
SL007181	TYK2	P29597	7297	TYK2	RFU	No	No	AE	0.246	0.021	0.327	0.246	0.011	0.481	0	0	0	0.754	0.519	0.989		
SL007195	CD70	P32970	970	CD70	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL007206	TSP2	P35442	7058	THBS2	RFU	No	No	AE	0.496	0.493	0.497	0.496	0.306	0.686	0	0	0	0.504	0.314	0.694		
SL007207	TSP4	P35443	7060	THBS4	RFU	No	No	AE	0.350	0.239	0.395	0.350	0.137	0.564	0	0	0	0.650	0.436	0.863		
SL007228	KPC1	P41743	5584	PRKC1	RFU	No	No	AE	0.756	0.821	0.713	0.756	0.655	0.857	0							

SL007502	ST4S6	Q7LFX5	51363	CHST15	RFU	No	No	AE	0.617	0.674	0.588	0.617	0.452	0.782	0	0	0	0	0	0	0.383	0.218	0.548
SL007531	BMPER	Q8N8U9	168667	BMPER	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1		
SL007547	TIMD3	Q8TDQ0	84868	HAVCR2	RFU	No	No	E	0.422	0.371	0.444	0.422	0.226	0.618	0	0	0	0	0	0.578	0.382	0.774	
SL007560	STAB2	Q8WVQ8	55576	STAB2	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1		
SL007620	IL12RB2	Q98665	3595	IL12RB2	RFU	No	No	E	0.315	0.136	0.376	0	0	0	0.315	0.069	0.560	0	0	0.685	0.440	0.931	
SL007640	CLC7A	Q98XN2	64591	CLEC7A	RFU	No	No	AE	0.707	0.786	0.663	0.707	0.570	0.845	0	0	0	0	0	0.293	0.155	0.430	
SL007642	ANGL4	Q98Y76	51129	ANGPTL4	RFU	No	No	DE	0.335	0.098	0.396	0	0	0	0.335	0.040	0.629	0	0	0.665	0.371	0.960	
SL007651	FGF23	Q9GZV9	8074	FGF23	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL007673	NET4	Q9HB63	59277	NTN4	RFU	No	No	DE	0.440	0.390	0.459	0	0	0	0.440	0.213	0.667	0	0	0.560	0.333	0.787	
SL007674	LY9	Q9HBG7	4063	LY9	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.541	0.397	0.686	0.459	0.314	
SL007680	ROBO2	Q9HC4K	6092	ROBO2	RFU	No	No	AE	0.505	0.508	0.504	0.505	0.314	0.696	0	0	0	0	0	0.495	0.304	0.686	
SL007729	ARTS1	Q9NZ08	51752	ERAP1	RFU	No	No	AE	0.724	0.796	0.681	0.724	0.603	0.846	0	0	0	0	0	0.276	0.154	0.397	
SL007747	TBK1	Q9UHD2	29110	TBK1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.294	0.106	0.482	0.706	0.518	
SL007752	DAPK2	Q9UIK4	23604	DAPK2	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL007756	GDF2	Q9UJK5	2959	GDF2	RFU	No	No	DE	0.675	0.740	0.638	0	0	0	0.675	0.539	0.811	0	0	0.325	0.189	0.461	
SL007774	JAG2	Q9Y219	3714	JAG2	RFU	No	No	DE	0.718	0.825	0.664	0	0	0	0.718	0.553	0.883	0	0	0.282	0.117	0.447	
SL007804	BGN	P21810	633	BGN	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.500	0.347	0.654	0.500	0.346	
SL007806	IL22RA1	Q8N6P7	58985	IL22RA1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL007869	PIIB	P23284	5479	PIIB	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL007871	Cytidylylate kise	P30085	51727	CMPK1	RFU	No	No	AE	0.503	0.505	0.502	0.503	0.325	0.681	0	0	0	0	0	0.497	0.319	0.675	
SL007888	Cystatin-S	P01036	1472	CS4	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL008008	ARGI1	P05089	383	ARG1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL008023	HPLN1	P10915	1404	HAPLN1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL008039	AKI1A1	P14550	10327	AKR1A1	RFU	No	No	AE	0.829	0.884	0.787	0.829	0.756	0.901	0	0	0	0	0	0.171	0.099	0.244	
SL008059	RS3	R23396	6188	RF53	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL008063	PPAC	P24666	52	ACP1	RFU	No	No	AE	0.824	0.882	0.781	0.824	0.748	0.900	0	0	0	0	0	0.178	0.100	0.252	
SL008072	CO8A1	P27658	1295	COL8A1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL008085	3HIDH	P31937	11112	HIBADH	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL008099	CAPG	P40121	822	CAPG	RFU	No	No	AE	0.759	0.820	0.718	0.759	0.665	0.854	0	0	0	0	0	0.241	0.146	0.335	
SL008102	MDHC	P40925	4190	MDH1	RFU	No	No	DE	0.784	0.856	0.736	0	0	0	0.784	0.682	0.886	0	0	0.216	0.114	0.318	
SL008122	DUS3	P51452	1845	DUSP3	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.551	0.409	0.693	0.449	0.307	
SL008143	UBE2N	P61088	7334	UBE2N	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.622	0.497	0.747	0.378	0.253	
SL008157	UBE2L3	P68036	7332	UBE2L3	RFU	No	No	AE	0.756	0.823	0.712	0.756	0.653	0.859	0	0	0	0	0	0.244	0.141	0.347	
SL008176	PSME1	Q06323	5720	PSME1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.467	0.307	0.628	0.533	0.372	
SL008177	C1QB2P	Q07021	708	C1QBP	RFU	No	No	DE	0.360	0.251	0.403	0	0	0	0.360	0.141	0.619	0	0	0.640	0.471	0.659	
SL008178	DERM	Q07507	1805	PPT	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.437	0.269	0.604	0.563	0.396	
SL008190	SPTA2	Q13813	6709	SPTAN1	RFU	No	No	DE	0.479	0.434	0.488	0	0	0	0.479	0.137	0.822	0	0	0.521	0.178	0.863	
SL008193	NID2	Q14112	22795	NID2	RFU	No	No	AE	0.249	0.016	0.331	0.249	0.008	0.490	0	0	0	0	0	0.751	0.510	0.992	
SL008309	RTN4	Q9NQC3	57142	RTN4	RFU	No	No	DE	0.572	0.607	0.555	0	0	0	0.572	0.410	0.735	0	0	0.428	0.265	0.590	
SL008331	PA2G4	Q9UQ80	5036	PA2G4	RFU	No	No	AE	0.719	0.783	0.678	0.719	0.605	0.832	0	0	0	0	0	0.281	0.168	0.395	
SL008378	4EBP2	Q13542	1979	EIF4EBP2	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL008380	CATZ	Q9UBR2	1522	CTSZ	RFU	No	No	AE	0.797	0.863	0.752	0.797	0.707	0.888	0	0	0	0	0	0.203	0.112	0.293	
SL008382	CYTD	P28325	1473	CST5	RFU	No	No	AE	0.605	0.649	0.581	0.605	0.458	0.753	0	0	0	0	0	0.395	0.247	0.542	
SL008414	EphB6	Q15197	2051	EPHB6	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.205	0.003	0.406	0.795	0.594	
SL008416	MRC2	Q9UBG0	9902	MRC2	RFU	No	No	DE	0.516	0.523	0.512	0	0	0	0.516	0.349	0.682	0	0	0.484	0.318	0.651	
SL008421	ATTS1	Q9UJH6	9510	ADAMTS1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL008504	GNS	P15586	2799	GNS	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.163	-0.036	0.362	0.837	0.638	
SL008516	CYTT	P09228	1470	CST2	RFU	No	No	AE	0.639	0.698	0.607	0.639	0.491	0.787	0	0	0	0	0	0.361	0.213	0.509	
SL008574	OMD	Q99983	4958	OMD	RFU	No	No	DE	0.594	0.638	0.572	0	0	0	0.594	0.437	0.752	0	0	0.406	0.248	0.563	
SL008588	SLAF5	Q9UIB8	8832	CD84	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.409	0.235	0.583	0.591	0.417	
SL008590	Olfactomedin-4	Q6UX06	10562	OLFM4	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL008611	ASAH1	Q02083	27163	AA	RFU	No	No	DE	0.654	0.719	0.619	0	0	0	0.654	0.506	0.802	0	0	0.346	0.198	0.494	
SL008623	CNTN2	Q02246	6900	CNTN2	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.520	0.371	0.670	0.480	0.330	
SL008639	IDS	P22304	3423	IDS	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.440	0.275	0.605	0.560	0.395	
SL008644	BST1	C10588	683	BST1	RFU	No	No	AE	0.887	0.931	0.852	0.887	0.837	0.938	0	0	0	0	0	0.113	0.062	0.163	
SL008703	CBPE	P16870	1363	CPE	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0.340	0.158	0.522	0.660	0.478	
SL008709	DSC3	Q14574	1825	DSC3	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL008728	NRX3B	Q9HDB5	9369	NRXN3	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.543	0.397	0.689	0.457	0.311	
SL008759	GPVI	Q9HCN6	51206	GP6	RFU	No	No	DCE	0.271	-0.030	0.334	0	0	0	0.271	-0.010	0.552	0.467	0.209	0.725	0.262	0.146	
SL008773	CD109	Q6YHK3	135228	CD109	RFU	No	No	AE	0.441	0.401	0.458	0.441	0.240	0.642	0	0	0	0	0	0.559	0.358	0.760	
SL008808	SKP1	P63208	6500	SKP1	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.624	0.499	0.749	0.376	0.251	
SL008822	EMR2	Q9UHX3	30817	EMR2	RFU	No	No	CE	0	0	0	0	0	0	0	0	0	0.385	0.211	0.559	0.615	0.441	
SL008835	ASGR1	P07306	432	ASGR1	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL008865	PSA2	P25787	5683	PSMA2	RFU	No	No	DE	0.493	0.488	0.495	0	0	0	0.493	0.291	0.695	0	0	0.507	0.305	0.709	
SL008904	LYVE1	Q8Y6V7	10894	LYVE1	RFU	No	No	DE	0.547	0.575	0.534	0	0	0	0.547	0.360	0.733	0	0	0.453	0.267	0.640	
SL008909	LGMN	Q99538	5641	LGMN	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL008916	DPP2	Q9UJH4	29952	DPP7	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL008933	PARK7	Q99497	11315	PARK7	RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1	1	1	
SL008936	CHL1	Q00533	10752	CHL1	RFU	No	No	AE	0.556	0.584	0.542	0.556	0.387	0.725	0								

SLO10289	Carbonic anhydrase II	P23280	765	CAB	RFLU	No	No	No	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
SLO103228	MED-1	Q15648	5469	MED1	RFLU	No	No	E	0	0	0	0	1	NA	NA	NA	NA	NA	NA	NA
SLO10348	FN1.4	P02751	2335	FN1	RFLU	No	No	DE	0.557	0.586	0.542	0	0	0	0.557	0.385	0.728	0	0	0
SLO10349	FN1.3	P02751	2335	FN1	RFLU	No	No	DE	0.734	0.804	0.691	0	0	0	0.734	0.619	0.849	0	0	0
SLO10368	IDUA	P35475	3425	IDUA	RFLU	No	No	DE	0.722	0.793	0.678	0	0	0	0.722	0.599	0.844	0	0	0
SLO10369	Carbonic Anhydrase I	P22748	762	CA4	RFLU	No	No	CE	0	0	0	0	0	0	0	0	0	0.189	-0.010	0.387
SLO10371	CD39	P49961	953	ENTPD1	RFLU	No	No	DE	0.536	0.582	0.523	0	0	0	0.536	0.256	0.816	0	0	0
SLO10372	Enterokise	P98073	5651	PRSS7	RFLU	No	No	DE	0.714	0.836	0.657	0	0	0	0.714	0.532	0.896	0	0	0
SLO10373	FCAR	P24071	2204	FCAR	RFLU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1
SLO10374	METAP1	P53562	23173	METAP1	RFLU	No	No	AE	0.742	0.816	0.696	0.742	0.625	0.860	0	0	0	0	0	0
SLO10375	ASAH2	Q8NR71	56624	ASAHI	RFLU	No	No	DE	0.697	0.763	0.658	0	0	0	0.697	0.572	0.822	0	0	0
SLO10376	MMEL2	Q495T6	79258	MMEL1	RFLU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1
SLO10378	RET	P07949	5979	RET	RFLU	No	No	AE	0.571	0.606	0.553	0.571	0.407	0.735	0	0	0	0	0	0
SLO10379	Semaphorin 3A	Q14563	10371	SEMA3A	RFLU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1
SLO10381	Soggy-1	Q9UK85	27120	DKKL1	RFLU	No	No	CE	0	0	0	0	0	0	0	0	0	0.407	0.235	0.579
SLO10384	Testican-1	Q08629	6695	SPOCK1	RFLU	No	No	AE	0.709	0.780	0.667	0.709	0.584	0.835	0	0	0	0	0	0
SLO10388	Trypsin B	P07478	5645	PRSS2	RFLU	No	No	AE	0.562	0.598	0.546	0.562	0.379	0.746	0	0	0	0	0	0
SLO10390	URB	Q76M96	151887	CDCD80	RFLU	No	No	DE	0.505	0.508	0.504	0	0	0	0.505	0.327	0.683	0	0	0
SLO10391	WFKN2	Q8TEU8	124857	WFKKN2	RFLU	No	No	AE	0.595	0.638	0.573	0.595	0.441	0.749	0	0	0	0	0	0
SLO10392	GASP-2	I14928	6	SPRASP2	RFLU	No	No	CE	0	0	0	0	0	0	0	0	0.222	0.028	0.417	0.778
SLO10393	KREM2	Q8NCWO	79412	KREMEI	RFLU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1
SLO10449	Carbonic Anhydrase X	QN5S85	56934	CA10	RFLU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1
SLO10450	CD48	P09326	962	CD48	RFLU	No	No	CE	0	0	0	0	0	0	0	0	0.296	0.105	0.486	0.704
SLO10451	CFC1	P0CG37	55997	CFC1	RFLU	No	No	DE	0.342	0.221	0.390	0	0	0	0.342	0.125	0.560	0	0	0
SLO10454	Contactin-4	Q8IWV2	152330	CNTN4	RFLU	No	No	CE	0	0	0	0	0	0	0	0	0.544	0.396	0.693	0.456
SLO10455	Contactin-5	Q94779	53942	CNTN5	RFLU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1
SLO10456	CYTIN	P01037	1469	CS1	RFLU	No	No	CE	0	0	0	0	0	0	0	0	0.283	0.096	0.471	0.717
SLO10457	DLL4	Q9NR61	54567	DLL4	RFLU	No	No	DE	0.338	-0.031	0.405	0	0	0	0.338	-0.010	0.686	0	0	0
SLO10458	Endocan	Q8NG30	11082	ESM1	RFLU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1
SLO10461	FCGR1	P12314	2209	FCGR1A	RFLU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1
SLO10462	FCN1	Q00602	2219	FCN1	RFLU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1
SLO10463	GPC2	Q8N158	221914	GPC2	RFLU	No	No	CE	0	0	0	0	0	0	0	0	0.261	0.070	0.451	0.739
SLO10464	LRIG3	Q6UXM1	121227	LRIG3	RFLU	No	No	AE	0.514	0.522	0.510	0.514	0.330	0.698	0	0	0	0	0	0
SLO10465	MATN2	Q00339	4147	MATN2	RFLU	No	No	AE	0.470	0.452	0.479	0.470	0.278	0.663	0	0	0	0	0	0
SLO10466	MFRP	Q9BY79	83552	MFRP	RFLU	No	No	CE	0	0	0	0	0	0	0	0	0.255	0.064	0.446	0.530
SLO10467	RGMA	Q96886	56963	RGMA	RFLU	No	No	AE	0.440	0.396	0.458	0.440	0.227	0.653	0	0	0	0	0	0
SLO10468	RGMB	Q6NV40	285704	RGMB	RFLU	No	No	CE	0	0	0	0	0	0	0	0	0.275	0.076	0.474	0.560
SLO10469	RGMC	Q6ZVN8	148738	HEF2	RFLU	No	No	AE	0.455	0.424	0.468	0.455	0.249	0.662	0	0	0	0	0	0
SLO10470	Semaphorin 3E	O15041	9723	SEMA3E	RFLU	No	No	AE	0.630	0.683	0.601	0.630	0.485	0.775	0	0	0	0	0	0
SLO10481	Testican-2	Q02563	9806	SPOCK2	RFLU	No	No	AE	0.699	0.761	0.661	0.699	0.581	0.817	0	0	0	0	0	0
SLO10488	ABL2	P42684	27	ABL2	RFLU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1
SLO10489	CAMK1	Q14012	8536	CAMK1	RFLU	No	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
SLO10490	CAMK1D	Q8IU85	57118	CAMK1D	RFLU	No	No	CE	0	0	0	0	0	0	0	0	0.214	0.016	0.412	0.786
SLO10491	CAMK2A	Q9UQM7	815	CAMK2A	RFLU	No	No	AE	0.699	0.763	0.660	0.699	0.577	0.821	0	0	0	0	0	0
SLO10492	CAMK2B	Q13554	816	CAMK2B	RFLU	No	No	DE	0.368	0.242	0.411	0	0	0	0.368	0.124	0.612	0	0	0
SLO10493	CAMK2D	Q13557	817	CAMK2D	RFLU	No	No	AE	0.769	0.835	0.725	0.769	0.671	0.867	0	0	0	0	0	0
SLO10494	CDK1/cyclin B	P06493	P14983	891	CDK2 CCNB1	RFLU	No	No	DE	0.870	0.926	0.827	0	0	0	0.870	0.805	0.936	0	0
SLO10495	CDK2/cyclin A	P24941	P201017	890	CDK2 CC2	RFLU	No	No	DE	0.267	0.000	0.348	0	0	0	0.267	0.000	0.534	0	0
SLO10496	CDK5/p35	Q00535	Q11020	8851	CDK5 CDK5R1	RFLU	No	No	E	0	0	0	0	0	0	0	0	0	0	1
SLO10498	EPHA3	P29320	2042	EPHA3	RFLU	No	No	AE	0.404	0.667	0.455	0.604	0.414	0.794	0	0	0	0	0	0
SLO10499	HCK	P08631	3055	HCK	RFLU	No	No	CE	0	0	0	0	0	0	0	0	0.216	0.019	0.414	0.784
SLO10500	LYN	P07948	4067	LYN	RFLU	No	No	CE	0	0	0	0	0	0	0	0	0.515	0.366	0.665	0.485
SLO10501	MP2K2	P36507	5605	MAP2K2	RFLU	No	No	DE	0.786	0.850	0.742	0	0	0	0.786	0.695	0.878	0	0	0
SLO10502	MK08	P45983	5599	MAPK8	RFLU	No	No	DE	0.621	0.689	0.589	0	0	0	0.621	0.443	0.800	0	0	0
SLO10503	MAPK2	P49137	9261	MAPKAPK2	RFLU	No	No	CE	0	0	0	0	0	0	0	0	0.471	0.313	0.630	0.379
SLO10504	MAPK5	Q8IW41	8550	MAPKAPK5	RFLU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1
SLO10505	MATK	P42679	4145	MATK	RFLU	No	No	DE	0.283	0	-0.133	0.369	0	0	0	0.283	-0.045	0.612	0	0
SLO10508	PAK3	Q75914	5063	PAK3	RFLU	No	No	CE	0	0	0	0	0	0	0	0	0.168	-0.033	0.369	0.832
SLO10509	PAK5	Q8NDU5	56924	PAK5	RFLU	No	No	AE	0.760	0.824	0.717	0.760	0.661	0.859	0	0	0	0	0	0
SLO10510	PAK7	Q9P286	57344	PAK7	RFLU	No	No	AE	0.449	0.418	0.463	0.449	0.263	0.634	0	0	0	0	0	0
SLO10512	PIK3CA/PIK3R1	P42336	P275290	5295	PIK3CA PIK3R1	RFLU	No	No	E	0	0	0	0	0	0	0	0	0	0	1
SLO10513	PRKACA	P17612	5566	PRKACA	RFLU	No	No	CE	0	0	0	0	0	0	0	0	0.553	0.412	0.695	0.447
SLO10514	PTK6	P13882	5753	PTK6	RFLU	No	No	DE	0.381	0.241	0.422	0	0	0	0.381	0.111	0.650	0	0	0
SLO10515	RPS6KA3	P51812	6197	RPS6KA3	RFLU	No	No	DE	0.670	0.745	0.630	0	0	0	0.670	0.517	0.823	0	0	0
SLO10516	SRCN1	P12931	6714	SRC	RFLU	No	No	AE	0.718	0.782	0.677	0.718	0.604	0.832	0	0	0	0	0	0
SLO10517	STK16	Q75716	8576	STK16	RFLU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1
SLO10518	TEC	P42680	7006	TEC	RFLU	No	No	DE	0.546	0.573	0.533	0	0	0	0.546	0.356	0.735	0	0	0
SLO10519	ZAP70	P43403	7535	ZAP70	RFLU	No	No	CE	0	0	0	0	0	0	0	0	0.426	0.256	0.595	0.574
SLO10520	AURKB	Q96GD4	9212	AURKB	RFLU	No	No	DE	0.180	-0.023	0.383	0.180	-0.023	0.383	0	0	0.180	-0.023	0.383	0.180
SLO10521	BTX	Q06187	695	BTX	RFLU	No	No	ACE	0.394	-0.508	0.424	0.394	-0.032	0.820	0	0	0.349	-0.051	0.748	0.257
SLO10522	CDK8/cyclin C	P49336	P241024	892	CDK8 CCNC	RFLU	No	No	CE	0	0	0	0	0	0	0	0.396	0.220	0.571	0.604
SLO10523	HIPK3	Q9H422	10114	HIPK3	RFLU	No	No	CE	0	0	0	0	0	0	0	0	0.206	0.010	0.401	0.794
SLO10528	UFM1	P61960	51569	UFM1	RFLU	No	No	CE	0	0	0	0	0	0	0	0	0.445	0.281	0.609	0.555
SLO10529	UFC1	Q9Y3C8	51506	UFC1	RFLU	No	No	CE	0	0	0	0	0	0	0	0	0.490	0.334	0.645	0.510
SLO10530	OCAD1	Q9NX40	54940	OCAD1	RFLU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1
SLO10610	CLC4K	Q9UJ71	50489	CD207	RFLU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	1
SLO10612	Dkk-4	Q9UBT3	27121	DKK																

[illegible]

SLO16548	AMPK a1b1g1	P13131 Q9J566Z 5564	PRKAA1 PRKAB RFU	No	No	E	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
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